O·I·Analytical

Methodology

Sulfide
by Flow Injection Analysis (FIA)

(Cartridge Part #A002762)

1.0 Scope and Application

1.1 This method is used for the determination of sulfide in drinking water, surface water, saline water, and domestic and industrial wastes.

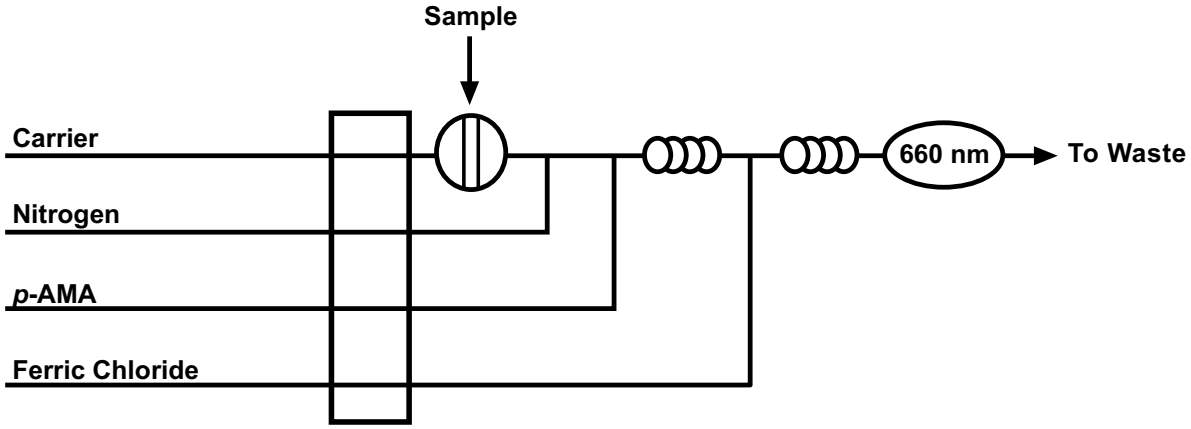
1.2 The Method Detection Limit (MDL) of this method is 0.002 mg/L sulfide. The applicable range of the method is 0.005–20.0 mg/L sulfide. A range of 0.005–1.00 mg/L sulfide is achieved using a 200-µL sample loop. Use a 100-µL sample loop to attain a range of 0.100–20.0 mg/L sulfide. The range may be extended to analyze higher concentrations by sample dilution.

2.0 Summary of Method

2.1 Sulfide reacts with *p*-aminodimethylaniline (*p*-AMA) and ferric chloride to form methylene blue. The absorbance is measured at 660 nm (Reference 15.2). This method does not detect acid insoluble sulfides.

2.2 The quality of the analysis is assured through reproducible calibration and testing of the Flow Injection Analysis (FIA) system.

2.3 A general flow diagram of the FIA system is shown below (see Section 17.0 for a detailed flow diagram).



3.0 Definitions

Definitions for terms used in this method are provided in Section 16.0, "Glossary of Definitions and Purposes."

4.0 Interferences

4.1 Strong reducing agents such as thiosulfate at concentrations above 10 mg/L inhibit color formation.

4.2 Samples with background absorbance at the analytical wavelength may interfere.

4.3 Filter or centrifuge turbid samples prior to determination.

4.4 Consult Reference 15.4 for treatment procedures for the removal of major interferences.

5.0 Safety

5.1 The toxicity or carcinogenicity of each compound or reagent used in this method has not been fully established. Each chemical should be treated as a potential health hazard. Exposure to these chemicals should be reduced to the lowest possible level.

5.2 For reference purposes, a file of Material Safety Data Sheets (MSDS) for each chemical used in this method should be available to all personnel involved in this chemical analysis. The preparation of a formal safety plan is also advisable.

5.3 The following chemicals used in this method may be highly toxic or hazardous and should be handled with extreme caution at all times. Consult the appropriate MSDS before handling.

5.3.1 *p*-Aminodimethylaniline (*N*,*N*-Dimethyl-1,3-Phenylenediamine Dihydrochloride), $C_8H_{12}N_2 \cdot 2HCl$ (FW 209.12)

5.3.2 Cadmium Sulfate Tri(octahydrate), $3CdSO_4 \cdot 8H_2O$ (FW 769.51)

5.3.3 Ferric Chloride Hexahydrate, $FeCl_3 \cdot 6H_2O$ (FW 270.30)

5.3.4 Hydrochloric Acid, concentrated, HCl (FW 36.46)

5.3.5 Potassium Biiodate, $KH(IO_3)_2$ (FW 389.92)

5.3.6 Sodium Carboxymethylcellulose, $R_n\text{-}OCH_2COONa$

5.3.7 Sodium Hydroxide, NaOH (FW 40.00)

5.3.8 Sodium Sulfide Nonahydrate, $Na_2S \cdot 9H_2O$ (FW 240.18)

5.3.9 Sodium Thiosulfate Pentahydrate, $Na_2S_2O_3 \cdot 5H_2O$ (FW 158.11)

5.3.10 Starch Indicator, 0.5% solution

5.3.11 Sulfuric Acid, concentrated, H_2SO_4 (FW 98.08)

5.3.12 Zinc Acetate Dihydrate, $Zn(CH_3CO_2)_2 \cdot 2H_2O$ (FW 219.50)

5.4 Unknown samples may be potentially hazardous and should be handled with extreme caution at all times.

5.5 Proper personal protective equipment (PPE) should be used when handling or working in the presence of chemicals.

5.6 This method does not address all safety issues associated with its use. The laboratory is responsible for maintaining a safe work environment and a current awareness file of OSHA regulations regarding the safe handling of the chemicals specified in this method.

6.0 Apparatus, Equipment, and Supplies

6.1 Flow Injection Analysis (FIA) System (OI Analytical Flow Solution® 3000) consisting of the following:

6.1.1 120-Place Autosampler

6.1.3 Expanded Range (ER) Photometric Detector with 5-mm path length flowcell and 660-nm optical filter

6.1.4 Data Acquisition System (PC or Notebook PC) with WinFLOW™ software

6.1.5 Sulfide Cartridge (Part #A002762)

6.2 Sampling equipment—Sample bottle, amber glass, with polytetrafluoroethylene (PTFE)-lined cap. Clean by washing with detergent and water, rinsing with two aliquots of reagent water, and drying by baking at 110°–150°C for a minimum of one hour.

6.3 Standard laboratory equipment including volumetric flasks, pipettes, syringes, etc. should all be cleaned, rinsed, and dried per bottle cleaning procedure in Section 6.2.

7.0 Reagents and Calibrants

7.1 Raw Materials

7.1.1 *p*-Aminodimethylaniline (*N*,*N*-Dimethyl-1,3-Phenylenediamine Dihydrochloride), $C_8H_{12}N_2 \cdot 2HCl$ (FW 209.12)

7.1.2 Brij®-35, 30% w/v (Part #A21-0110-33)

7.1.3 Cadmium Sulfate Tri(octahydrate), $3CdSO_4 \cdot 8H_2O$ (FW 769.51)

7.1.4 Deionized Water (ASTM Type I or II)

7.1.5 Ferric Chloride Hexahydrate, $FeCl_3 \cdot 6H_2O$ (FW 270.30)

7.1.6 Hydrochloric Acid, concentrated, HCl (FW 36.46)

7.1.7 Potassium Biiodate, $KH(IO_3)_2$ (FW 389.92)

7.1.8 Sodium Carboxymethylcellulose, $R_n\text{-}OCH_2COONa$

7.1.9 Sodium Hydroxide, NaOH (FW 40.00)

7.1.10 Sodium Sulfide Nonahydrate, $Na_2S \cdot 9H_2O$ (FW 240.18)

7.1.11 Sodium Thiosulfate Pentahydrate, $Na_2S_2O_3 \cdot 5H_2O$ (FW 158.11)

7.1.12 Starch Indicator, 0.5% solution

7.1.13 Sulfuric Acid, concentrated, H_2SO_4 (FW 98.08)

7.1.14 Zinc Acetate Dihydrate, $Zn(CH_3CO_2)_2 \cdot 2H_2O$ (FW 219.50)

7.2 Reagent Preparation

Note: For best results, filter and degas all reagents prior to use.

7.2.1 Reagent Water

7.2.1.1 Degassed and deionized reagent water can be prepared in one of the following manners:

7.2.1.1.1 Place distilled/deionized water under a strong vacuum for 15–20 minutes. Magnetic stirring or sonification will aid in the degassing process.

7.2.1.1.2 Purge distilled/deionized water with a stream of nitrogen gas (or other inert gas) through a glass frit for approximately 5 minutes.

7.2.1.1.3 Boil distilled/deionized water in an Erlenmeyer flask for 15–20 minutes. Remove the flask from the heat source, cover it with an inverted beaker, and allow it to cool to room temperature.

7.2.1.2 After preparing the degassed reagent water, store the reagent water in a tightly sealed container to protect it from reabsorption of atmospheric gases. For best results, store degassed reagent water under a slight vacuum when not in use.

7.2.2 Start-up Solution/Carrier (1 L)

7.2.2.1 Add 2 mL of Brij-35 to approximately 800 mL of reagent water (Section 7.2.1) in a 1-L volumetric flask.

7.2.2.2 Dilute to 1,000 mL with reagent water and mix gently.

7.2.3 *p*-Aminodimethylaniline Solution (*p*-AMA) (1 L)

7.2.3.1 While stirring, carefully add 50 mL of concentrated hydrochloric acid to approximately 800 mL of reagent water in a 1-L volumetric flask.

7.2.3.2 Add 0.38 g of *p*-aminodimethylaniline.

7.2.3.3 Add 0.5 mL of Brij-35.

7.2.3.4 Dilute to 1,000 mL with reagent water and mix well.

Warning: Mixing hydrochloric acid with water releases a great amount of heat. Take appropriate precautions.

Note: Store in an amber bottle. Prepare this solution fresh daily.

7.2.4 Ferric Chloride Solution (1 L)

7.2.4.1 While stirring, carefully add 50 mL of concentrated hydrochloric acid to approximately 800 mL of reagent water in a 1-L volumetric flask.

7.2.4.2 Add 5.13 g of ferric chloride hexahydrate.

7.2.4.3 Add 0.5 mL of Brij-35.

7.2.4.4 Dilute to 1,000 mL with reagent water and mix well.

Warning: Mixing hydrochloric acid with water releases a great amount of heat. Take appropriate precautions.

Note: Store in an amber bottle. Prepare this solution fresh daily.

7.2.5 1 N Sodium Hydroxide (500 mL)

7.2.5.1 Dissolve 20 g of sodium hydroxide in approximately 400 mL of reagent water in a 500-mL volumetric flask.

7.2.5.2 Dilute to 500 mL with reagent water and mix well.

Warning: Mixing sodium hydroxide with water releases a great amount of heat. Take appropriate precautions.

7.2.6 Absorbing Solution (1 L)

7.2.6.1 While stirring, slowly add 1 g of sodium carboxymethylcellulose to approximately 800 mL of reagent water in a 1-L beaker.

7.2.6.2 Heat the solution to 50°C to completely dissolve the sodium carboxymethylcellulose.

7.2.6.3 Dissolve 0.5 g of cadmium sulfate tri(octahydrate) in 50 mL of reagent water in a separate beaker. Add this solution to the sodium carboxymethylcellulose solution (Section 7.2.6.2).

7.2.6.4 Add 0.8 mL of 1 N sodium hydroxide (Section 7.2.5).

7.2.6.5 Quantitatively transfer the solution to a 1-L volumetric flask and dilute to 1,000 mL with reagent water. Mix well.

Note: This solution is normally turbid. Do not filter.

7.2.7 2 N Zinc Acetate (100 mL)

7.2.7.1 Dissolve 21.9 g of zinc acetate dihydrate in approximately 60 mL of reagent water in a 100-mL volumetric flask.

7.2.7.2 Dilute to 100 mL with reagent water and mix well.

7.2.8 Thiosulfate Solution, 0.025 N (1 L)

7.2.8.1 Dissolve 6.205 g of sodium thiosulfate pentahydrate in approximately 800 mL of reagent water in a 1-L volumetric flask.

7.2.8.2 Dilute to 1,000 mL with reagent water and mix well.

7.2.9 Biiodate Stock Solution, 0.1 N (1 L)

7.2.9.1 Dissolve 3.249 g of potassium biiodate in approximately 800 mL of reagent water in a 1-L volumetric flask.

7.2.9.2 Dilute to 1,000 mL with reagent water and mix well.

7.2.10 Working Biiodate Solution, 0.025 N (1 L)

7.2.10.1 Dilute 250 mL of biiodate stock solution (Section 7.2.9) to 1,000 mL with reagent water in a 1-L volumetric flask. Mix well.

7.2.11 Standardized Thiosulfate Solution (1 L)

7.2.11.1 Use a volumetric pipet to add 20 mL of working biiodate solution (Section 7.2.10) to a 250-mL Erlenmeyer flask.

7.2.11.2 Add 0.1 g of potassium iodide.

7.2.11.3 Dilute to 50 mL with reagent water. Add 2 mL of concentrated sulfuric acid.

7.2.11.4 Titrate the solution with 0.025 N thiosulfate solution (Section 7.2.8) until a pale, straw yellow color is reached.

7.2.11.5 Add 1 mL of starch indicator and mix well. The solution will turn blue.

7.2.11.6 Continue to titrate with 0.025 N thiosulfate solution until the solution turns clear or milky white.

7.2.11.7 The volume of the thiosulfate solution used in the titration should equal the volume of the biiodate solution added to the flask. If not, adjust the strength of the thiosulfate solution as needed (see Equation 1).

EQUATION 1

$$N_1 = \frac{N_2 V_2}{V_1}$$

Where:
N_1 = *Normality of thiosulfate solution*
V_1 = *Volume (in mL) of thiosulfate solution titrated*
N_2 = *Normality of working biiodate solution*
V_2 = *Volume (in mL) of working biiodate solution used*

Solve the following equation for the volume of reagent water to be used (V_3) to adjust the strength of the thiosulfate solution:

$$V_3 = \left[\frac{N_1 V_4}{0.025}\right] - V_4$$

V_3 = *Volume (in mL) of reagent water used to adjust the thiosulfate solution strength*
V_4 = *Volume (in mL) of remaining thiosulfate solution*

7.2.11.8 If the normality of the thiosulfate solution is less than 0.025 N, add sodium thiosulfate pentahydrate to the solution. Dilute to 1,000 mL with reagent water and retitrate. Adjust the strength of the thiosulfate solution as needed.

7.3 Calibrant Preparation

7.3.1 Stock Calibrant 1,000 mg/L Sulfide (1 L)

7.3.1.1 Wash approximately 10 g of sodium sulfide nonahydrate with reagent water and dry on filter paper.

7.3.1.2 Using only large transparent crystals, dissolve 7.491 g of sodium sulfide nonahydrate in approximately 800 mL of reagent water.

7.3.1.3 Dilute to 1,000 mL with reagent water and mix well.

Note: Store this solution at room temperature in an airtight amber bottle. If stored properly, this reagent is stable for 4–6 weeks.

7.3.2 Intermediate Calibrant 100 mg/L Sulfide (1 L)

7.3.2.1 Dilute 100 mL of stock calibrant (Section 7.3.1) to 1,000 mL with absorbing solution (Section 7.2.6).

Note: Store at room temperature in an airtight amber bottle. Prepare this solution fresh weekly.

7.3.3 Standardized Intermediate Calibrant (1 L)

7.3.3.1 Using a volumetric pipet, add 10 mL of working biiodate solution (Section 7.2.10) into a 250-mL Erlenmeyer flask.

7.3.3.2 Add 100 mL of intermediate calibrant (Section 7.3.2), making sure that the tip of the pipet is below the surface of the liquid.

7.3.3.3 Add 100 mg of potassium iodide and swirl the solution to dissolve.

7.3.3.4 Add 2 mL of concentrated sulfuric acid.

7.3.3.5 Titrate with standardized thiosulfate solution (Section 7.2.11) until a pale, straw yellow color is reached.

7.3.3.6 Add 1 mL of starch indicator and mix well. The solution will turn blue.

7.3.3.7 Continue to titrate with standardized thiosulfate solution until the solution turns clear or milky white.

7.3.3.8 Titrate a blank in the same manner. For the blank use 100 mL of absorbing solution instead of the intermediate calibrant.

7.3.3.9 Since 1 mL of 0.025 N potassium biiodate solution reacts with 0.4 mg of sulfide, use the following equation to calculate the concentration of sulfide in the standardized intermediate calibrant (see Equation 2).

Note: Store at room temperature in an airtight amber bottle. Prepare this solution fresh weekly.

EQUATION 2

$$\text{mg/L Sulfide} = 400 \times \left[\frac{A - (B_1 - B_2)}{C_1} \right]$$

Where:
A = Volume (in L) of working potassium biiodate solution
B_1 = Volume (in L) of standardized thiosulfate solution used to titrate the intermediate calibrant
B_2 = Volume (in L) of standardized thiosulfate solution used to titrate the blank
C_1 = Volume (in L) of intermediate calibrant

7.3.4 Working Calibrants (100 mL)

7.3.4.1 Add the designated volumes of standardized intermediate calibrant (see Equation 3) to the required number of 100-mL volumetric flasks that each contain approximately 80 mL of reagent water.

7.3.4.2 Dilute each solution to the mark with reagent water and mix well.

Note: Prepare working calibrants fresh daily.

EQUATION 3

$$C_1V_1 = C_2V_2$$

Where:
C_1 = *Concentration (in mg/L) of stock solution (standardized intermediate calibrant)*
V_1 = *Volume (in L) of stock solution to be used*
C_2 = *Desired concentration (in mg/L) of working calibrant to be prepared*
V_2 = *Final volume (in L) of working calibrant to be prepared*

By solving this equation for the volume of stock solution to be used (V_1), the following equation is obtained:

$$V_1 = \frac{C_2V_2}{C_1}$$

Since the desired concentration (C_2), the final volume (V_2), and the concentration of the stock solution (C_1) are all known for any given calibrant concentration in a defined volume, the volume of stock solution to be used (V_1) is easily calculated.

7.3.4.3 Calibrants covering the entire range of this analysis can be prepared from the following tables.

7.3.4.3.1 For range 0.005–1.00 mg/L using a 200-µL sample loop:

Final Concentration (mg/L)	Vol. of Inter. Cal. (µL)	Nominal Conc. of Inter. Cal. (mg/L)	Final Volume (mL)
0.005	5	100	100
0.010	10	100	100
0.050	50	100	100
0.100	100	100	100
0.500	500	100	100
1.00	1,000	100	100

7.3.4.3.2 For range 0.100–20.0 mg/L using a 100-µL sample loop:

Final Concentration (mg/L)	Vol. of Inter. Cal. (mL)	Nominal Conc. of Inter. Cal. (mg/L)	Final Volume (mL)
0.100	0.10	100	100
1.00	1.0	100	100
5.00	5.0	100	100
10.0	10	100	100
15.0	15	100	100
20.0	20	100	100

8.0 Sample Collection, Preservation, and Storage

8.1 Samples should be collected in plastic or glass bottles that have been thoroughly cleaned and rinsed with reagent water (Section 7.2.1).

8.2 The volume of sample collected should be sufficient to ensure that a representative sample is obtained, replicate analysis is possible, and waste disposal is minimized.

8.3 Collect samples with a minimum of aeration. Sample analysis should be performed as soon as possible to eliminate loss of analyte.

8.4 Preserve samples by adding 200 µL of 2 N zinc acetate (Section 7.2.7) to a 100-mL bottle. Fill the bottle completely with sample and cap tightly.

8.5 Holding time for preserved samples is 24 hours from the time of collection (Reference 15.3).

9.0 Quality Control

Note: The following QC procedures are provided for reference purposes only and are not a substitute for any QC procedures that may be required for regulatory compliance.

9.1 It is recommended that each laboratory that uses this method operate a formal quality control program. The minimum requirements of such a program should consist of an initial demonstration of laboratory capability and the periodic analysis of Laboratory Control Samples (LCSs) and Matrix Spike/Matrix Spike Duplicates (MS/MSDs) as a continuing check on performance. Laboratory performance should be compared to established performance criteria to determine if the results of the analyses meet the performance characteristics of the method.

9.2 Method Detection Limit (MDL)—To establish the ability to detect sulfide at low levels, the analyst should determine the MDL using the apparatus, reagents, and calibrants that will be used in the practice of this method. An MDL less than or equal to the MDL listed in Section 1.2 should be achieved prior to practice of this method.

9.2.1 An MDL is calculated by analyzing a matrix spike at a concentration of two to three times the expected detection limit of the analyzer. Seven consecutive replicate analyses of this matrix spike should be analyzed, and the MDL should be calculated using Equation 4.

EQUATION 4

$$MDL = (t) \times (S)$$

Where:
t = Student's t value for a 99% confidence level and a standard deviation estimate with n–1 degrees of freedom (t = 3.14 for seven replicates)
S = Standard deviation of the replicate analyses

9.2.2 It is recommended that the MDL be calculated after every six months of operation, when a new operator begins work, or whenever there is any significant change in the instrument response.

9.3 Analyses of MS/MSD samples are required to demonstrate method accuracy and precision and to monitor matrix interferences (interferences caused by the sample matrix).

9.3.1 Matrix Spike/Matrix Spike Duplicate (MS/MSD)—The laboratory should spike, in duplicate, a minimum of 10% of all samples (one sample in duplicate in each batch of 10 samples) from a given sampling site.

9.3.2 The concentration of the spike in the sample shall be determined as follows:

9.3.2.1 If, as in compliance monitoring, the concentration of sulfide in the sample is being checked against a regulatory concentration limit, the spiking level shall be at that limit.

9.3.2.2 If the concentration of sulfide in a sample is not being checked against a limit, the spike shall be at the concentration of the LCS or at least four times greater than the MDL.

9.4 Analyses of Laboratory Reagent Blanks (LRBs) are required to demonstrate freedom from contamination and that the compounds of interest and interfering compounds have not been carried over from a previous analysis.

9.5 As part of the QC program for the laboratory, method precision and accuracy for samples should be assessed and records should be maintained.

9.5.1 An LCS should be analyzed with every sample batch, and the mean (m) and the standard deviation (S) should be recorded. After multiple analyses, the mean should be plotted with limits of $m+2S$ and $m–2S$. The mean and the limits should be recalculated after every 5–10 new measurements.

9.5.2 If the LCS measurement falls outside the range calculated in Section 9.5.1, then the problem should be addressed, and that sample batch should be reanalyzed if necessary.

9.6 Reference Sample—To demonstrate that the analytical system is in control, the laboratory may wish to periodically test an external reference sample, such as a Standard Reference Material (SRM) available from the National Institute of Standards and Technology (NIST). Corrective action should be taken if the measured concentration significantly differs from the stated concentration.

10.0 Configuration and Start-up

10.1 Instrument Configuration

10.1.1 Configure the OI Analytical Flow Solution 3000 Analyzer according to the Operator's Manual and verify that each module is properly powered on.

10.1.2 Verify that the Sulfide Cartridge (Part #A002762) is configured as illustrated in the flow diagram shown in Section 17.0.

10.1.3 Connect the appropriate pump tubes to the cartridge and to their appropriate reagent containers according to the flow diagram.

10.2 Instrument Stabilization

10.2.1 Connect the reagent pump tubes to a reagent bottle containing the start-up solution (Section 7.2.2). Start the pump at low speed, allowing the start-up solution to flow through the entire system.

10.2.2 Verify that the flowcell of each detector is purged of all bubbles and that the flow is stable and free from surging before proceeding.

10.3 Baseline Verification

10.3.1 Create and save a Method in WinFLOW. Refer to the WinFLOW Operator's Manual (Reference 15.5) for help on creating a Method.

10.3.2 Create and save a Sample Table in WinFLOW that will be used to generate a calibration curve using at least three calibrants that cover the full range of expected concentrations in the samples to be analyzed. This Sample Table should also be used to analyze all necessary QC samples as well as the analytical batch of samples to be analyzed. For help on creating a Sample Table, refer to the WinFLOW Operator's Manual (Reference 15.5).

10.3.3 Select **Collect Data** in the WinFLOW main window, enter the user's identification, select the appropriate Method and Sample Table, and begin to collect baseline data. Very sharp fluctuations in the baseline and/or consistent drifting are typically signs of bubbles in the flowcell. The flowcell must be free of bubbles prior to beginning analysis.

10.4 Calibration and Standardization

10.4.1 Prepare a series of at least three working calibrants using the stock solutions (Section 7.3) according to Equation 3, covering the desired analysis range.

10.4.2 Place the calibrants in the autosampler in order of decreasing concentration. Each calibrant should be analyzed according to the analytical procedures in Section 11.0. A calibration curve will be calculated by the WinFLOW software.

10.4.3 Acceptance or control limits for the calibration results should be established using the difference between the measured value of each calibrant and the corresponding "true" concentration.

10.4.4 Each calibration curve should be verified by analysis of a Laboratory Control Sample (LCS, Section 9.5). Using WinFLOW software, calibration, verification, and sample analysis may be performed in one continuous analysis.

11.0 Procedure

11.1 Analysis

11.1.1 Begin pump flow with the start-up solution (Section 7.2.2). Verify a stable baseline (Section 10.3).

11.1.2 After the baseline has been verified according to Section 10.3, place all reagents on-line and allow to pump at least 10–15 minutes. Verify there are no bubbles in the flowcell. Obtain a stable baseline at 660 nm and autozero the baseline before beginning analysis.

11.1.3 Load the sampler tray with calibrants, blanks, samples, and QC samples.

Note: The matrix of the working standards, blanks, and QC samples should match that of the samples being analyzed.

11.1.4 Using the Method and Sample Table created for the analytical batch to be analyzed and with the baseline verified to be stable, begin the analysis by selecting the "Fast Forward" button on the left side of the Data Analysis window in WinFLOW. This will initiate the sequential analysis of samples as defined in the Sample Table.

11.1.5 When analysis is complete, pump start-up solution through the system for at least 10–15 minutes. Stop the pump, release the tension on all pump tubes, and power off the system.

11.2 Operating Notes

11.2.1 The carrier solution (Section 7.2.2) must be thoroughly degassed by submitting the solution to a strong vacuum for 15–20 minutes with stirring and/or heating.

11.2.2 Use nitrogen as the segmentation gas to prevent oxidation of sulfide.

11.2.3 If the baseline is noisy, refilter all reagents except the absorbing solution through a 0.45-µm filter to remove particulates.

12.0 Data Analysis and Calculations

12.1 The calibration curve allows for accurate quantitation of the concentration in each sample.

12.2 WinFLOW software reports the concentration of each sample relative to the calibration curve.

13.0 Method Performance

Range:	
200-µL sample loop	0.005–1.00 mg/L
100-µL sample loop	0.100–20.0 mg/L
Throughput:	28 samples/hour
Precision:	
0.005 mg/L	<3% RSD
0.100 mg/L	<3% RSD
1.00 mg/L	<2% RSD
10.0 mg/L	<1% RSD
20.0 mg/L	<1% RSD
Method Detection Limit (MDL):	0.002 mg/L

14.0 Pollution Prevention and Waste Management

14.1 It is the laboratory's responsibility to comply with all federal, state, and local regulations governing waste management, particularly the hazardous waste identification rules and land-disposal restrictions. In addition, it is the laboratory's responsibility to protect air, water, and land resources by minimizing and controlling all releases from fume hoods and bench operations. Also, compliance is required with any sewage discharge permits and regulations.

14.2 For further information on waste management, consult Section 13.6 of *Less is Better: Laboratory Chemical Management for Waste Reduction* (Reference 15.1).

15.0 References

15.1 *Less is Better: Laboratory Chemical Management for Waste Reduction.* Available from the American Chemical Society, Department of Government Regulations and Science Policy, 1155 16th Street, NW, Washington, DC, 20036.

15.2 Sulfide (Colorimetric, Methylene Blue). *Methods for Chemical Analysis of Water and Wastewater;* EPA-600/4-79-020; U.S. Environmental Protection Agency, Office of Research and Development, Environmental Monitoring and Support Laboratory: Cincinnati, OH, 1984; Method 376.2.

15.3 Sample Preservation. *Methods for Chemical Analysis of Water and Wastes*; EPA-600/4-79-020; U.S. Environmental Protection Agency, Office of Research and Development, Environmental Monitoring and Support Laboratory: Cincinnati, OH, 1984; xvii.

15.4 *Standard Methods for the Examination of Water and Wastewater*, 20th ed.; American Public Health Association: Washington, D.C., 1998.

15.5 WinFLOW Software and Operator's Manual (Part #A002877). Available from OI Analytical, P.O. Box 9010, College Station, TX, 77842-9010.

16.0 Glossary of Definitions and Purposes

The definitions and purposes are specific to this method but have been conformed to common usage as much as possible.

16.1 Units of weights and measures and their abbreviations

16.1.1 Symbols

°C	degrees Celsius
%	percent
±	plus or minus
≥	greater than or equal to
≤	less than or equal to

16.1.2 Alphabetical characters

g	gram
L	liter
mg	milligram
mg/L	milligram per liter
µg	microgram
µg/L	microgram per liter
mL	milliliter
ppm	parts per million
ppb	parts per billion
M	molar solution
N	normal solution

16.2 Definitions

16.2.1 Laboratory Control Sample (LCS)—An aliquot of LRB to which a quantity of the analyte of interest is added in the laboratory. The LCS is analyzed like a sample. Its purpose is to determine whether the methodology is in control and whether the laboratory is capable of making accurate and precise measurements.

16.2.2 Laboratory Reagent Blank (LRB)—An aliquot of reagent water and other blank matrix that is treated like a sample, including exposure to all glassware, equipment, and reagents that are used with other samples. The LRB is used to determine if the method analyte or other interferences are present in the laboratory environment, reagents, or apparatus.

16.2.3 Matrix Spike/Matrix Spike Duplicate (MS/MSD)—An aliquot of an environmental sample to which a quantity of the method analyte is added in the laboratory. The MS/MSD is analyzed like a sample. Its purpose is to determine whether the sample matrix contributes bias to the analytical results. The background concentration of the analyte in the sample matrix must be determined in a separate aliquot, and the measured values in the MS/MSD must be corrected for the background concentration.

16.2.4 Method Detection Limit (MDL)—The minimum concentration of a substance that can be measured and reported with 99% confidence that the analyte concentration is greater than zero.

17.0 Figures

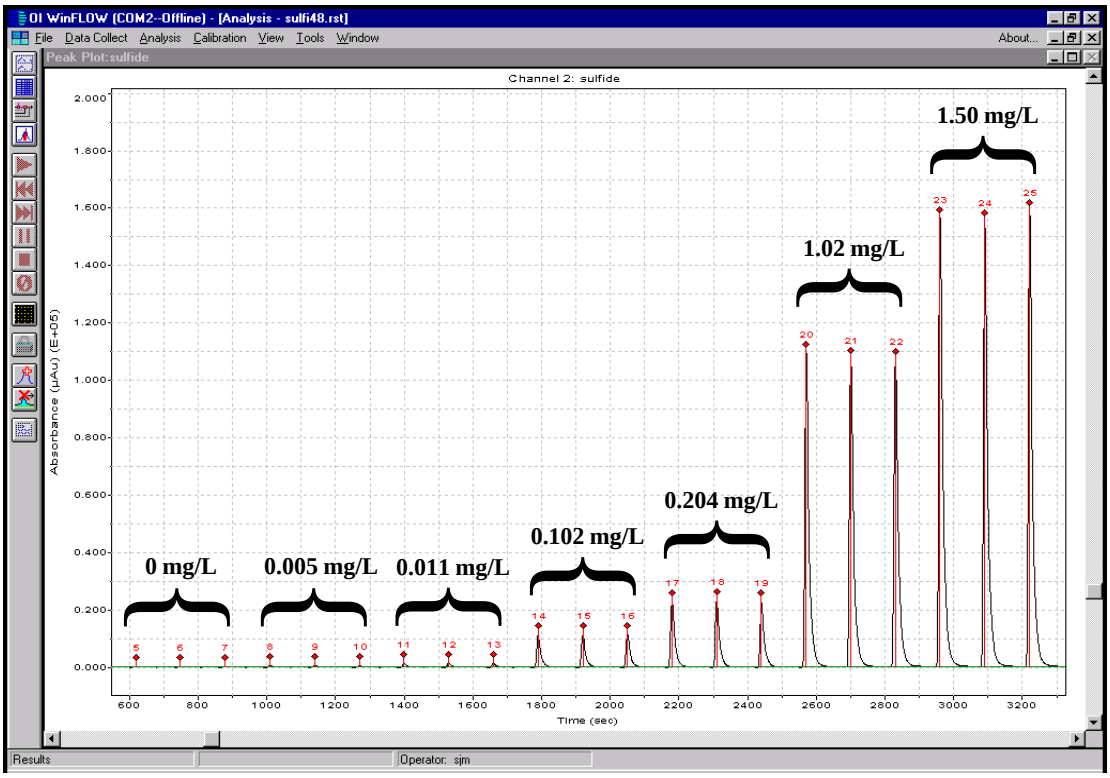


Figure 1. Calibration (0.005–1.50 mg/L) Using a 200-µL Sample Loop

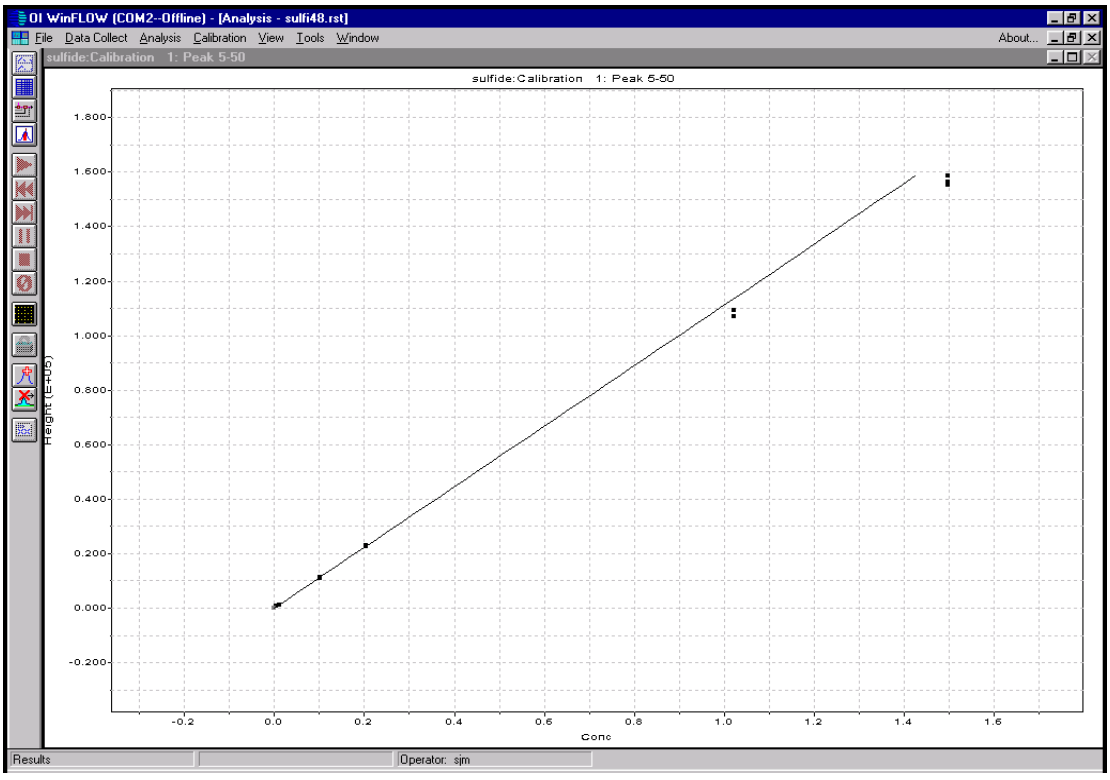


Figure 2. Calibration Curve (0.005–1.50 mg/L) Using a 200-µL Sample Loop

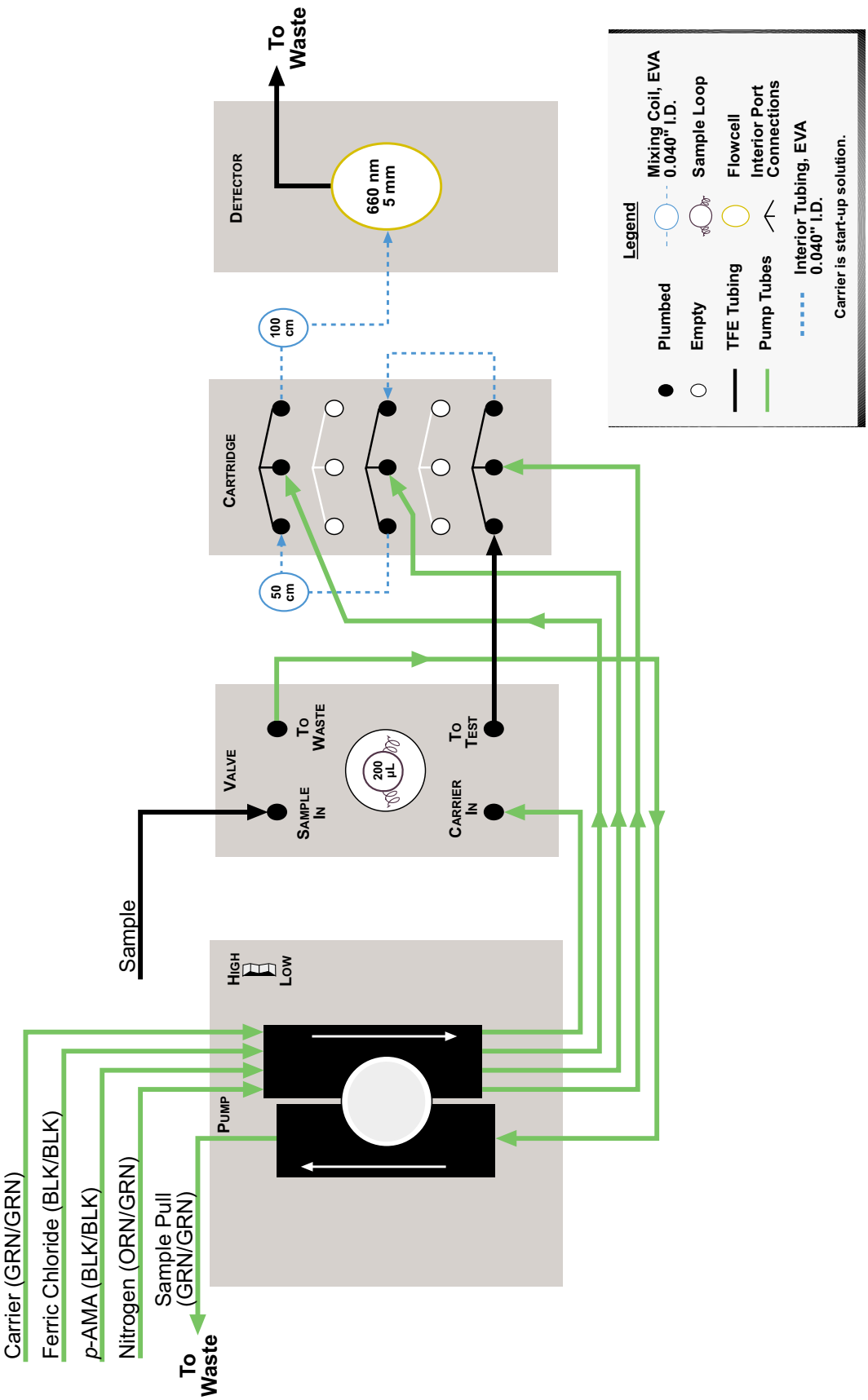


Figure 3. Detailed Flow Diagram for Sulfide by FIA on a Flow Solution 3000, Cartridge Part #A002762

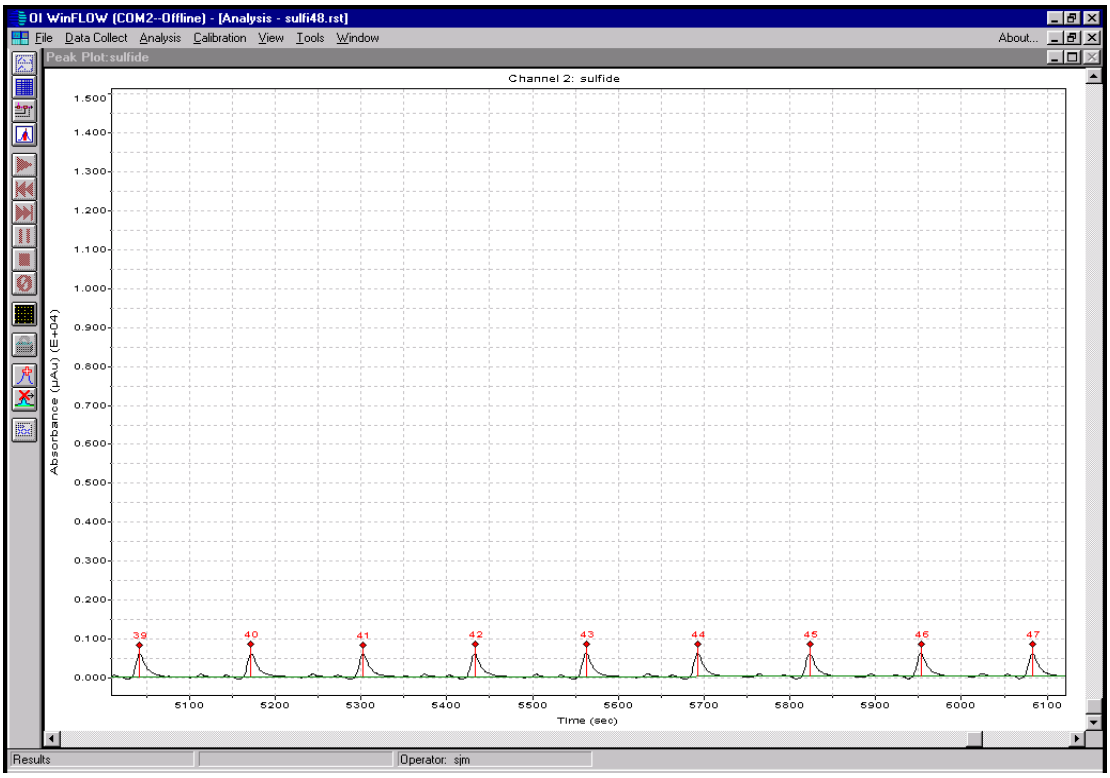


Figure 4. Precision at 0.005 mg/L (<3% RSD) Using a 200-µL Sample Loop

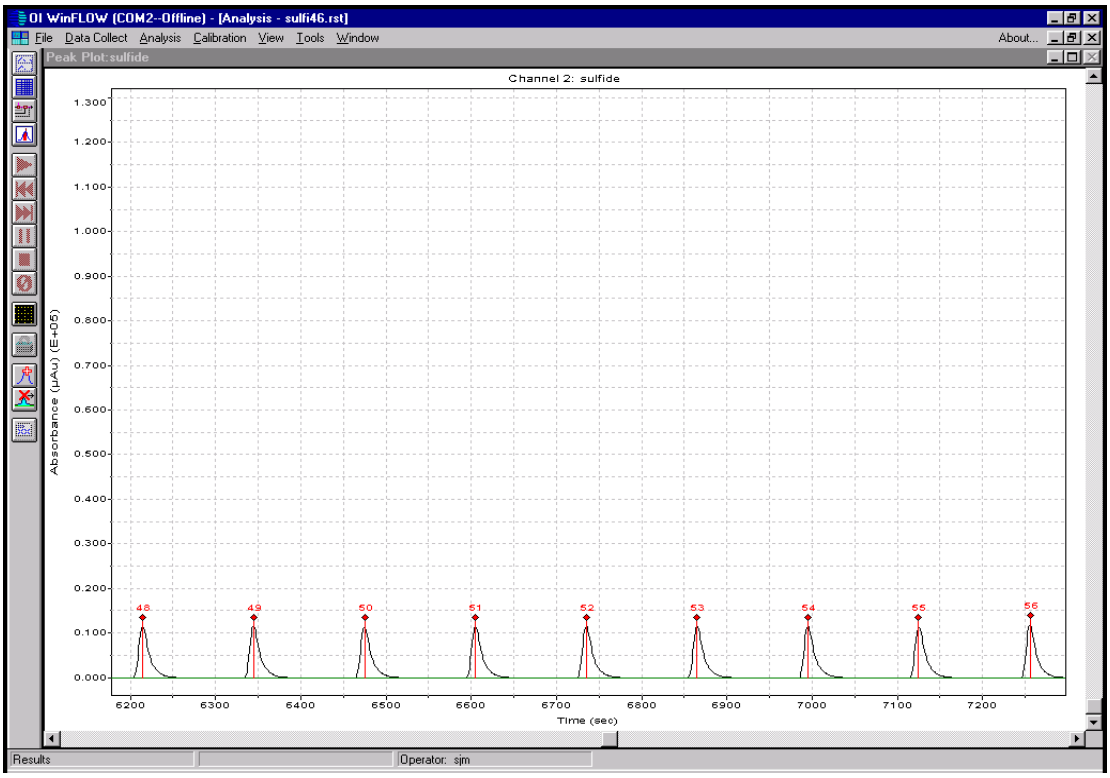


Figure 5. Precision at 0.102 mg/L (<3% RSD) Using a 200-µL Sample Loop

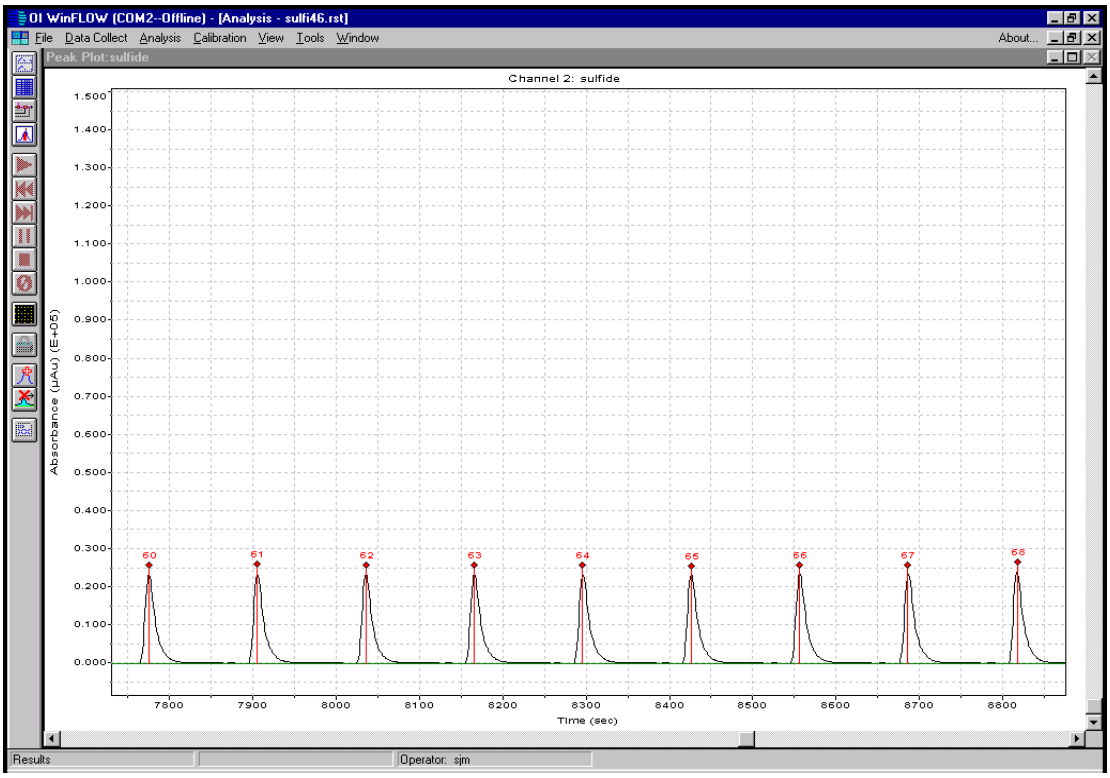


Figure 6. Precision at 0.204 mg/L (<2% RSD) Using a 200-µL Sample Loop

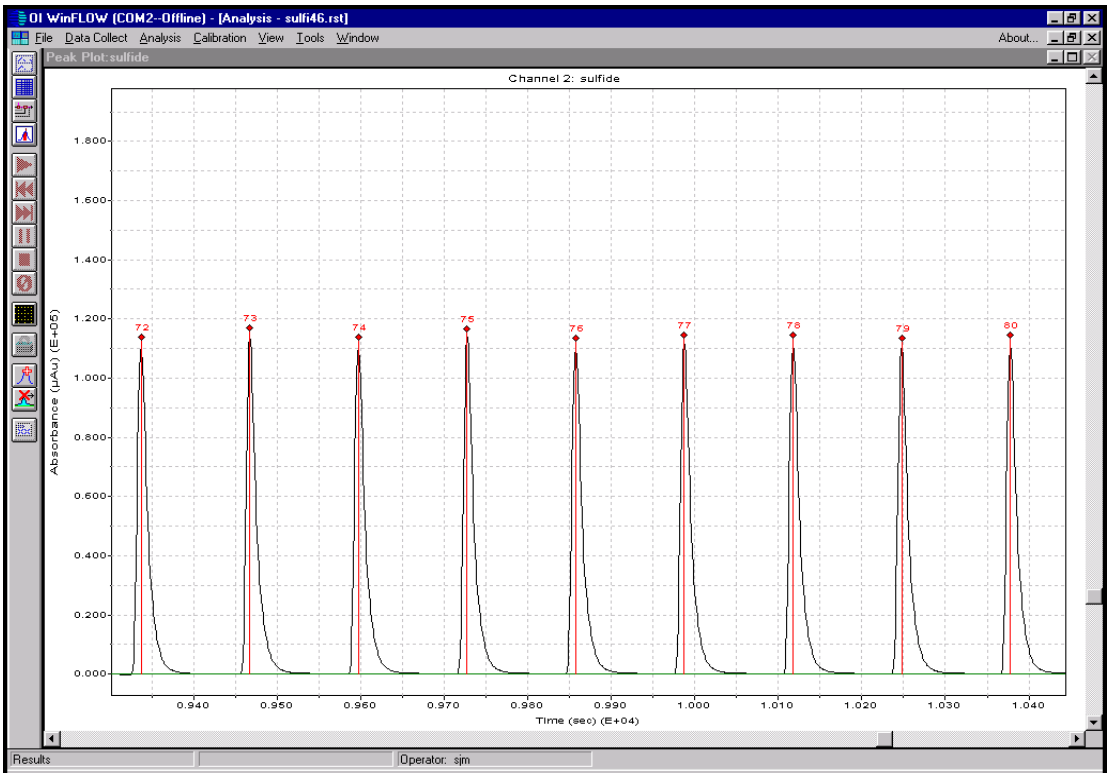


Figure 7. Precision at 1.02 mg/L (<2% RSD) Using a 200-µL Sample Loop

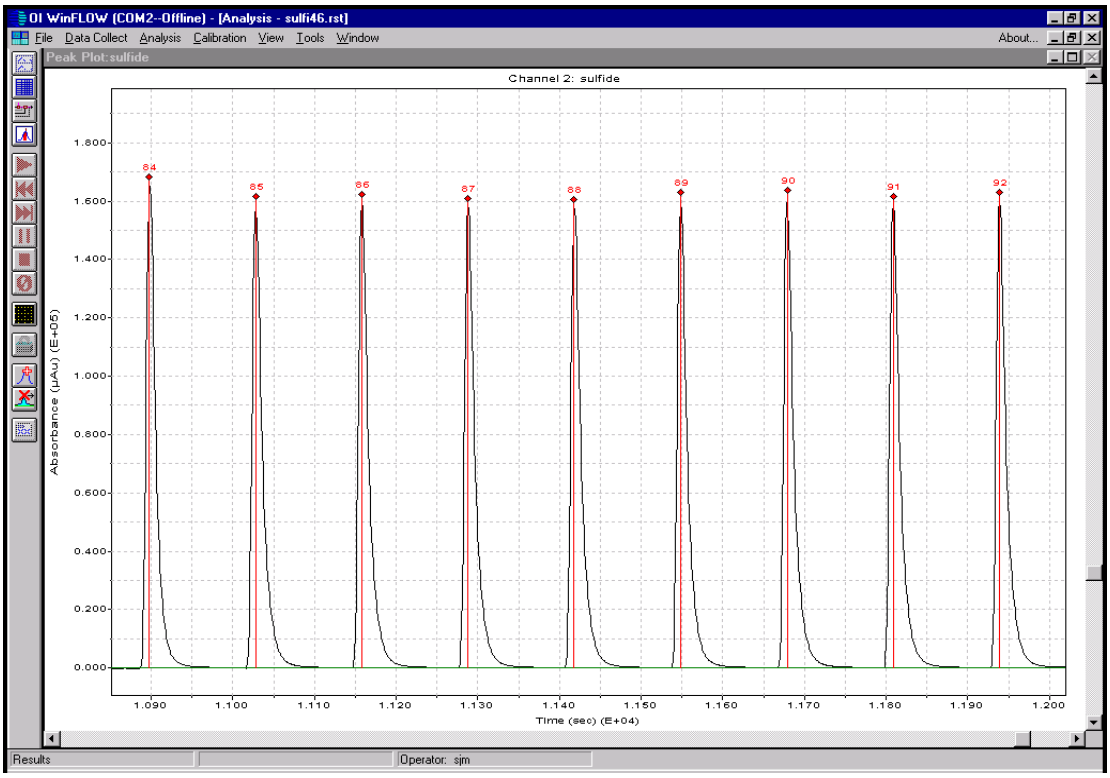


Figure 8. Precision at 1.50 mg/L (<2% RSD) Using a 200-µL Sample Loop

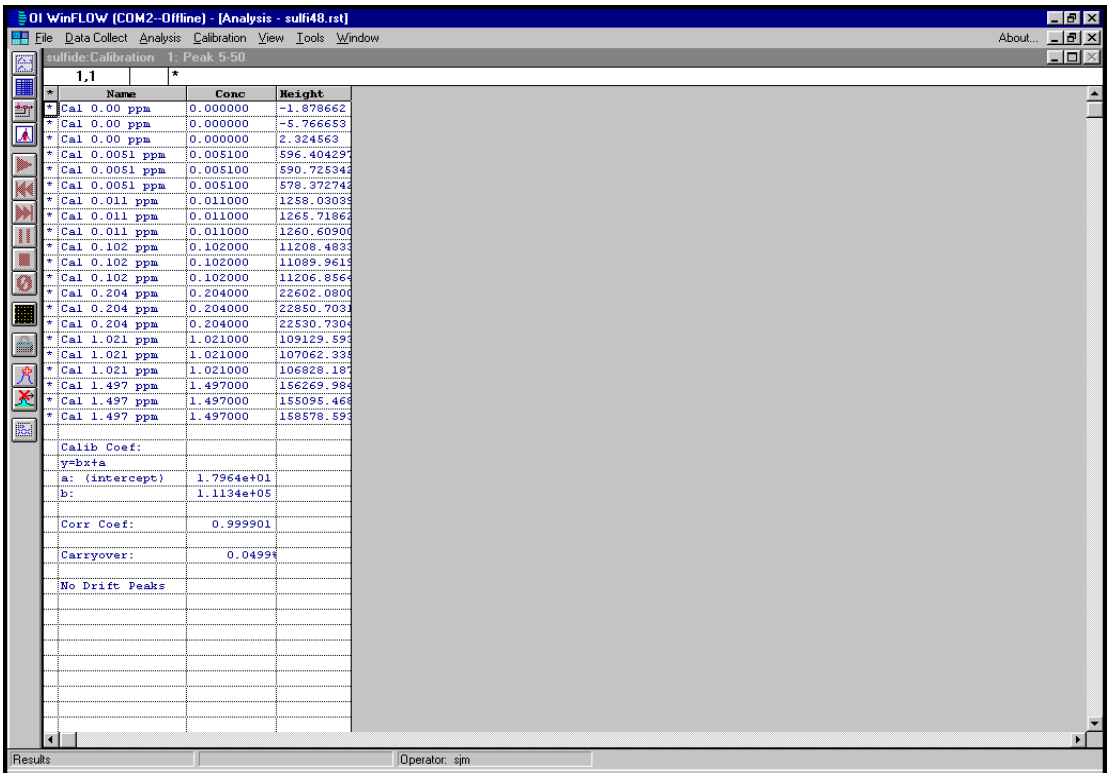


Figure 9. Calibration Results (0.005–1.50 mg/L) Using a 200-µL Sample Loop

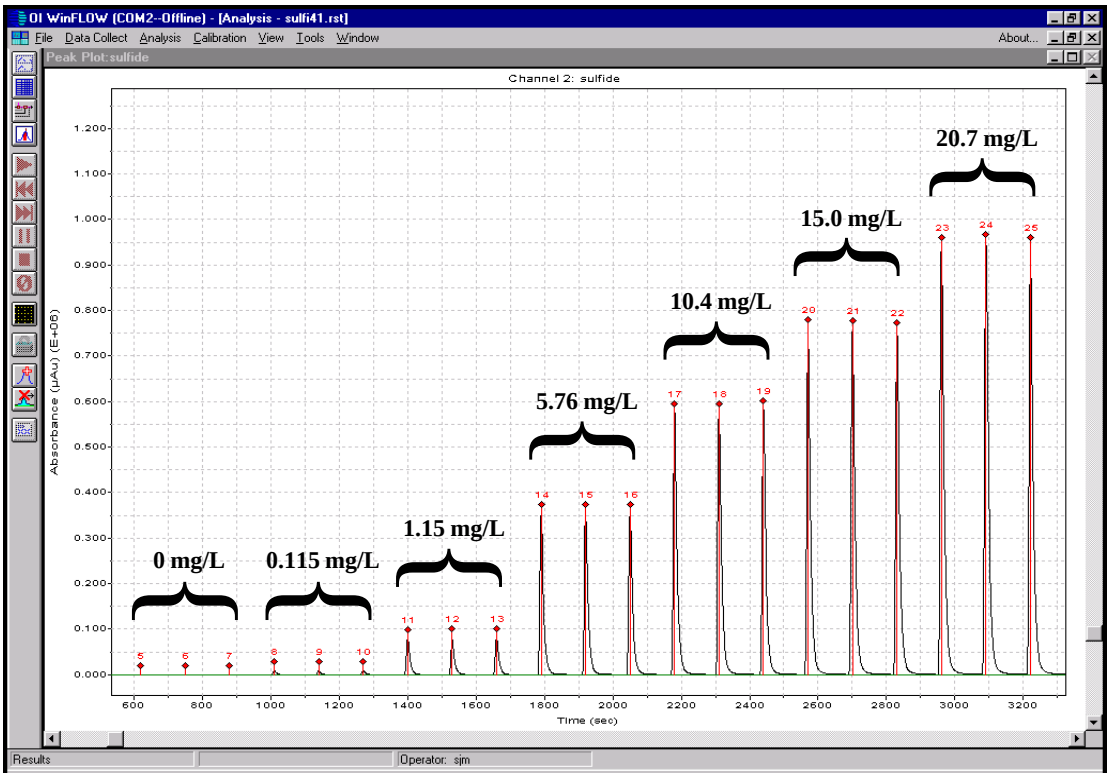


Figure 10. Calibration (0.115–20.7 mg/L) Using a 100-µL Sample Loop

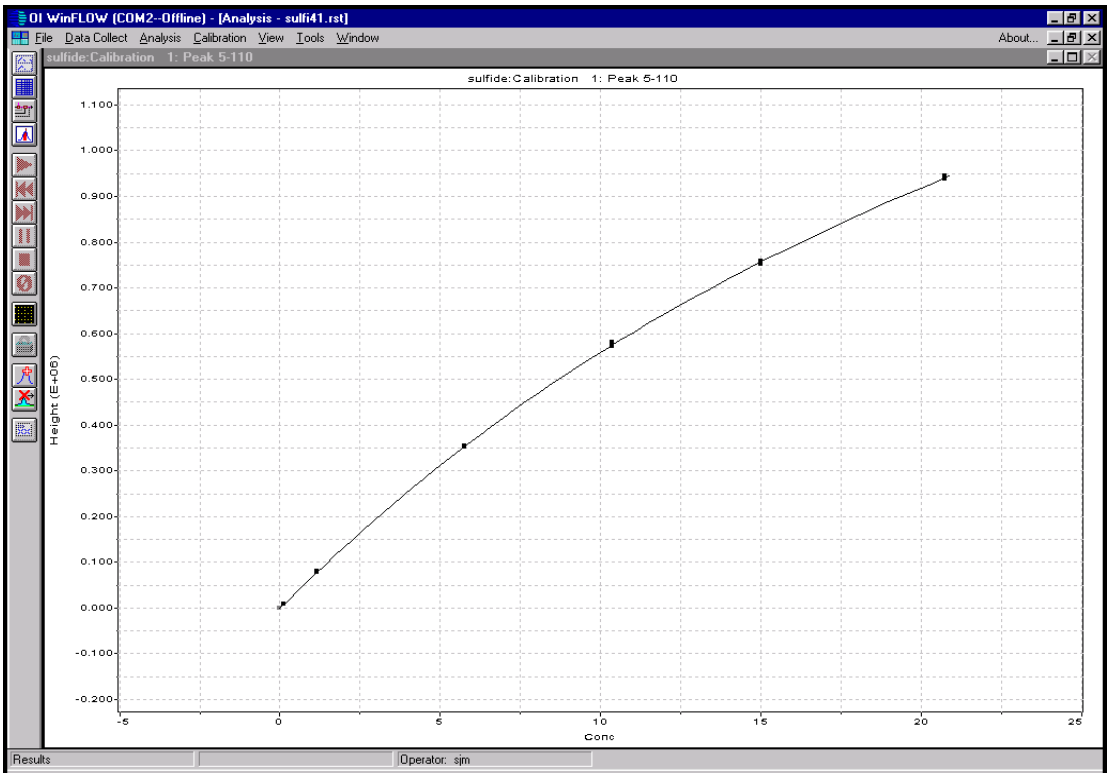


Figure 11. Calibration Curve (0.115–20.7 mg/L) Using a 100-µL Sample Loop

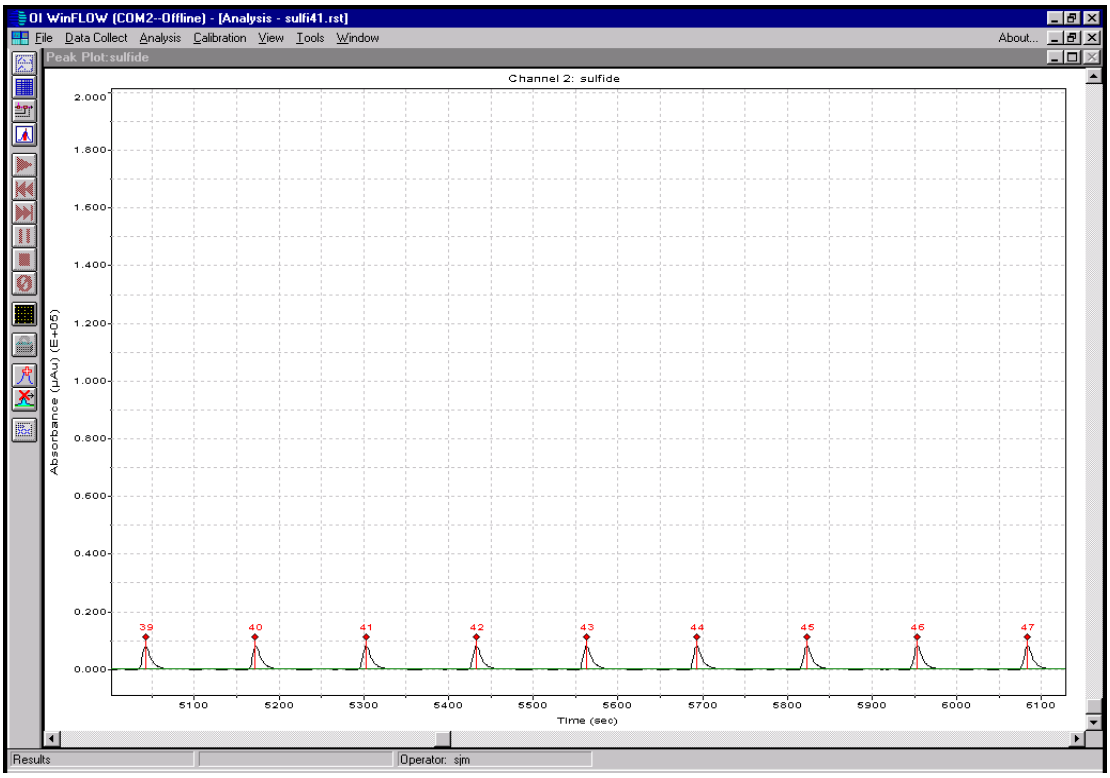


Figure 12. Precision at 0.115 mg/L (<3% RSD) Using a 100-µL Sample Loop

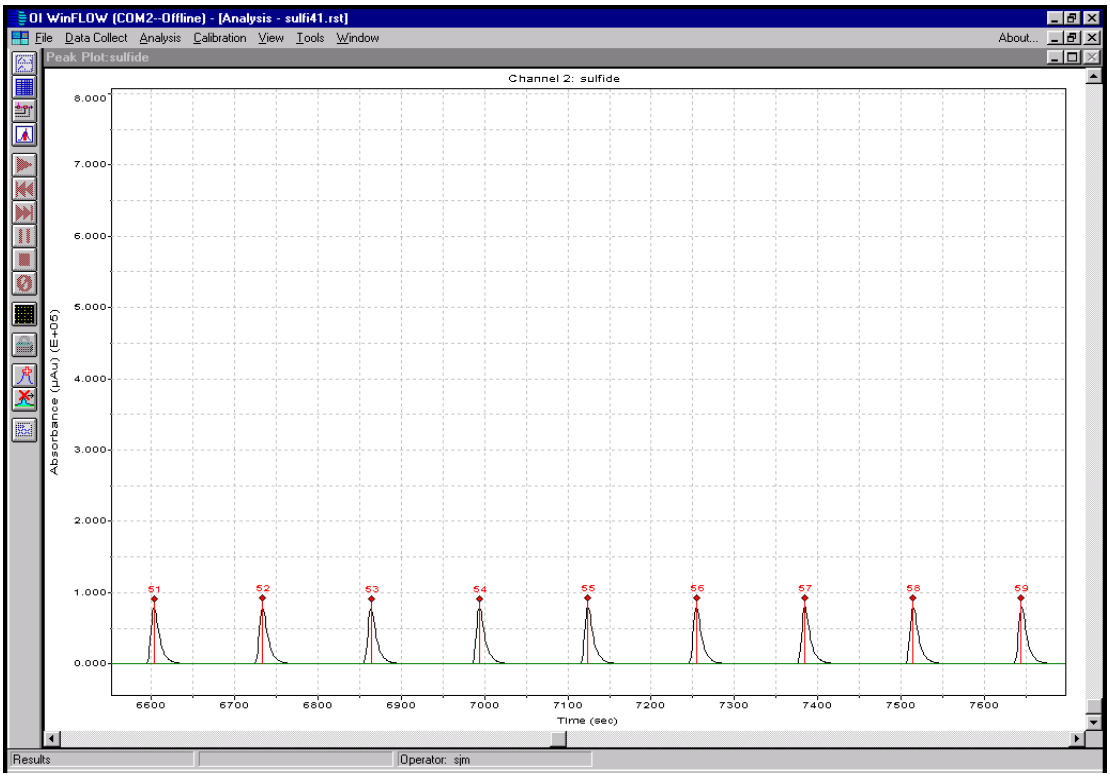


Figure 13. Precision at 1.15 mg/L (<2% RSD) Using a 100-µL Sample Loop

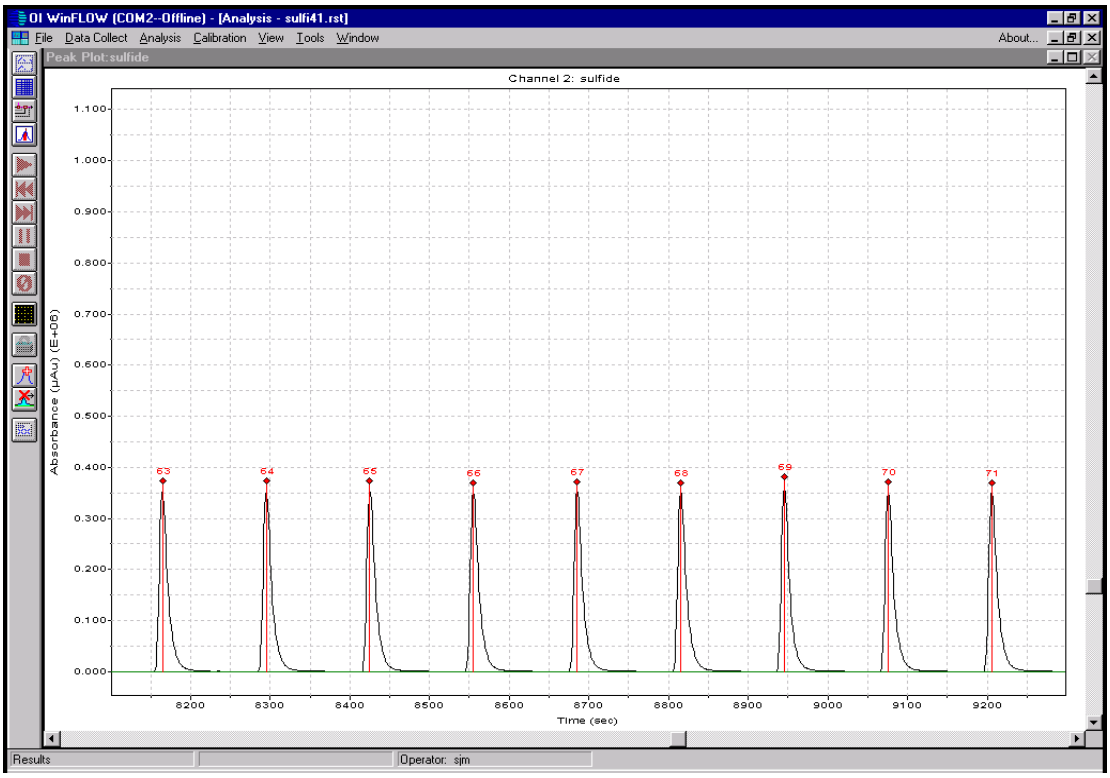


Figure 14. Precision at 5.76 mg/L (<2% RSD) Using a 100-µL Sample Loop

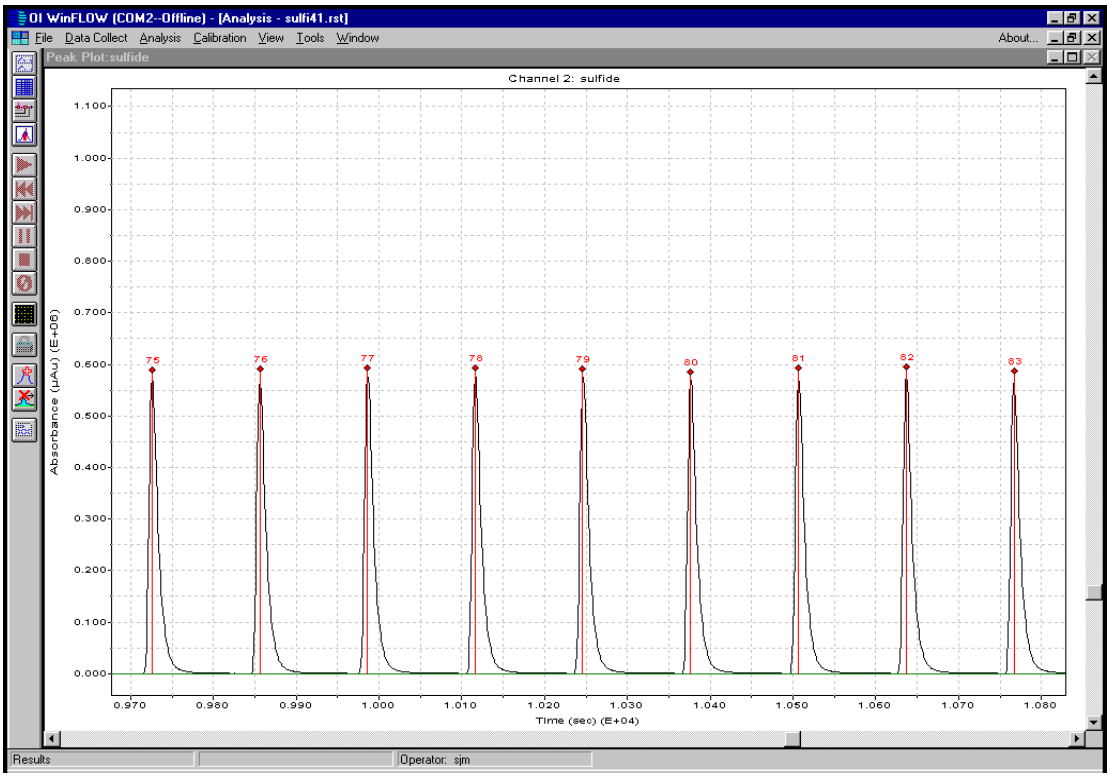


Figure 15. Precision at 10.4 mg/L (<1% RSD) Using a 100-µL Sample Loop

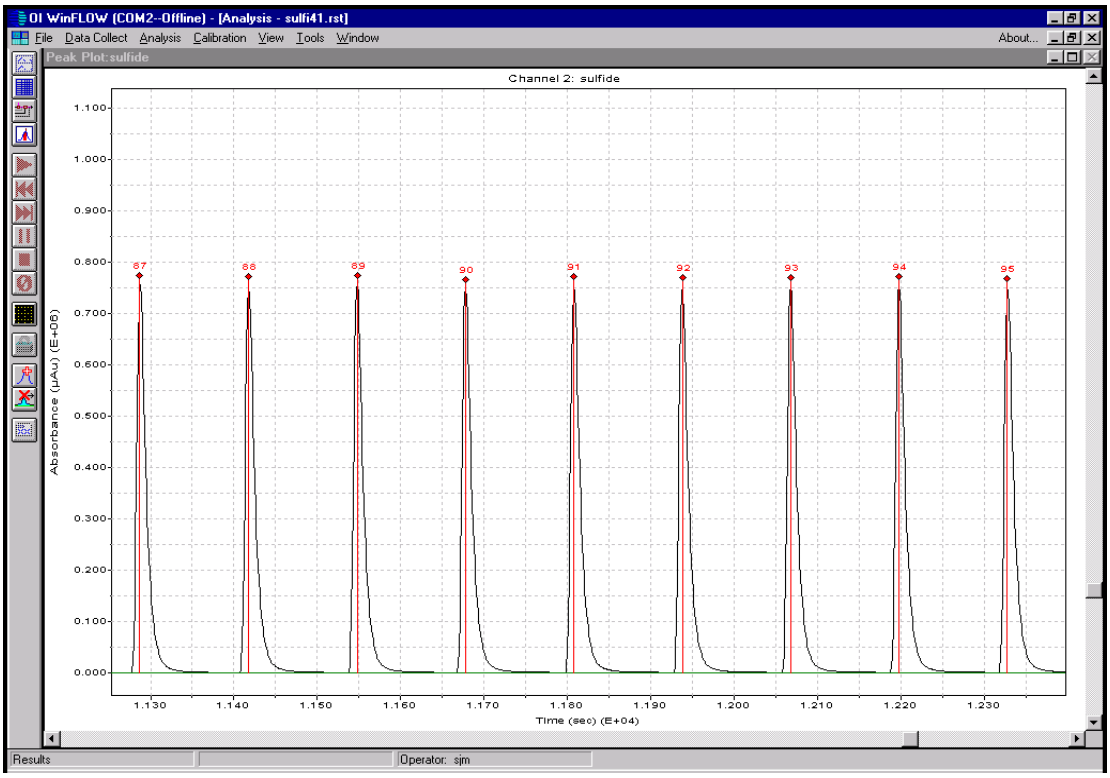


Figure 16. Precision at 15.0 mg/L (<1% RSD) Using a 100-µL Sample Loop

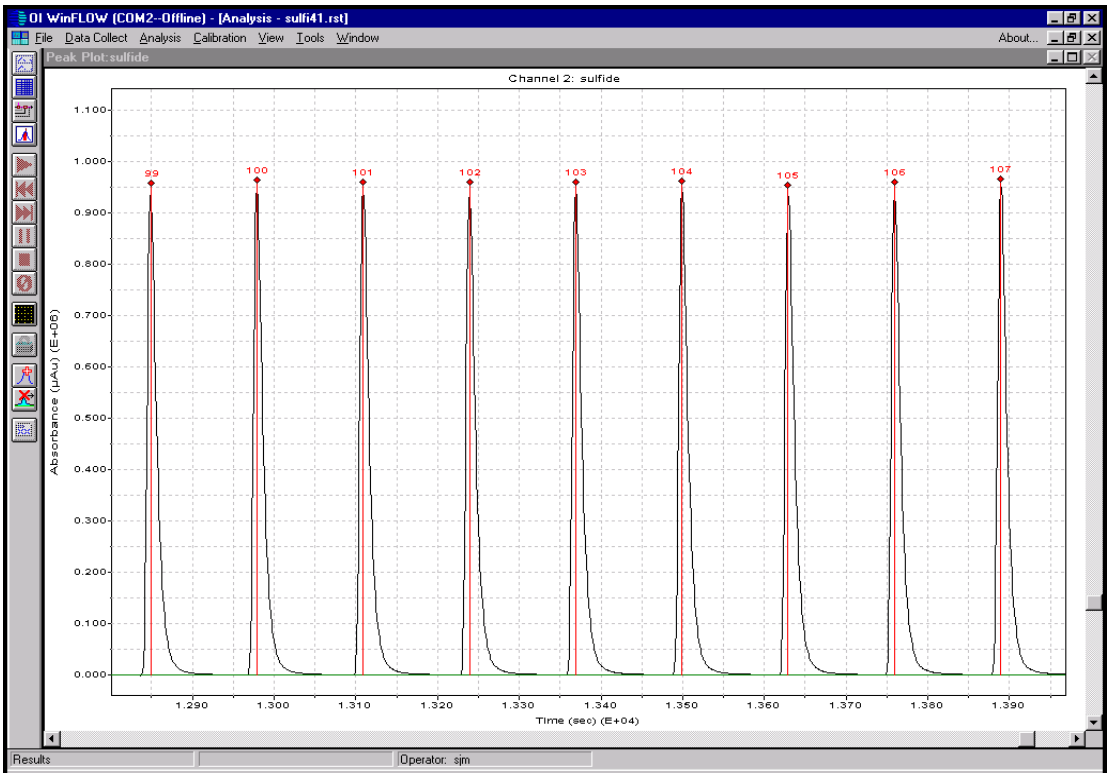


Figure 17. Precision at 20.7 mg/L (<1% RSD) Using a 100-µL Sample Loop

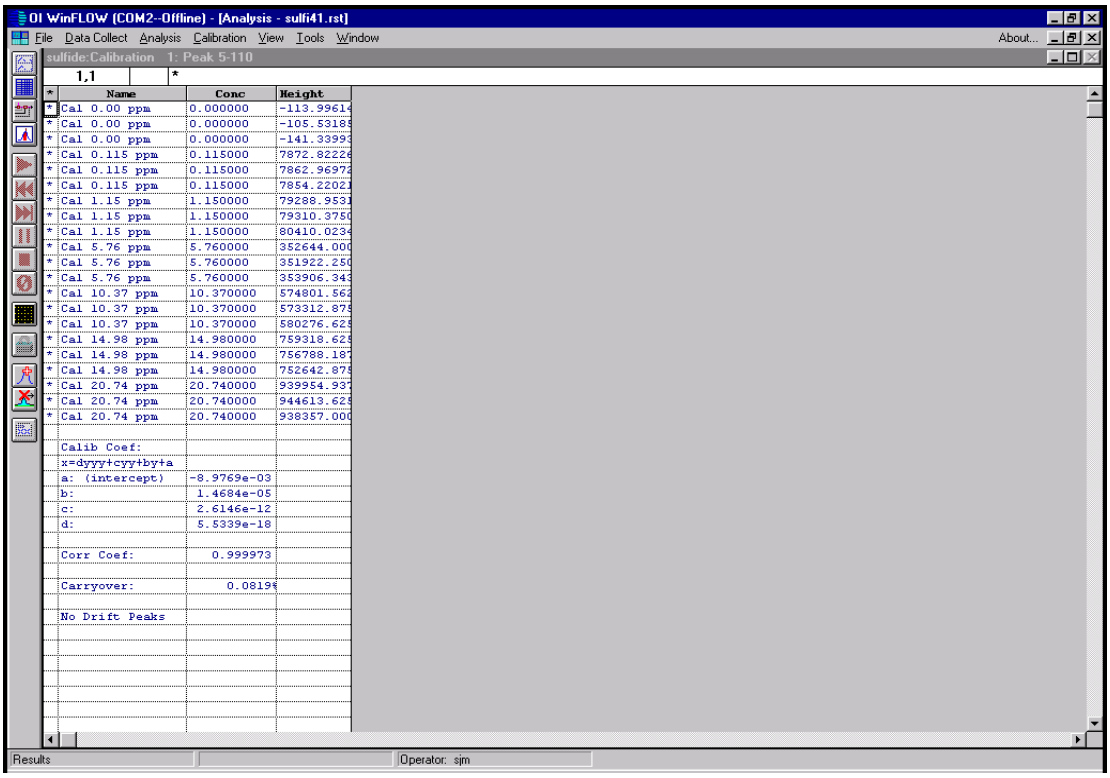


Figure 18. Calibration Results (0.115–20.7 mg/L) Using a 100-µL Sample Loop

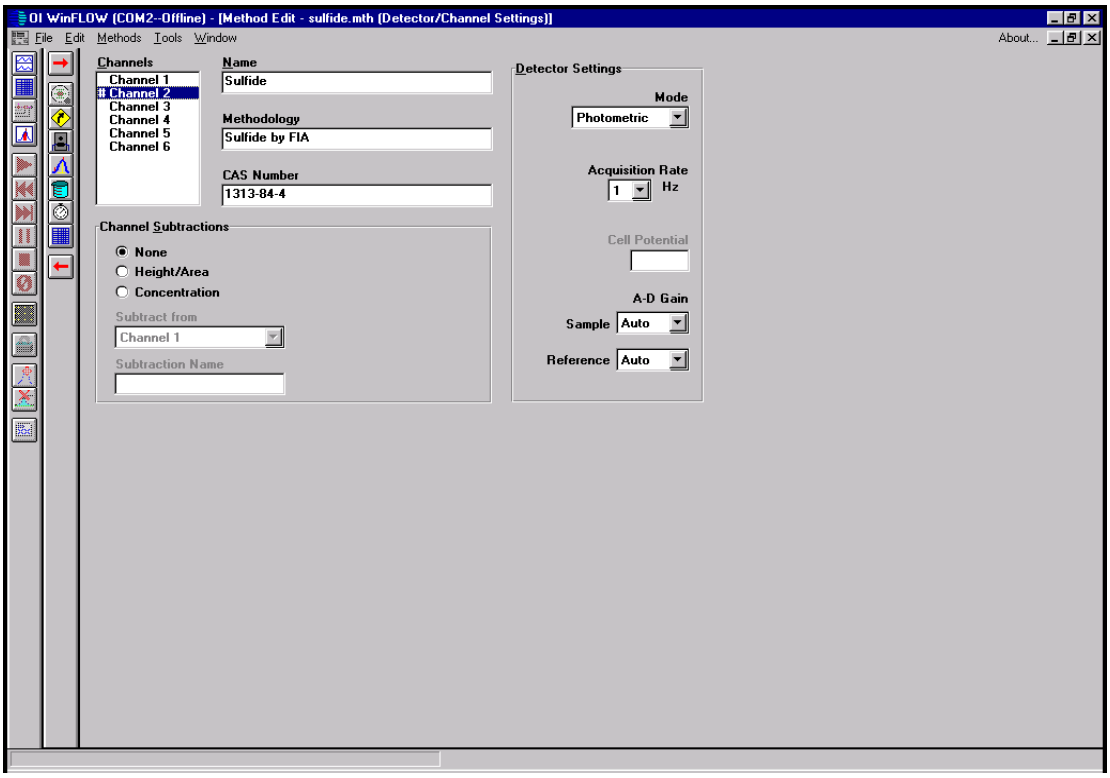


Figure 19. WinFLOW Method Editor—Detector/Channel Settings

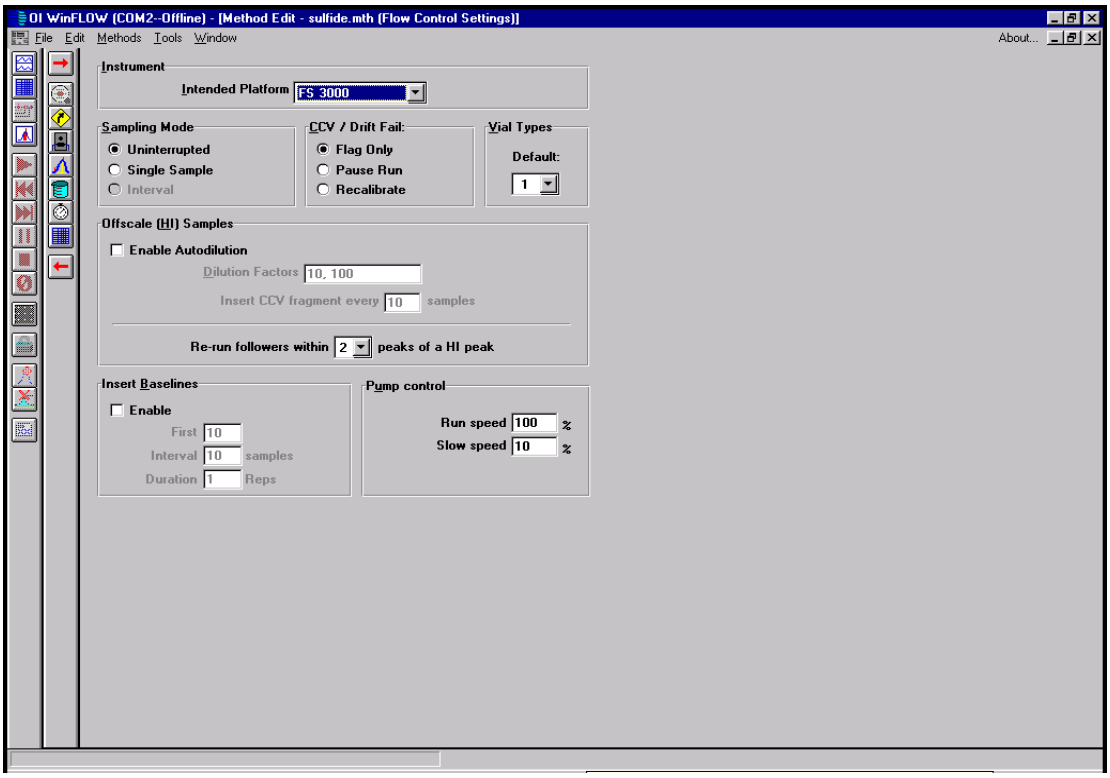


Figure 20. WinFLOW Method Editor—Flow Control Settings

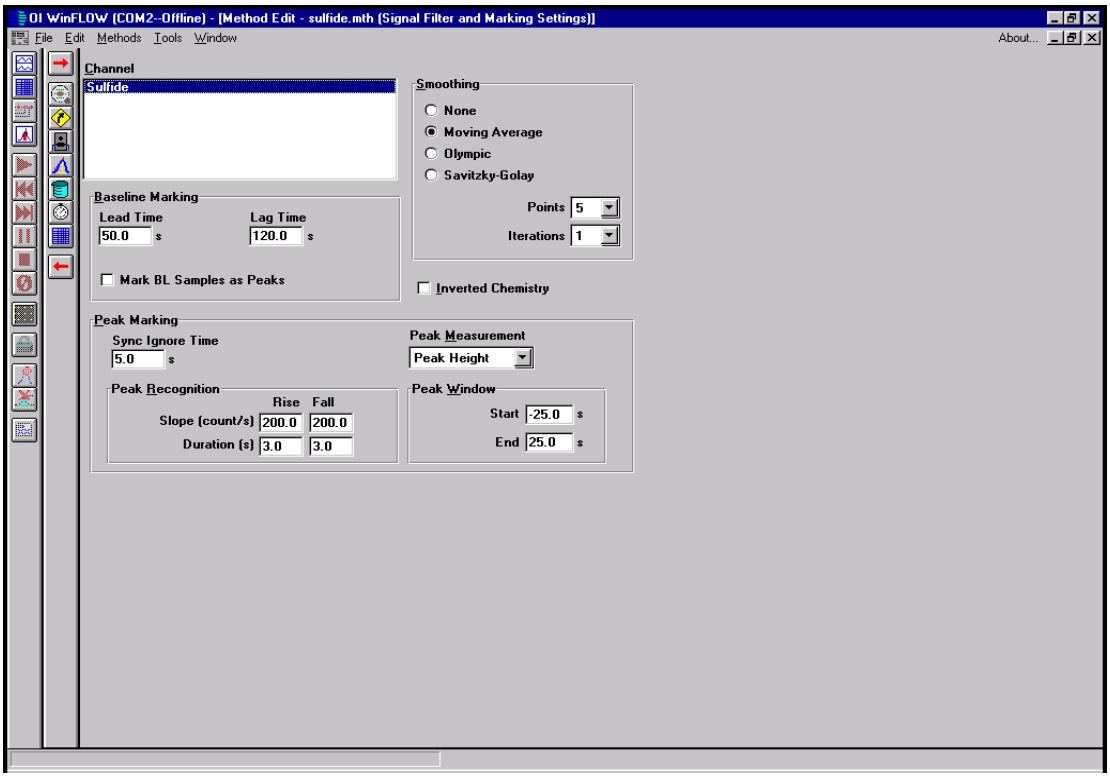


Figure 21. WinFLOW Method Editor—Signal Filter and Marking Settings

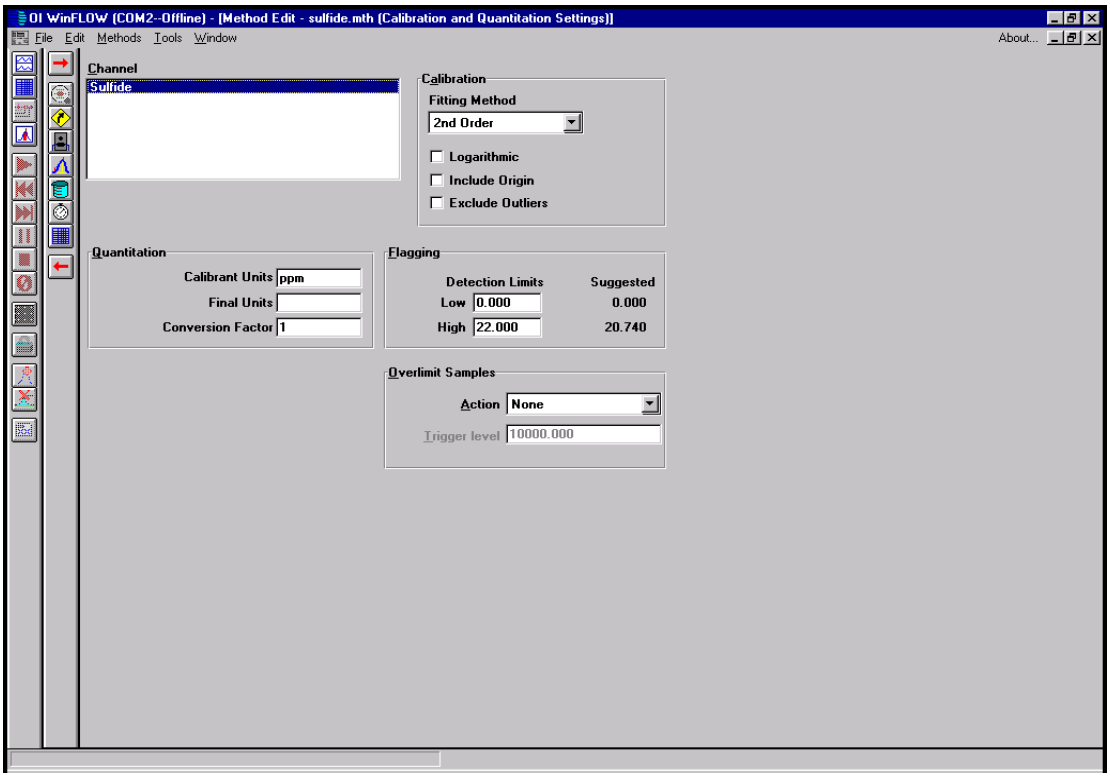


Figure 22. WinFLOW Method Editor—Calibration and Quantitation Settings

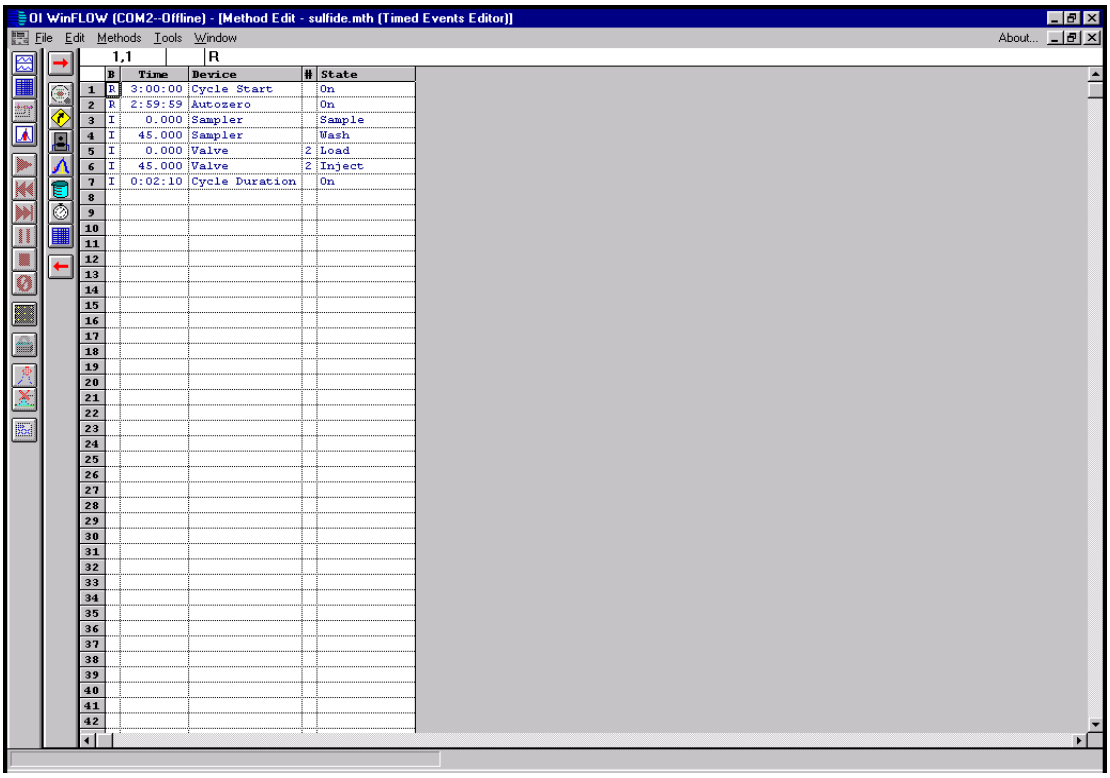


Figure 23. WinFLOW Method Editor—Timed Events Editor

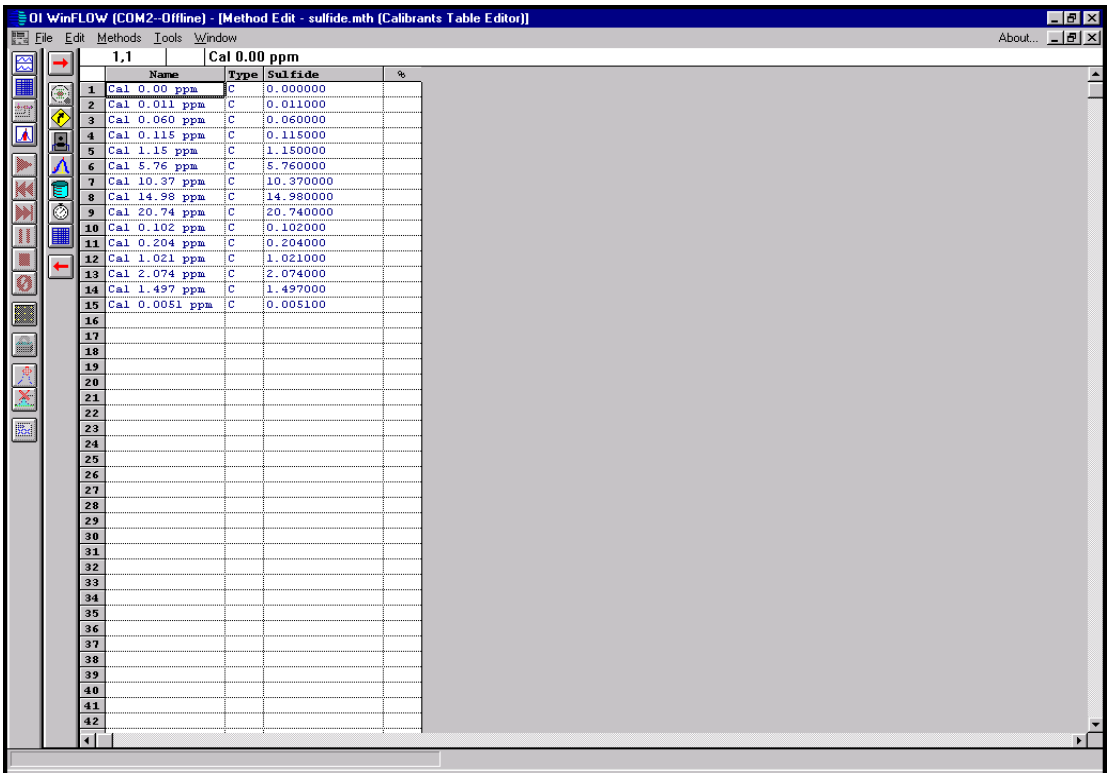


Figure 24. WinFLOW Method Editor—Calibrants Table Editor

Results were obtained under optimal operating conditions. Actual results may vary depending on sample introduction, cleanliness of sample containers, reagent purity, operator skill, and maintenance of instruments.

O·I·Analytical

P.O. Box 9010
College Station, Texas 77842-9010
Tel: (979) 690-1711 · FAX: (979) 690-0440

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