O·I·Analytical

Methodology

Total Kjeldahl Nitrogen by Gas Diffusion and Flow Injection Analysis (FIA)

(Cartridge Part #A002380)

1.0 Scope and Application

1.1 This method is used for the determination of Total Kjeldahl Nitrogen (TKN) in drinking water, surface water, and domestic and industrial wastes.

1.2 Nitrogen components of biological origin such as amino acids, proteins, and peptides are converted to ammonia via Kjeldahl digestion. It may not convert nitrogenous compounds of some industrial wastes, such as amines, nitro compounds, hydrazones, oximes, semicarbazones, and some refractory tertiary amines.

1.3 The Method Detection Limit (MDL) of this method is 0.02 mg/L TKN. The applicable range of the method is 0.20–10 mg/L TKN using a 200-µL sample loop. The range may be extended to analyze higher concentrations by sample dilution.

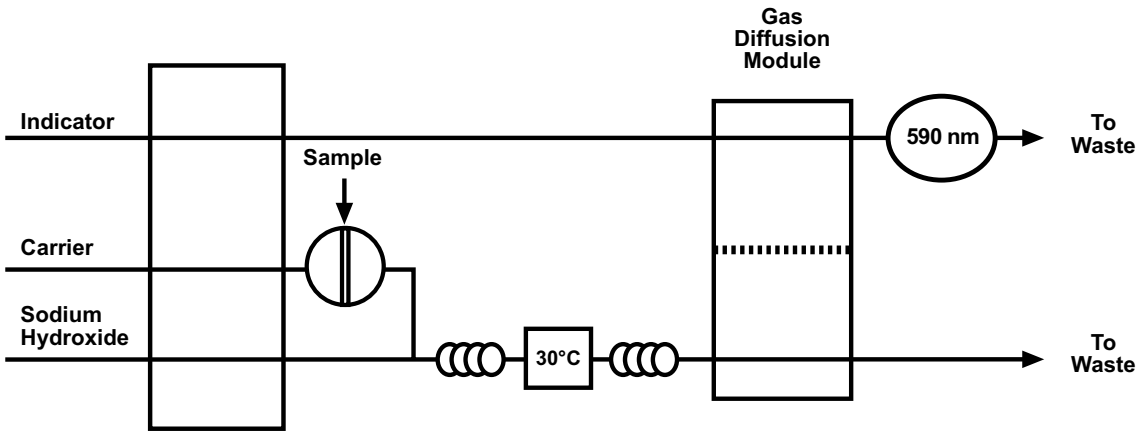
2.0 Summary of Method

2.1 Prior to analysis, the sample is digested in the presence of sulfuric acid, potassium sulfate, and a mercury catalyst at a final temperature of 380°C. Free ammonia and organic nitrogen compounds are converted to ammonium sulfate under these conditions.

2.2 The sample is neutralized and made alkaline with sodium hydroxide. The ammonia gas that is generated passes under a gas diffusion membrane and into a receiving solution that contains an indicator reagent. The absorbance is measured at 590 nm. The extent of the color change is proportional to the concentration of TKN present in the sample (References 15.1–15.3).

2.3 The quality of the analysis is assured through reproducible calibration and testing of the Flow Injection Analysis (FIA) system.

2.4 A general flow diagram of the FIA system is shown below (see Section 17.0 for a detailed flow diagram).



3.0 Definitions

Definitions for terms used in this method are provided in Section 16.0, "Glossary of Definitions and Purposes."

4.0 Interferences

4.1 High nitrate concentrations (greater than 10 times the TKN level) negatively interfere with the analysis.

4.2 Prevent the reaction between nitrate and ammonia by using an anion exchange resin (chloride form) to remove nitrate prior to the analysis.

5.0 Safety

5.1 The toxicity or carcinogenicity of each compound or reagent used in this method has not been fully established. Each chemical should be treated as a potential health hazard. Exposure to these chemicals should be reduced to the lowest possible level.

5.2 For reference purposes, a file of Material Safety Data Sheets (MSDS) for each chemical used in this method should be available to all personnel involved in this chemical analysis. The preparation of a formal safety plan is also advisable.

5.3 The following chemicals used in this method may be highly toxic or hazardous and should be handled with extreme caution at all times. Consult the appropriate MSDS before handling.

5.3.1 Ammonium Chloride, NH_4Cl (FW 53.49)

5.3.2 Hydrochloric Acid, concentrated, HCl (FW 36.46)

5.3.3 Potassium Sulfate, K_2SO_4 (FW 174.26)

5.3.4 Red Mercuric Oxide, HgO (FW 216.61)

5.3.5 Sodium Hydroxide, NaOH (FW 40.00)

5.3.6 Sulfuric Acid, concentrated, H_2SO_4 (FW 98.08)

5.3.7 TKN/Gas Diffusion Indicator (Part #A002510)

5.4 Unknown samples may be potentially hazardous and should be handled with extreme caution at all times.

5.5 Proper personal protective equipment (PPE) should be used when handling or working in the presence of chemicals.

5.6 This method does not address all safety issues associated with its use. The laboratory is responsible for maintaining a safe work environment and a current awareness file of OSHA regulations regarding the safe handling of the chemicals specified in this method.

6.0 Apparatus, Equipment, and Supplies

6.1 Flow Injection Analysis (FIA) System (OI Analytical Flow Solution® 3000) consisting of the following:

6.1.1 120-Place Autosampler

6.1.2 Expanded Range (ER) Photometric Detector with 5-mm path length flowcell and 590-nm optical filter

6.1.4 Data Acquisition System (PC or Notebook PC) with WinFLOW™ software

6.1.5 TKN by Gas Diffusion Cartridge (Part #A002380)

6.2 Sampling equipment—Sample bottle, amber glass, with polytetrafluoroethylene (PTFE)-lined cap. Clean by washing with detergent and water, rinsing with two aliquots of reagent water, and drying by baking at 110°–150°C for a minimum of one hour.

6.3 Standard laboratory equipment including volumetric flasks, pipettes, syringes, etc. should all be cleaned, rinsed, and dried per bottle cleaning procedure in Section 6.2.

7.0 Reagents and Calibrants

7.1 Raw Materials

7.1.1 Ammonium Chloride, NH_4Cl (FW 53.49)

7.1.2 Deionized Water (ASTM Type I or II)

7.1.3 Hydrochloric Acid, concentrated, HCl (FW 36.46)

7.1.4 Potassium Sulfate, K_2SO_4 (FW 174.26)

7.1.5 Red Mercuric Oxide, HgO (FW 216.61)

7.1.6 Sodium Hydroxide, NaOH (FW 40.00)

7.1.7 Sulfuric Acid, concentrated, H_2SO_4 (FW 98.08)

7.1.8 TKN/Gas Diffusion Indicator (Part #A002510)

7.2 Reagent Preparation

Note: For best results, filter and degas all reagents prior to use.

7.2.1 Reagent Water

7.2.1.1 Degassed and deionized reagent water can be prepared in one of the following manners:

7.2.1.1.1 Place distilled/deionized water under a strong vacuum for 15–20 minutes. Magnetic stirring or sonification will aid in the degassing process.

7.2.1.1.2 Purge distilled/deionized water with a stream of nitrogen gas (or other inert gas) through a glass frit for approximately 5 minutes.

7.2.1.1.3 Boil distilled/deionized water in an Erlenmeyer flask for 15–20 minutes. Remove the flask from the heat source, cover it with an inverted beaker, and allow it to cool to room temperature.

7.2.1.2 After preparing the degassed reagent water, store the reagent water in a tightly sealed container to protect it from reabsorption of atmospheric gases. For best results, store degassed reagent water under a slight vacuum when not in use.

7.2.2 Digestion Reagents

7.2.2.1 1:4 Sulfuric Acid Solution (50 mL)

7.2.2.1.1 While stirring, carefully add 10 mL of concentrated sulfuric acid to 40 mL of reagent water (Section 7.2.1).

Warning: Mixing sulfuric acid with water releases a great amount of heat. Take appropriate precautions.

7.2.2.2 Mercuric Sulfate Solution (100 mL)

7.2.2.2.1 Dissolve 8 g of red mercuric oxide in 50 mL of 1:4 sulfuric acid solution (Section 7.2.2.1) in a 100-mL volumetric flask.

7.2.2.2.2 Dilute to 100 mL with reagent water and mix well.

Warning: Mercury compounds are highly toxic. Take appropriate precautions.

7.2.2.3 Digestion Solution (2 L)

7.2.2.3.1 While stirring, carefully add 400 mL of concentrated sulfuric acid to approximately 1,300 mL of reagent water in a 2-L volumetric flask.

7.2.2.3.2 Add 8 g of red mercuric oxide and stir until dissolved.

7.2.2.3.3 Add 25 mL of mercuric sulfate solution (Section 7.2.2.2). Allow the solution to cool to room temperature.

7.2.2.3.4 Dilute to 2,000 mL with reagent water and mix well.

Warning: Mixing sulfuric acid with water releases a great amount of heat. Take appropriate precautions.

Warning: Mercury compounds are highly toxic. Take appropriate precautions.

7.2.3 Color Reagents

7.2.3.1 Start-up Solution and Carrier, 5 M Sodium Hydroxide (1 L)

7.2.3.1.1 While stirring, carefully add 200 g of sodium hydroxide to approximately 700 mL of reagent water in a 1-L volumetric flask.

7.2.3.1.2 Allow the solution to cool to room temperature. Dilute to 1,000 mL with reagent water and mix well.

Warning: Mixing sodium hydroxide with water releases a great amount of heat. Take appropriate precautions.

Note: Store in a polyethylene container.

7.2.3.2 0.1 M Sodium Hydroxide Solution (100 mL)

7.2.3.2.1 Use a volumetric pipet to add 5 mL of start-up solution (Section 7.2.3.1) to approximately 80 mL of reagent water in a 100-mL volumetric flask.

7.2.3.2.2 Dilute to 100 mL with reagent water and mix well.

7.2.3.3 Stock Indicator Solution (200 mL)

7.2.3.3.1 Dissolve 1 g of TKN/gas diffusion indicator in 5 mL of 0.1 M sodium hydroxide solution (Section 7.2.3.2) in a 200-mL volumetric flask.

7.2.3.3.2 Dilute to 200 mL with reagent water and mix well.

Note: This solution is stable for six months.

7.2.3.4 Working Indicator Solution (500 mL)

7.2.3.4.1 Use a volumetric pipet to add 10 mL of stock indicator solution (Section 7.2.3.3) to approximately 400 mL of reagent water in a 500-mL volumetric flask.

7.2.3.4.2 Dilute to 500 mL with reagent water and mix well.

Note: This solution is stable for 4–6 weeks.

Note: Prepare this solution at least 24 hours prior to use and adjust to the proper absorbance value daily. See Section 11.2.1 for complete instructions.

7.2.3.5 0.01 M Sodium Hydroxide Solution (500 mL)

7.2.3.5.1 Use a volumetric pipet to add 1 mL of start-up solution (Section 7.2.3.1) to approximately 400 mL of reagent water in a 500-mL volumetric flask.

7.2.3.5.2 Dilute to 500 mL with reagent water and mix well.

7.2.3.6 0.01 M Hydrochloric Acid Solution (500 mL)

7.2.3.6.1 While stirring, carefully add 0.42 mL of concentrated hydrochloric acid to approximately 400 mL of reagent water in a 500-mL volumetric flask.

7.2.3.6.2 Dilute to 500 mL with reagent water and mix well.

7.3 Calibrant Preparation

7.3.1 Stock Calibrant 1,000 mg/L Ammonia (1 L)

7.3.1.1 Dissolve 3.819 g of ammonium chloride in approximately 900 mL of reagent water in a 1-L volumetric flask.

7.3.1.2 Dilute to 1,000 mL with reagent water and mix well.

Note: Store at 4°C in a tightly sealed container. If stored properly, this solution is stable for 4–6 weeks.

7.3.2 Intermediate Calibrant 100 mg/L Ammonia (100 mL)

7.3.2.1 Use a volumetric pipet to add 10 mL of stock calibrant (Section 7.3.1) to approximately 80 mL of reagent water in a 100-mL volumetric flask.

7.3.2.2 Dilute to 100 mL with reagent water and mix well.

Note: Prepare this solution fresh daily.

7.3.3 Working Calibrants (100 mL)

7.3.3.1 Add the designated volumes of stock calibrant (see Equation 1) to the required number of 100-mL volumetric flasks that each contain approximately 80 mL of reagent water.

7.3.3.2 Dilute each solution to the mark with reagent water and mix well.

Note: Prepare working calibrants fresh daily.

EQUATION 1

$$C_1V_1 = C_2V_2$$

Where:
C_1 = *Concentration (in mg/L) of stock solution (or calibrant)*
V_1 = *Volume (in L) of stock solution (or calibrant) to be used*
C_2 = *Desired concentration (in mg/L) of working calibrant to be prepared*
V_2 = *Final volume (in L) of working calibrant to be prepared*

By solving this equation for the volume of stock solution to be used (V_1), the following equation is obtained:

$$V_1 = \frac{C_2V_2}{C_1}$$

Since the desired concentration (C_2), the final volume (V_2), and the concentration of the stock solution (C_1) are all known for any given calibrant concentration in a defined volume, the volume of stock solution to be used (V_1) is easily calculated.

7.3.3.3 Calibrants covering the entire range of this analysis using a 200-µL injection loop can be prepared from the following tables.

Final Concentration (mg/L)	Vol. of Inter. Cal. (mL)	Conc. of Inter. Cal. (mg/L)	Final Volume (mL)
0.20	0.20	100	100
0.40	0.40	100	100
0.70	0.70	100	100
1.0	1.0	100	100

Final Concentration (mg/L)	Vol. of Stock Cal. (mL)	Conc. of Stock Cal. (mg/L)	Final Volume (mL)
2.0	0.20	1,000	100
4.0	0.40	1,000	100
6.0	0.60	1,000	100
8.0	0.80	1,000	100
10	1.0	1,000	100

8.0 Sample Collection, Preservation, and Storage

8.1 Samples should be collected in plastic or glass bottles that have been thoroughly cleaned and rinsed with reagent water (Section 7.2.1).

8.2 The volume of sample collected should be sufficient to ensure that a representative sample is obtained, replicate analysis is possible, and waste disposal is minimized.

8.3 Determine TKN in unpreserved samples within 24 hours of collection.

8.4 Preserve and store samples by adding 2 mL of concentrated sulfuric acid per liter of sample and refrigerating at 4°C. Sample analysis should be performed as soon as possible to eliminate loss of analyte.

8.5 Holding time for preserved samples is 28 days from the time of collection (Reference 15.5).

9.0 Quality Control

Note: The following QC procedures are provided for reference purposes only and are not a substitute for any QC procedures that may be required for regulatory compliance.

9.1 It is recommended that each laboratory that uses this method operate a formal quality control program. The minimum requirements of such a program should consist of an initial demonstration of laboratory capability and the periodic analysis of Laboratory Control Samples (LCSs) and Matrix Spike/Matrix Spike Duplicates (MS/MSDs) as a continuing check on performance. Laboratory performance should be compared to established performance criteria to determine if the results of the analyses meet the performance characteristics of the method.

9.2 Method Detection Limit (MDL)—To establish the ability to detect TKN at low levels, the analyst should determine the MDL using the apparatus, reagents, and calibrants that will be used in the practice of this method. An MDL less than or equal to the MDL listed in Section 1.3 should be achieved prior to practice of this method.

9.2.1 An MDL is calculated by analyzing a matrix spike at a concentration of two to three times the expected detection limit of the analyzer. Seven consecutive replicate analyses of this matrix spike should be analyzed, and the MDL should be calculated using Equation 2.

EQUATION 2

$$MDL = (t) \times (S)$$

Where:

t = Student's t value for a 99% confidence level and a standard deviation estimate with n–1 degrees of freedom (t = 3.14 for seven replicates)

S = Standard deviation of the replicate analyses

9.2.2 It is recommended that the MDL be calculated after every six months of operation, when a new operator begins work, or whenever there is any significant change in the instrument response.

9.3 Analyses of MS/MSD samples are required to demonstrate method accuracy and precision and to monitor matrix interferences (interferences caused by the sample matrix).

9.3.1 Matrix Spike/Matrix Spike Duplicate (MS/MSD)—The laboratory should spike, in duplicate, a minimum of 10% of all samples (one sample in duplicate in each batch of 10 samples) from a given sampling site.

9.3.2 The concentration of the spike in the sample shall be determined as follows:

9.3.2.1 If, as in compliance monitoring, the concentration of TKN in the sample is being checked against a regulatory concentration limit, the spiking level shall be at that limit.

9.3.2.2 If the concentration of TKN in a sample is not being checked against a limit, the spike shall be at the concentration of the LCS or at least four times greater than the MDL.

9.4 Analyses of Laboratory Reagent Blanks (LRBs) are required to demonstrate freedom from contamination and that the compounds of interest and interfering compounds have not been carried over from a previous analysis.

9.5 As part of the QC program for the laboratory, method precision and accuracy for samples should be assessed and records should be maintained.

9.5.1 An LCS should be analyzed with every sample batch, and the mean (m) and the standard deviation (S) should be recorded. After multiple analyses, the mean should be plotted with limits of $m+2S$ and $m–2S$. The mean and the limits should be recalculated after every 5–10 new measurements.

9.5.2 If the LCS measurement falls outside the range calculated in Section 9.5.1, then the problem should be addressed, and that sample batch should be reanalyzed if necessary.

9.6 Reference Sample—To demonstrate that the analytical system is in control, the laboratory may wish to periodically test an external reference sample, such as a Standard Reference Material (SRM) available from the National Institute of Standards and Technology (NIST). Corrective action should be taken if the measured concentration significantly differs from the stated concentration.

10.0 Configuration and Start-up

10.1 Instrument Configuration

10.1.1 Configure the OI Analytical Flow Solution 3000 Analyzer according to the Operator's Manual and verify that each module is properly powered on.

10.1.2 Verify that the TKN by Gas Diffusion Cartridge (Part #A002380) is configured as illustrated in the flow diagram shown in Section 17.0.

10.1.3 Connect the appropriate pump tubes to the cartridge and to their appropriate reagent containers according to the flow diagram.

10.2 Instrument Stabilization

10.2.1 Connect the reagent pump tubes to a reagent bottle containing the start-up solution (Section 7.2.3.1). Start the pump at low speed, allowing the start-up solution to flow through the entire system.

10.2.2 Verify that the flowcell of each detector is purged of all bubbles and that the flow is stable and free from surging before proceeding.

10.3 Baseline Verification

10.3.1 Create and save a Method in WinFLOW. Refer to the WinFLOW Operator's Manual (Reference 15.6) for help on creating a Method.

10.3.2 Create and save a Sample Table in WinFLOW that will be used to generate a calibration curve using at least three calibrants that cover the full range of expected concentrations in the samples to be analyzed. This Sample Table should also be used to analyze all necessary QC samples as well as the analytical batch of samples to be analyzed. For help on creating a Sample Table, refer to the WinFLOW Operator's Manual (Reference 15.6).

10.3.3 Select **Collect Data** in the WinFLOW main window, enter the user's identification, select the appropriate Method and Sample Table, and begin to collect baseline data. Very sharp fluctuations in the baseline and/or consistent drifting are typically signs of bubbles in the flowcell. The flowcell must be free of bubbles prior to beginning analysis.

10.4 Calibration and Standardization

10.4.1 Prepare a series of at least three working calibrants using the stock solutions (Section 7.3) according to Equation 1, covering the desired analysis range.

10.4.2 Place the calibrants in the autosampler in order of increasing concentration. Each calibrant should be analyzed according to the analytical procedures in Section 11.0. A calibration curve will be calculated by the WinFLOW software.

10.4.3 Acceptance or control limits for the calibration results should be established using the difference between the measured value of each calibrant and the corresponding "true" concentration.

10.4.4 Each calibration curve should be verified by analysis of a Laboratory Control Sample (LCS, Section 9.5). Using WinFLOW software, calibration, verification, and sample analysis may be performed in one continuous analysis.

11.0 Procedure

11.1 Digestion

11.1.1 Blanks, Calibrants, Samples, and QC Samples

Note: The final diluted volume after digestion will be 25 mL.

11.1.1.1 For blanks, add 25 mL of reagent water to the tube.

11.1.1.2 For calibrants, use a volumetric pipet to add the appropriate amount of stock or intermediate calibrant (Section 7.3) to each tube. Completely rinse the material down the side of the tube with reagent water to an approximate volume of 25 mL. Mix well with a vortex mixer.

11.1.1.3 For samples, use a volumetric pipet to add 25 mL to the tube and mix well with a vortex mixer. If samples are known to be out of range, smaller volumes can be used and the result multiplied by the appropriate dilution factor.

11.1.1.3.1 For example, if 10 mL of sample is digested, multiply the result by 2.5 after digestion and analysis. If 5 mL of sample is digested, multiply the result by 5 after digestion and analysis.

11.1.1.4 For QC samples, prepare the samples according to its instructions and add 25 mL to the tube. Mix well with a vortex mixer.

11.1.2 Use a volumetric pipet to add 5 mL of digestion solution (Section 7.2.2.3) to each tube and mix well with a vortex mixer.

Note: Measure the digestion solution accurately to ensure uniform acid content in each tube.

11.1.3 Add 4–6 Teflon® or glass boiling stones to each tube.

Note: Too many boiling stones will cause the sample to boil over.

11.1.4 Place the tubes in the block digester and set the temperature to 160°C. Allow the volume of each tube to be reduced to about 5 mL (approximately one hour).

Note: The evaporation step reduces the volume of liquid in the tube before the temperature is increased for digestion.

11.1.5 Increase the temperature to 380°C for 2½ hours. When the digestion is complete, the final volume will be about 2–3 mL. The liquid will be clear or straw-colored.

Note: If any amount of liquid boils out of the tube during the digestion, the result will be in error.

Note: If an exhaust system or scrubber is placed on top of the tubes during the digestion, care must be taken not to draw significant amount of fumes during the high temperature stage. Variable acid concentrations may result. An exhaust system is not necessary if the digestion apparatus is placed in a properly operating fume hood.

11.1.6 Remove the tubes from the block to cool. Do not allow the digests to crystallize while cooling. To prevent crystallization, add a few milliliters of reagent water to the digests after cooling the tubes for several minutes.

Caution: If the reagent water is added too soon and the acid is still hot, the contents of the tube may react vigorously and some material may be lost from the tube.

11.1.7 Dilute the digestate to 25 mL with reagent water. The digests must be accurately diluted to a known volume in a calibrated container in order to ensure consistent acid content in the final solutions. If the contents of the tubes are removed to calibrated containers during this step, ensure that all of the material is transferred.

11.2 Analysis

11.2.1 Adjust the working indicator solution (Section 7.2.3.4) so that its absorbance, with reagent water as a reference, is between 0.25–0.30 absorbance units.

11.2.1.1 Place the 590-nm filter in the detector.

11.2.1.2 Pump reagent water through all the lines for 5–10 minutes.

11.2.1.3 Exchange the water in the appropriate line with working indicator solution (see the flow diagram in Section 17.0).

11.2.1.4 Turn the heater on. Once the heater unit has reached 30°C, verify a stable baseline (Section 10.3). Verify there are no bubbles in the flowcell. Obtain a stable baseline at 590 nm and autozero the baseline before proceeding.

11.2.1.5 The absorbance should fall within the range of 0.25–0.30 units. If the absorbance value is low, adjust the working indicator solution by dropwise addition of 0.01 M sodium hydroxide (Section 7.2.3.5). If the absorbance is high, adjust the solution by dropwise addition of 0.01 M hydrochloric acid (Section 7.2.3.6).

11.2.1.6 Note the absorbance reading obtained with the adjusted working indicator solution. Exchange the reagent water in the remaining lines with start-up solution (Section 7.2.3.1). Wait for a stable absorbance reading.

Note: The increase in absorbance should not be greater than 0.02–0.04 absorbance units. If the increase is greater or if there is continuous drift, the gas diffusion membrane may be damaged.

11.2.1.7 Following the stable reading, inject the highest standard three or more times, or until a stable response is again achieved.

11.2.2 Load the sampler tray with calibrants, blanks, samples, and QC samples.

Note: The matrix of the working standards, blanks, and QC samples should match that of the samples being analyzed.

11.2.3 Using the Method and Sample Table created for the analytical batch to be analyzed and with the baseline verified to be stable, begin the analysis by selecting the “Fast Forward” button on the left side of the Data Analysis window in WinFLOW. This will initiate the sequential analysis of samples as defined in the Sample Table.

11.2.4 When analysis is complete, pump start-up solution through the system for at least 10–15 minutes. Stop the pump, release the tension on all pump tubes, and power off the system.

11.3 Operating Notes

11.3.1 Degas all reagents prior to use. Avoid using a vacuum filtration unit to prevent introduction of ammonia or carbon dioxide. Carbon dioxide decreases the sensitivity of the working indicator solution.

11.3.2 For all ranges, use a second or third order curve fit.

11.3.3 For determining TKN at low levels, take the following precautions:

11.3.3.1 Clean all glassware with acid and base prior to use.

11.3.3.2 Use polyethylene containers instead of glass.

11.3.3.3 Store all solutions in tightly stoppered containers to prevent contamination from gaseous ammonia present in ambient air.

12.0 Data Analysis and Calculations

12.1 The calibration curve allows for accurate quantitation of the concentration in each sample.

12.2 WinFLOW software reports the concentration of each sample relative to the calibration curve.

13.0 Method Performance

Range:	0.20–10 mg/L TKN
Throughput:	40 samples/hour
Precision:	
0.50 mg/L	<3% RSD
5.0 mg/L	<2% RSD
Method Detection Limit (MDL):	0.02 mg/L TKN

14.0 Pollution Prevention and Waste Management

14.1 It is the laboratory's responsibility to comply with all federal, state, and local regulations governing waste management, particularly the hazardous waste identification rules and land-disposal restrictions. In addition, it is the laboratory's responsibility to protect air, water, and land resources by minimizing and controlling all releases from fume hoods and bench operations. Also, compliance is required with any sewage discharge permits and regulations.

14.2 For further information on waste management, consult Section 13.6 of *Less is Better: Laboratory Chemical Management for Waste Reduction* (Reference 15.4).

15.0 References

15.1 Ramsing, A.; Ruzicka, J.; Hansen, E.H. *Acta Chem. Scand.* **1980**, *114*, 165.

15.2 Svensson, G.; Anfalt, T. *Acta Chem. Scand.* **1982**, *119*, 7.

15.3 Ruzicka, J.; Hansen, E.H. *Flow Injection Analysis*; Wiley & Sons: New York, 1981.

15.4 *Less is Better: Laboratory Chemical Management for Waste Reduction.* Available from the American Chemical Society, Department of Government Regulations and Science Policy, 1155 16th Street, NW, Washington, DC, 20036.

15.5 Sample Preservation. *Methods for Chemical Analysis of Water and Wastes*; EPA-600/4-79-020; U.S. Environmental Protection Agency, Office of Research and Development, Environmental Monitoring and Support Laboratory: Cincinnati, OH, 1984; xvii.

15.6 WinFLOW Software and Operator's Manual (Part #A002877). Available from OI Analytical, P.O. Box 9010, College Station, TX, 77842-9010.

16.0 Glossary of Definitions and Purposes

The definitions and purposes are specific to this method but have been conformed to common usage as much as possible.

16.1 Units of weights and measures and their abbreviations

16.1.1 Symbols

°C	degrees Celsius
%	percent
±	plus or minus
≥	greater than or equal to
≤	less than or equal to

16.1.2 Alphabetical characters

g	gram
L	liter
mg	milligram
mg/L	milligram per liter
µg	microgram
µg/L	microgram per liter
mL	milliliter
ppm	parts per million
ppb	parts per billion
M	molar solution
N	normal solution

16.2 Definitions

16.2.1 Laboratory Control Sample (LCS)—An aliquot of LRB to which a quantity of the analyte of interest is added in the laboratory. The LCS is analyzed like a sample. Its purpose is to determine whether the methodology is in control and whether the laboratory is capable of making accurate and precise measurements.

16.2.2 Laboratory Reagent Blank (LRB)—An aliquot of reagent water and other blank matrix that is treated like a sample, including exposure to all glassware, equipment, and reagents that are used with other samples. The LRB is used to determine if the method analyte or other interferences are present in the laboratory environment, reagents, or apparatus.

16.2.3 Matrix Spike/Matrix Spike Duplicate (MS/MSD)—An aliquot of an environmental sample to which a quantity of the method analyte is added in the laboratory. The MS/MSD is analyzed like a sample. Its purpose is to determine whether the sample matrix contributes bias to the analytical results. The background concentration of the analyte in the sample matrix must be determined in a separate aliquot, and the measured values in the MS/MSD must be corrected for the background concentration.

16.2.4 Method Detection Limit (MDL)—The minimum concentration of a substance that can be measured and reported with 99% confidence that the analyte concentration is greater than zero.

17.0 Figures

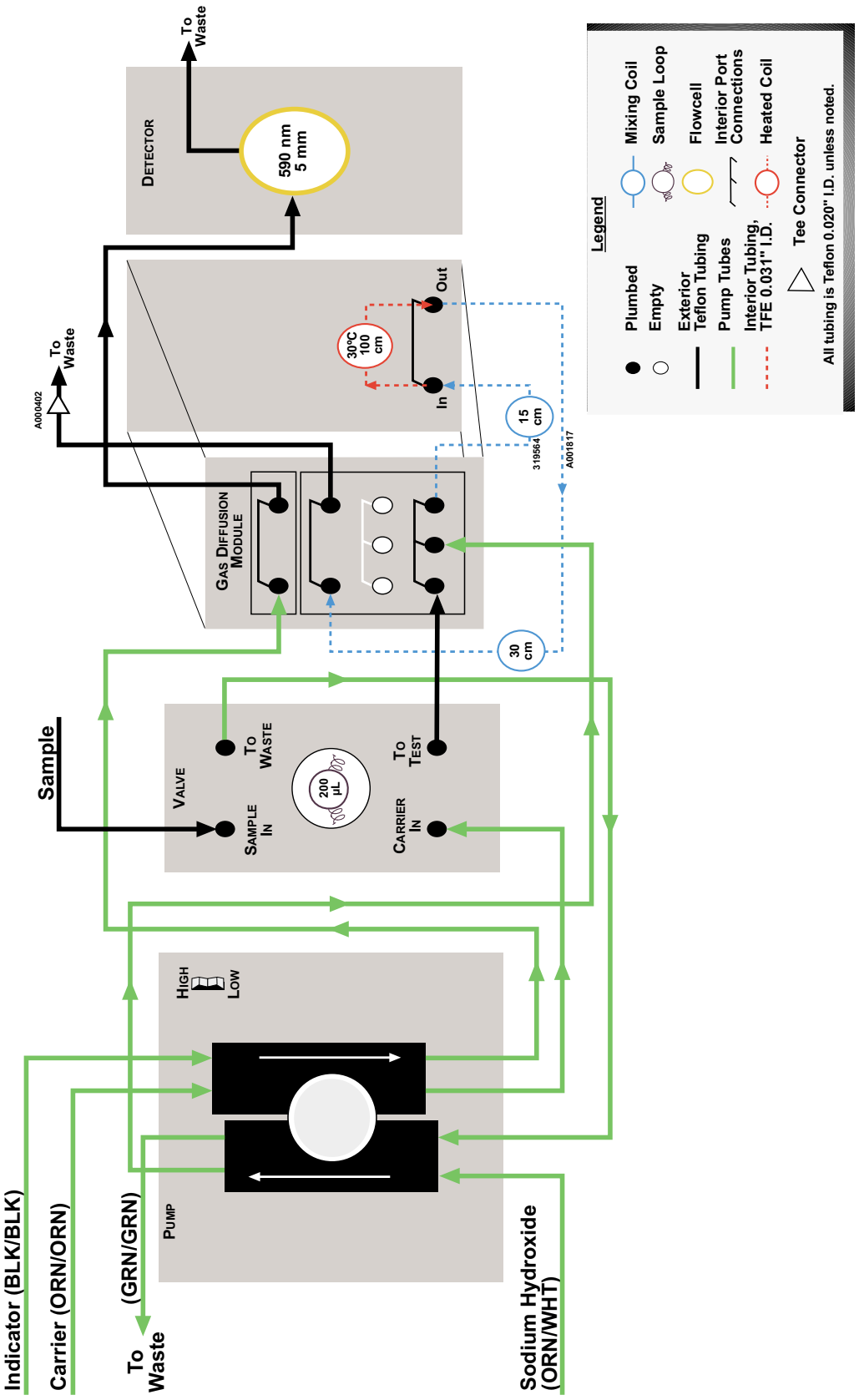


Figure 1. Detailed Flow Diagram for TKN by Gas Diffusion and FIA on a Flow Solution 3000, Cartridge Part #A002380

Results were obtained under optimal operating conditions. Actual results may vary depending on sample introduction, cleanliness of sample containers, reagent purity, operator skill, and maintenance of instruments.

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