



## Ammonia in Soil and Plant Extracts by Segmented Flow Analysis (SFA)

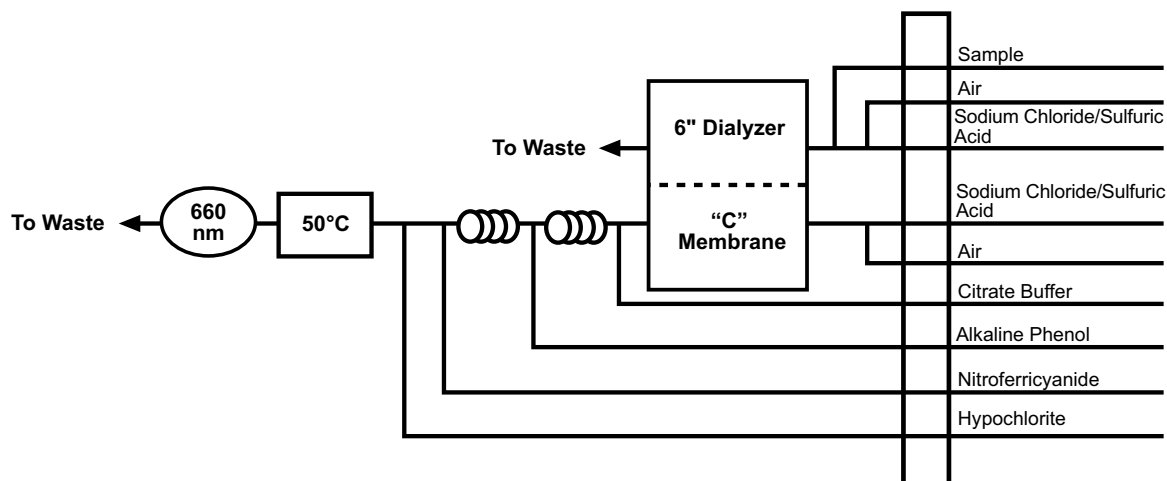
(Cartridge Part #A002617)

### 1.0 Scope and Application

- 1.1 This method is used for the determination of ammonia in soil and plant extracts.
- 1.2 The Method Detection Limit (MDL) of this method is 0.01 mg/L ammonia as nitrogen (N). The applicable range of the method is 0.20–250 mg/L ammonia as nitrogen. The range may be extended to analyze higher concentrations by sample dilution.

### 2.0 Summary of Method

- 2.1 Ammonia reacts with alkaline phenol and hypochlorite to form indophenol blue in an amount that is proportional to the ammonia concentration. The blue color is intensified with sodium nitroferricyanide, and the absorbance is measured at 660 nm (Reference 15.2).
- 2.2 The quality of the analysis is assured through reproducible calibration and testing of the Segmented Flow Analysis (SFA) system.
- 2.3 A general flow diagram of the SFA system is shown below (see Section 17.0 for a detailed flow diagram).



### 3.0 Definitions

Definitions for terms used in this method are provided in Section 16.0, "Glossary of Definitions and Purposes."

### 4.0 Interferences

- 4.1 Eliminate precipitation of calcium and magnesium hydroxides by adding sodium citrate.
- 4.2 Filter turbid samples prior to analysis.
- 4.3 Samples with background absorbance at the analytical wavelength may interfere.

### 5.0 Safety

- 5.1 The toxicity or carcinogenicity of each compound or reagent used in this method has not been fully established. Each chemical should be treated as a potential health hazard. Exposure to these chemicals should be reduced to the lowest possible level.
- 5.2 For reference purposes, a file of Material Safety Data Sheets (MSDS) for each chemical used in this method should be available to all personnel involved in this chemical analysis. The preparation of a formal safety plan is also advisable.
- 5.3 The following chemicals used in this method may be highly toxic or hazardous and should be handled with extreme caution at all times. Consult the appropriate MSDS before handling.
  - 5.3.1 Ammonium Sulfate,  $(\text{NH}_4)_2\text{SO}_4$  (FW 132.14)
  - 5.3.2 Phenol, solid or liquified, 88%,  $\text{C}_6\text{H}_5\text{OH}$  (FW 94.11)
  - 5.3.3 Sodium Chloride,  $\text{NaCl}$  (FW 58.44)
  - 5.3.4 Sodium Citrate Dihydrate,  $\text{C}_6\text{H}_5\text{Na}_3\text{O}_7 \cdot 2\text{H}_2\text{O}$  (FW 294.10)
  - 5.3.5 Sodium Hydroxide,  $\text{NaOH}$  (FW 40.00)
  - 5.3.6 Sodium Hypochlorite, 5.25% available chlorine (household bleach) (FW 74.44)
  - 5.3.7 Sodium Nitroferricyanide Dihydrate,  $\text{Na}_2\text{Fe}(\text{CN})_5\text{NO} \cdot 2\text{H}_2\text{O}$  (FW 297.95)
  - 5.3.8 Sulfuric Acid, concentrated,  $\text{H}_2\text{SO}_4$  (FW 98.08)
- 5.4 Unknown samples may be potentially hazardous and should be handled with extreme caution at all times.
- 5.5 Proper personal protective equipment (PPE) should be used when handling or working in the presence of chemicals.

- 5.6 This method does not address all safety issues associated with its use. The laboratory is responsible for maintaining a safe work environment and a current awareness file of OSHA regulations regarding the safe handling of the chemicals specified in this method.

## 6.0 Apparatus, Equipment, and Supplies

- 6.1 Segmented Flow Analysis (SFA) System (OI Analytical Flow Solution® IV) consisting of the following:
- 6.1.1 Model 502 Multichannel Peristaltic Pump
  - 6.1.2 Random Access (RA) Autosampler
  - 6.1.3 Expanded Range (ER) Photometric Detector with 5-mm path length flowcell and 660-nm optical filter
  - 6.1.4 Data Acquisition System (PC or Notebook PC) with WinFLOW™ software
  - 6.1.5 Ammonia/TKN in Soil and Plant Extracts Cartridge (Part #A002617)
- 6.2 Sampling equipment—Sample bottle, amber glass, with polytetrafluoroethylene (PTFE)-lined cap. Clean by washing with detergent and water, rinsing with two aliquots of reagent water, and drying by baking at 110°–150°C for a minimum of one hour.
- 6.3 Standard laboratory equipment including volumetric flasks, pipettes, syringes, etc. should all be cleaned, rinsed, and dried per bottle cleaning procedure in Section 6.2.

## 7.0 Reagents and Calibrants

- 7.1 Raw Materials
- 7.1.1 Ammonium Sulfate,  $(\text{NH}_4)_2\text{SO}_4$  (FW 132.14)
  - 7.1.2 Brij®-35, 30% w/v (Part #A21-0110-33)
  - 7.1.3 Deionized Water (ASTM Type I or II)
  - 7.1.4 Phenol, solid or liquified, 88%,  $\text{C}_6\text{H}_5\text{OH}$  (FW 94.11)
  - 7.1.5 Sodium Chloride, NaCl (FW 58.44)
  - 7.1.6 Sodium Citrate Dihydrate,  $\text{C}_6\text{H}_5\text{Na}_3\text{O}_7 \cdot 2\text{H}_2\text{O}$  (FW 294.10)
  - 7.1.7 Sodium Hydroxide, NaOH (FW 40.00)
  - 7.1.8 Sodium Hypochlorite, 5.25% available chlorine (household bleach) (FW 74.44)

7.1.9 Sodium Nitroferricyanide Dihydrate,  $\text{Na}_2\text{Fe}(\text{CN})_5\text{NO}\cdot 2\text{H}_2\text{O}$  (FW 297.95)

7.1.10 Sulfuric Acid, concentrated,  $\text{H}_2\text{SO}_4$  (FW 98.08)

## 7.2 Reagent Preparation

**Note:** For best results, filter and degas all reagents prior to use.

### 7.2.1 Reagent Water

7.2.1.1 Degassed and deionized reagent water can be prepared in one of the following manners:

7.2.1.1.1 Place distilled/deionized water under a strong vacuum for 15–20 minutes. Magnetic stirring or sonification will aid in the degassing process.

7.2.1.1.2 Purge distilled/deionized water with a stream of nitrogen gas (or other inert gas) through a glass frit for approximately 5 minutes.

7.2.1.1.3 Boil distilled/deionized water in an Erlenmeyer flask for 15–20 minutes. Remove the flask from the heat source, cover it with an inverted beaker, and allow it to cool to room temperature.

7.2.1.2 After preparing the degassed reagent water, store the reagent water in a tightly sealed container to protect it from reabsorption of atmospheric gases. For best results, store degassed reagent water under a slight vacuum when not in use.

### 7.2.2 Start-up Solution (1 L)

7.2.2.1 Add 2 mL of Brij-35 to approximately 800 mL of reagent water (Section 7.2.1) in a 1-L volumetric flask.

7.2.2.2 Dilute to 1,000 mL with reagent water and mix gently.

### 7.2.3 Sodium Citrate Solution (200 mL)

7.2.3.1 Dissolve 28 g of sodium citrate dihydrate in approximately 150 mL of reagent water in a 200-mL volumetric flask.

7.2.3.2 Add 6 g of sodium hydroxide and mix to dissolve.

7.2.3.3 Dilute to 200 mL with reagent water and mix well.

**Warning:** Mixing sodium hydroxide with water produces a great amount of heat. Take appropriate precautions.

**Note:** Store this solution at room temperature.

#### 7.2.4 Sodium Nitroferricyanide Solution (200 mL)

7.2.4.1 Dissolve 0.1 g of sodium nitroferricyanide dihydrate in approximately 150 mL of reagent water in a 200-mL volumetric flask.

7.2.4.2 Dilute to 200 mL with reagent water and mix well.

**Note:** Store in an amber bottle at room temperature.

#### 7.2.5 Alkaline Phenol Solution (200 mL)

7.2.5.1 Add 7.35 mL of liquified (or 8.3 g of solid) phenol and 4 g of sodium hydroxide to approximately 150 mL of reagent water in a 200-mL volumetric flask.

7.2.5.2 Dilute to 200 mL with reagent water and mix well.

**Warning:** Mixing sodium hydroxide with water produces a great amount of heat. Take appropriate precautions.

**Note:** Prepare this solution fresh daily.

#### 7.2.6 Working Hypochlorite Solution (200 mL)

7.2.6.1 Add 12 mL of sodium hypochlorite (5.25% available chlorine) to approximately 150 mL of reagent water in a 200-mL volumetric flask.

7.2.6.2 Dilute to 200 mL with reagent water and mix well.

**Note:** Prepare this solution fresh daily.

#### 7.2.7 Sodium Chloride/Sulfuric Acid Solution (1 L)

7.2.7.1 Dissolve 100 g of sodium chloride in approximately 800 mL of reagent water in a 1-L volumetric flask.

7.2.7.2 Add 7.5 mL of concentrated sulfuric acid.

7.2.7.3 Dilute to 1,000 mL with reagent water and mix well.

7.2.7.4 Add 1 mL of Brij-35 and mix gently.

**Warning:** Mixing sulfuric acid with water produces a great amount of heat. Take appropriate precautions.

**Note:** Prepare this solution fresh daily.

### 7.2.8 1 N Sodium Hydroxide Solution (1 L)

7.2.8.1 Dissolve 40 g of sodium hydroxide in approximately 800 mL of reagent water in a 1-L volumetric flask.

7.2.8.2 Dilute to 1,000 mL with reagent water and mix well.

**Warning:** Mixing sodium hydroxide with water produces a great amount of heat. Take appropriate precautions.

### 7.2.9 Sampler Wash—Reagent Water

## 7.3 Calibrant Preparation

### 7.3.1 Stock Calibrant 1,000 mg/L Ammonia Nitrogen (1 L)

7.3.1.1 Dissolve 4.717 g of ammonium sulfate (dried at 100°C) in approximately 800 mL of reagent water in a 1-L volumetric flask.

7.3.1.2 Dilute to 1,000 mL with reagent water and mix well.

**Note:** Store at 4°C. If stored properly, this solution is stable for 4–6 weeks.

### 7.3.2 Intermediate Calibrant 100 mg/L Ammonia Nitrogen (100 mL)

7.3.2.1 Use a volumetric pipet to add 10 mL of stock calibrant (Section 7.3.1) to approximately 80 mL of reagent water in a 100-mL volumetric flask.

7.3.2.2 Dilute to 100 mL with reagent water and mix well.

**Note:** Prepare this solution fresh daily.

### 7.3.3 Working Calibrants (100 mL)

7.3.3.1 Add the designated volumes of stock calibrant (see Equation 1) to the required number of 100-mL volumetric flasks that each contain approximately 80 mL of reagent water.

7.3.3.2 Dilute each solution to the mark with reagent water and mix well.

**Note:** Prepare working calibrants fresh daily.

## EQUATION 1

$$C_1 V_1 = C_2 V_2$$

Where:

$C_1$  = Concentration (in mg/L) of stock solution (or calibrant)

$V_1$  = Volume (in L) of stock solution (or calibrant) to be used

$C_2$  = Desired concentration (in mg/L) of working calibrant to be prepared

$V_2$  = Final volume (in L) of working calibrant to be prepared

By solving this equation for the volume of stock solution to be used ( $V_1$ ), the following equation is obtained:

$$V_1 = \frac{C_2 V_2}{C_1}$$

Since the desired concentration ( $C_2$ ), the final volume ( $V_2$ ), and the concentration of the stock solution ( $C_1$ ) are all known for any given calibrant concentration in a defined volume, the volume of stock solution to be used ( $V_1$ ) is easily calculated.

7.3.3.3 Calibrants covering the entire range of this analysis can be prepared from the following tables.

Final Concentration (mg/L)	Vol. of Calibrant (mL)	Conc. of Calibrant (mg/L)	Final Volume (mL)
0.20	0.20	100	100
0.50	0.50	100	100
1.0	1.0	100	100

Final Concentration (mg/L)	Vol. of Calibrant (mL)	Conc. of Calibrant (mg/L)	Final Volume (mL)
5.0	0.50	1,000	100
10	1.0	1,000	100
50	5.0	1,000	100
100	10	1,000	100
250	25	1,000	100

## 8.0 Sample Collection, Preservation, and Storage

- 8.1 Samples should be collected in plastic or glass bottles that have been thoroughly cleaned and rinsed with reagent water (Section 7.2.1).
- 8.2 The volume of sample collected should be sufficient to ensure that a representative sample is obtained, replicate analysis is possible, and waste disposal is minimized.
- 8.3 Determine ammonia nitrogen in unpreserved samples as soon as possible to eliminate loss of analyte.
- 8.4 Preserve and store samples with 2 mL of concentrated sulfuric acid per liter of sample and refrigerate at 4°C.
- 8.5 Holding time for preserved samples is 28 days from the time of collection (Reference 15.3).

## 9.0 Quality Control

**Note:** The following QC procedures are provided for reference purposes only and are not a substitute for any QC procedures that may be required for regulatory compliance.

- 9.1 It is recommended that each laboratory that uses this method operate a formal quality control program. The minimum requirements of such a program should consist of an initial demonstration of laboratory capability and the periodic analysis of Laboratory Control Samples (LCSs) and Matrix Spike/Matrix Spike Duplicates (MS/MSDs) as a continuing check on performance. Laboratory performance should be compared to established performance criteria to determine if the results of the analyses meet the performance characteristics of the method.
- 9.2 Method Detection Limit (MDL)—To establish the ability to detect ammonia at low levels, the analyst should determine the MDL using the apparatus, reagents, and calibrants that will be used in the practice of this method. An MDL less than or equal to the MDL listed in Section 1.2 should be achieved prior to practice of this method.
  - 9.2.1 An MDL is calculated by analyzing a matrix spike at a concentration of two to three times the expected detection limit of the analyzer. Seven consecutive replicate analyses of this matrix spike should be analyzed, and the MDL should be calculated using Equation 2.



## EQUATION 2

$$MDL = (t) \times (S)$$

Where:

*t* = Student's *t* value for a 99% confidence level and a standard deviation estimate with *n*–1 degrees of freedom (*t* = 3.14 for seven replicates)

*S* = Standard deviation of the replicate analyses

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- 9.2.2 It is recommended that the MDL be calculated after every six months of operation, when a new operator begins work, or whenever there is any significant change in the instrument response.
- 9.3 Analyses of MS/MSD samples are required to demonstrate method accuracy and precision and to monitor matrix interferences (interferences caused by the sample matrix).
- 9.3.1 Matrix Spike/Matrix Spike Duplicate (MS/MSD)—The laboratory should spike, in duplicate, a minimum of 10% of all samples (one sample in duplicate in each batch of 10 samples) from a given sampling site.
- 9.3.2 The concentration of the spike in the sample shall be determined as follows:
- 9.3.2.1 If, as in compliance monitoring, the concentration of ammonia in the sample is being checked against a regulatory concentration limit, the spiking level shall be at that limit.
- 9.3.2.2 If the concentration of ammonia in a sample is not being checked against a limit, the spike shall be at the concentration of the LCS or at least four times greater than the MDL.
- 9.4 Analyses of Laboratory Reagent Blanks (LRBs) are required to demonstrate freedom from contamination and that the compounds of interest and interfering compounds have not been carried over from a previous analysis.
- 9.5 As part of the QC program for the laboratory, method precision and accuracy for samples should be assessed and records should be maintained.
- 9.5.1 An LCS should be analyzed with every sample batch, and the mean (*m*) and the standard deviation (*S*) should be recorded. After multiple analyses, the mean should be plotted with limits of *m*+2*S* and *m*–2*S*. The mean and the limits should be recalculated after every 5–10 new measurements.
- 9.5.2 If the LCS measurement falls outside the range calculated in Section 9.5.1, then the problem should be addressed, and that sample batch should be reanalyzed if necessary.
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- 9.6 Reference Sample—To demonstrate that the analytical system is in control, the laboratory may wish to periodically test an external reference sample, such as a Standard Reference Material (SRM) available from the National Institute of Standards and Technology (NIST). Corrective action should be taken if the measured concentration significantly differs from the stated concentration.

## 10.0 Configuration and Start-up

### 10.1 Instrument Configuration

- 10.1.1 Configure the OI Analytical Flow Solution IV Analyzer according to the Operator's Manual and verify that each module is properly powered on.
- 10.1.2 Verify that the Ammonia/TKN in Soil and Plant Extracts Cartridge (Part #A002617) is configured as illustrated in the flow diagram shown in Section 17.0.
- 10.1.3 Connect the appropriate pump tubes to the cartridge and to their appropriate reagent containers according to the flow diagram.

### 10.2 Instrument Stabilization

- 10.2.1 Connect the reagent pump tubes to a reagent bottle containing the start-up solution (Section 7.2.2). Start the pump at 40% speed, allowing the start-up solution to flow through the entire system.
- 10.2.2 Verify that the flowcell of each detector is purged of all bubbles and that the flow is stable and free from surging before proceeding.

### 10.3 Baseline Verification

- 10.3.1 Create and save a Method in WinFLOW. Refer to the WinFLOW Operator's Manual (Reference 15.4) for help on creating a Method.
- 10.3.2 Create and save a Sample Table in WinFLOW that will be used to generate a calibration curve using at least three calibrants that cover the full range of expected concentrations in the samples to be analyzed. This Sample Table should also be used to analyze all necessary QC samples as well as the analytical batch of samples to be analyzed. For help on creating a Sample Table, refer to the WinFLOW Operator's Manual (Reference 15.4).
- 10.3.3 Select **Collect Data** in the WinFLOW main window, enter the user's identification, select the appropriate Method and Sample Table, and begin to collect baseline data. Very sharp fluctuations in the baseline and/or consistent drifting are typically signs of bubbles in the flowcell. The flowcell must be free of bubbles prior to beginning analysis.

### 10.4 Calibration and Standardization

- 10.4.1 Prepare a series of at least three working calibrants using the stock solutions (Section 7.3) according to Equation 1, covering the desired analysis range.

- 10.4.2 Place the calibrants in the autosampler in order of increasing concentration. Each calibrant should be analyzed according to the analytical procedures in Section 11.0. A calibration curve will be calculated by the WinFLOW software.
- 10.4.3 Acceptance or control limits for the calibration results should be established using the difference between the measured value of each calibrant and the corresponding “true” concentration.
- 10.4.4 Each calibration curve should be verified by analysis of a Laboratory Control Sample (LCS, Section 9.5). Using WinFLOW software, calibration, verification, and sample analysis may be performed in one continuous analysis.

## 11.0 Procedure

**Note:** Pay special attention to Sections 11.1.2 and 11.2.6 to avoid precipitation of the alkaline phenol solution.

### 11.1 Analysis

- 11.1.1 Set the temperature of the heater to 50°C. Begin pump flow with the start-up solution (Section 7.2.2). Once the heater unit has reached 50°C, verify a stable baseline (Section 10.3).
- 11.1.2 Place the alkaline phenol line in reagent water while pumping all the other reagents for at least 5 minutes. Then put the alkaline phenol line into its reagent bottle and pump the alkaline phenol solution (Section 7.2.5) through the system.
- 11.1.3 If precipitate forms after the addition of the alkaline phenol solution, it was added too soon or the pH of the reagent stream was too low.
  - 11.1.3.1 Immediately stop the pump and disconnect the manifold from the debubbler.
  - 11.1.3.2 Place all reagent lines in 1 N sodium hydroxide (Section 7.2.8).
  - 11.1.3.3 Turn on the pump and flush the system for at least 10 minutes or until all of the precipitate has been removed from the system.
  - 11.1.3.4 Rinse with reagent water for 5 minutes. Return the reagent lines to their bottles, except the alkaline phenol. Repeat Section 11.1.2.
- 11.1.4 When the system has stabilized, allow to pump at least 10–15 minutes and verify that there are no bubbles in the flowcell. Obtain a stable baseline at 660 nm and autozero the baseline before beginning the analysis.
- 11.1.5 Load the sampler tray with calibrants, blanks, samples, and QC samples.

**Note:** The matrix of the working standards, blanks, and QC samples should match that of the samples being analyzed.

- 11.1.6 Using the Method and Sample Table created for the analytical batch to be analyzed and with the baseline verified to be stable, begin the analysis by selecting the “Fast Forward” button on the left side of the Data Analysis window in WinFLOW. This will initiate the sequential analysis of samples as defined in the Sample Table.
- 11.1.7 When the analysis is complete, remove the alkaline phenol line from its reagent bottle and place it in reagent water. Pump the other reagents for at least 5 minutes. Then place the other lines in reagent water and pump for at least 10–15 minutes. Stop the pump, release the tension on all pump tubes, and power off the system.

## 11.2 Operating Notes

- 11.2.1 If a stable baseline on reagents cannot be achieved, investigate the following:

- 11.2.1.1 Filter the working reagents. Make sure the reagent containers are free from dust, debris, or other contamination.
- 11.2.1.2 Disconnect the debubbler from the flowcell by disconnecting the transmission tubing from the debubbler. Use a syringe fitted with a union to flush the flowcell with start-up solution (Section 7.2.2) followed by reagent water. Reconnect the debubbler.
- 11.2.1.3 Clean the cartridge with 1 N sodium hydroxide (Section 7.2.8). Remove the alkaline phenol line to reagent water for 5 minutes before placing all of the reagent lines in the 1 N sodium hydroxide to prevent precipitation in the cartridge.

- 11.2.2 If poor peak shape of carryover is observed, investigate the following:

- 11.2.2.1 Verify that the bubbles from the sampler travel smoothly through the tubing to the cartridge. If not, replace the pump tube and the transmission tubing.
- 11.2.2.2 Verify that the flow through the cartridge is smooth and consistent. Check for loose fittings.
- 11.2.2.3 Remake all reagents, including the stock reagents. The working hypochlorite solution (Section 7.2.6) must be fresh.

- 11.2.3 If high background or blank values are observed, carefully evaluate the source of water, the cleaning procedure for glassware, and the laboratory environment.

- 11.2.4 If poor sensitivity or nonlinearity is observed, investigate the following:

- 11.2.4.1 Check for excessive pump tube wear or a flow restriction. Replace pump tubes and verify that all lines are pumping freely.
- 11.2.4.2 Verify that the heater unit is on and that the temperature is not too low.

11.2.4.3 Remake all reagents, including the stock reagents. The working hypochlorite solution (Section 7.2.6) must be fresh.

11.2.5 The pH of the stream from the flowcell should be approximately 13.

## 12.0 Data Analysis and Calculations

12.1 The calibration curve allows for accurate quantitation of the concentration in each sample.

12.2 WinFLOW software reports the concentration of each sample relative to the calibration curve.

## 13.0 Method Performance

Range:	0.20–250 mg/L
Throughput:	30 samples/hour
Precision:	
0.20 mg/L	<3% RSD
250 mg/L	<2% RSD
Method Detection Limit (MDL):	0.01 mg/L

## 14.0 Pollution Prevention and Waste Management

14.1 It is the laboratory's responsibility to comply with all federal, state, and local regulations governing waste management, particularly the hazardous waste identification rules and land-disposal restrictions. In addition, it is the laboratory's responsibility to protect air, water, and land resources by minimizing and controlling all releases from fume hoods and bench operations. Also, compliance is required with any sewage discharge permits and regulations.

14.2 For further information on waste management, consult Section 13.6 of *Less is Better: Laboratory Chemical Management for Waste Reduction* (Reference 15.1).

## 15.0 References

- 15.1 *Less is Better: Laboratory Chemical Management for Waste Reduction*. Available from the American Chemical Society, Department of Government Regulations and Science Policy, 1155 16<sup>th</sup> Street, NW, Washington, DC, 20036.
- 15.2 Nitrogen, Kjeldahl, Total. *Methods for Chemical Analysis of Water and Wastewater*; EPA-600/4-79-020; U.S. Environmental Protection Agency, Office of Research and Development, Environmental Monitoring and Support Laboratory: Cincinnati, OH, 1984; Method 351.2.
- 15.3 Sample Preservation. *Methods for Chemical Analysis of Water and Wastes*; EPA-600/4-79-020; U.S. Environmental Protection Agency, Office of Research and Development, Environmental Monitoring and Support Laboratory: Cincinnati, OH, 1984; xvii.

15.4 *Standard Methods for the Examination of Water and Wastewater*, 20th ed.; American Public Health Association: Washington, D.C., 1998.

15.5 WinFLOW Software and Operator's Manual (Part #A002877). Available from OI Analytical, P.O. Box 9010, College Station, TX, 77842-9010.

## 16.0 Glossary of Definitions and Purposes

The definitions and purposes are specific to this method but have been conformed to common usage as much as possible.

### 16.1 Units of weights and measures and their abbreviations

#### 16.1.1 Symbols

°C	degrees Celsius
%	percent
±	plus or minus
≥	greater than or equal to
≤	less than or equal to

#### 16.1.2 Alphabetical characters

g	gram
L	liter
mg	milligram
mg/L	milligram per liter
µg	microgram
µg/L	microgram per liter
mL	milliliter
ppm	parts per million
ppb	parts per billion
M	molar solution
N	normal solution

### 16.2 Definitions

16.2.1 **Laboratory Control Sample (LCS)**—An aliquot of LRB to which a quantity of the analyte of interest is added in the laboratory. The LCS is analyzed like a sample. Its purpose is to determine whether the methodology is in control and whether the laboratory is capable of making accurate and precise measurements.

16.2.2 **Laboratory Reagent Blank (LRB)**—An aliquot of reagent water and other blank matrix that is treated like a sample, including exposure to all glassware, equipment, and reagents that are used with other samples. The LRB is used to determine if the method analyte or other interferences are present in the laboratory environment, reagents, or apparatus.

- 16.2.3 Matrix Spike/Matrix Spike Duplicate (MS/MSD)—An aliquot of an environmental sample to which a quantity of the method analyte is added in the laboratory. The MS/MSD is analyzed like a sample. Its purpose is to determine whether the sample matrix contributes bias to the analytical results. The background concentration of the analyte in the sample matrix must be determined in a separate aliquot, and the measured values in the MS/MSD must be corrected for the background concentration.
- 16.2.4 Method Detection Limit (MDL)—The minimum concentration of a substance that can be measured and reported with 99% confidence that the analyte concentration is greater than zero.

## 17.0 Figures

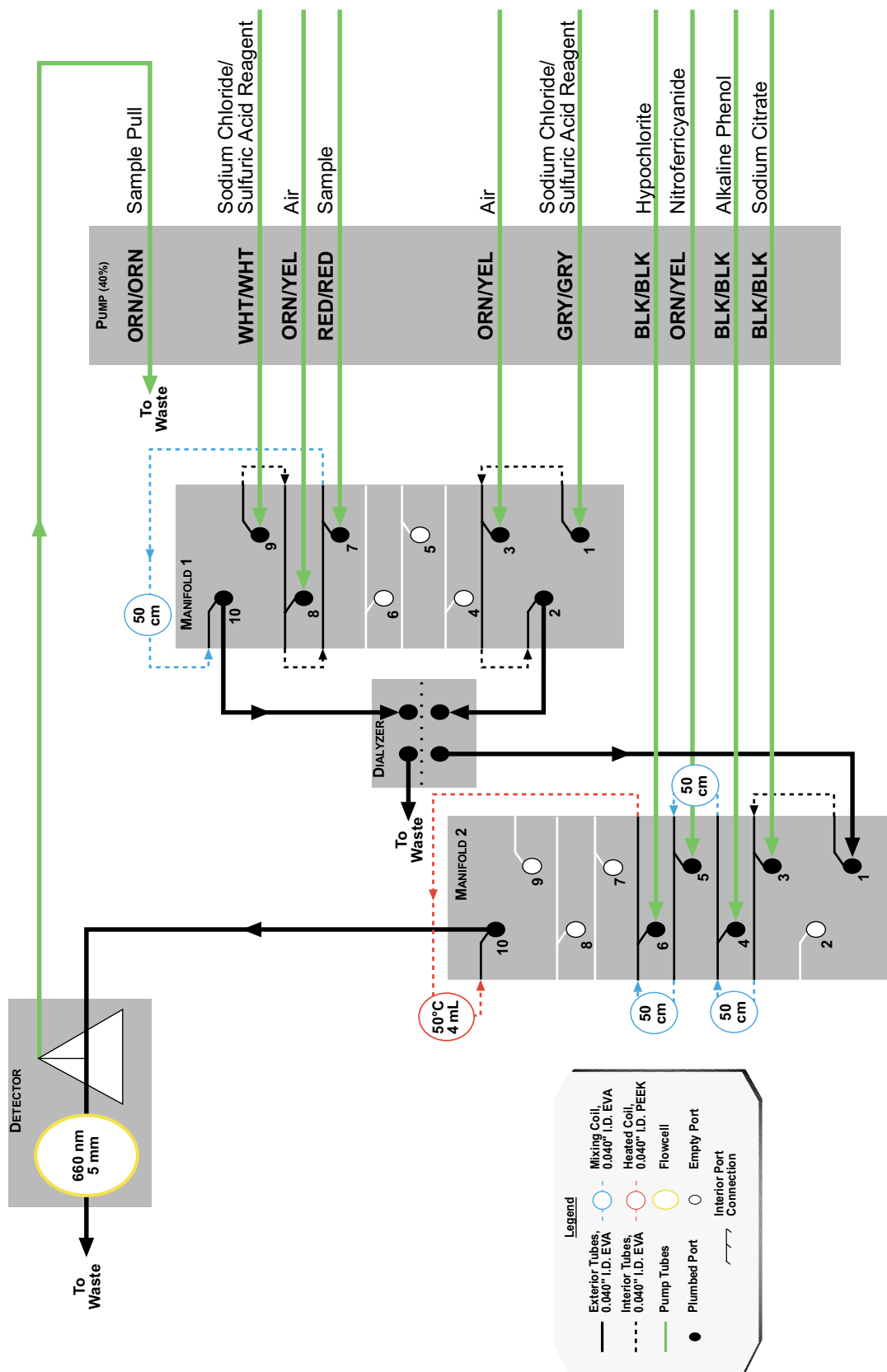


Figure 1. Detailed Flow Diagram for Ammonia/TKN in Soil and Plant Extracts by SFA on a Flow Solution IV, Cartridge Part #A002617



Results were obtained under optimal operating conditions. Actual results may vary depending on sample introduction, cleanliness of sample containers, reagent purity, operator skill, and maintenance of instruments.

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P.O. Box 9010  
College Station, Texas 77842-9010  
Tel: (979) 690-1711 · FAX: (979) 690-0440