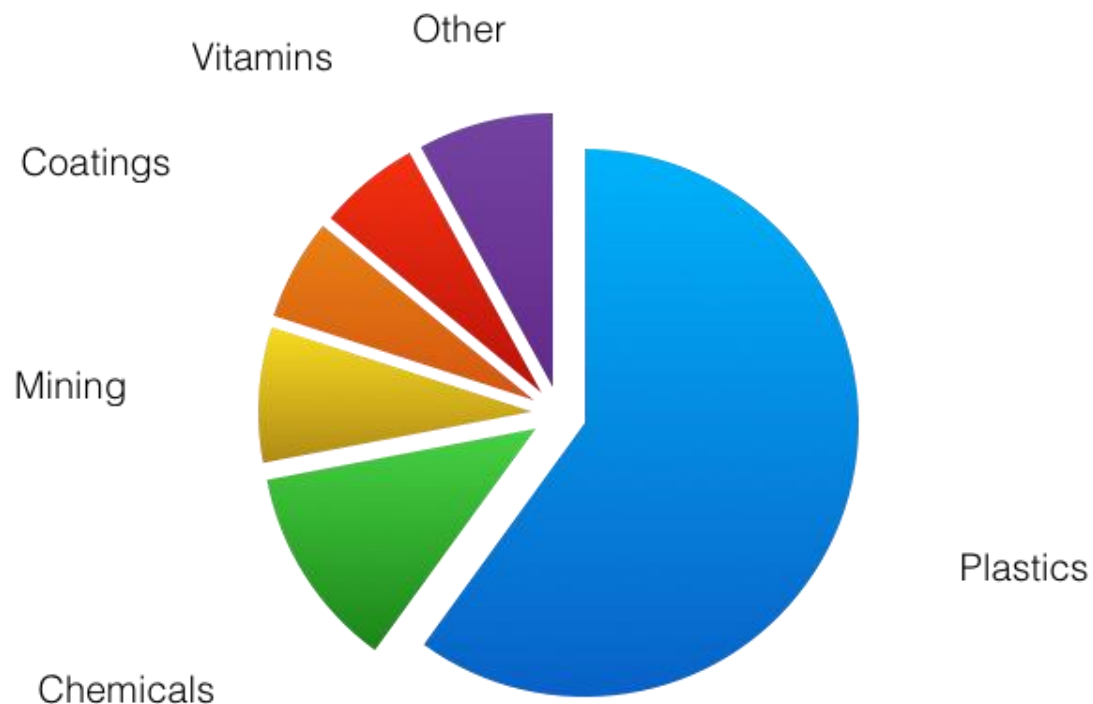


# **Cyanide Analysis:**

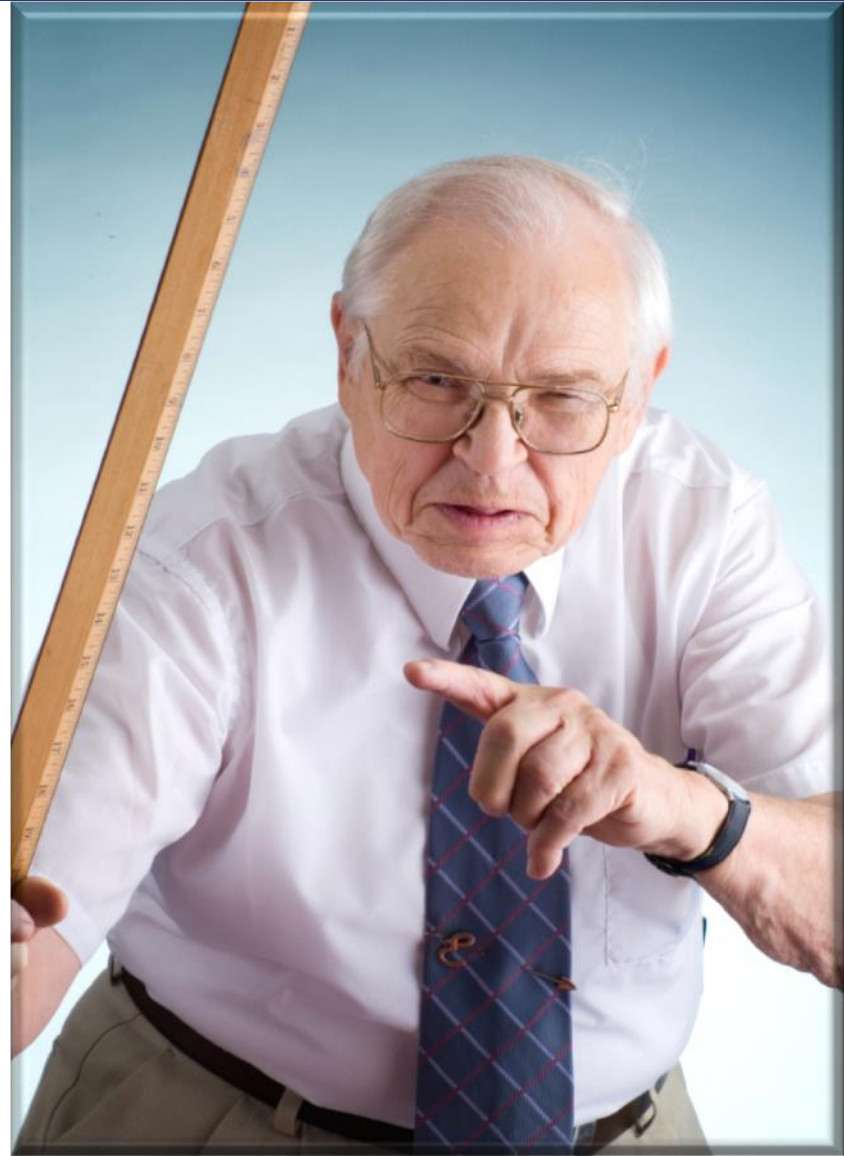
## **Cyanide Chemistry, Methodology, Interferences, Sample Handling and Regulatory Updates**

# Distribution of the Industrial Uses of CN



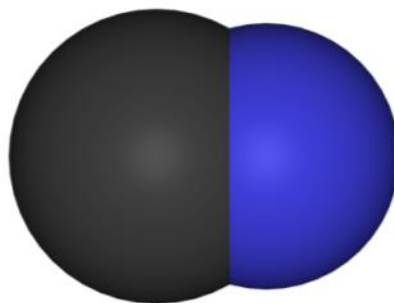
# Who is measuring cyanide?

- NPDES
- Pretreatment
- SDWA
- Industrial hygiene
- Foods
- Beverages
- Mining
- Manufacturing

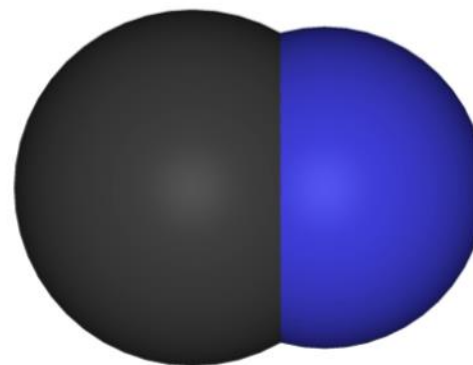


# A generalized summary of cyanide and its metal – cyanide species

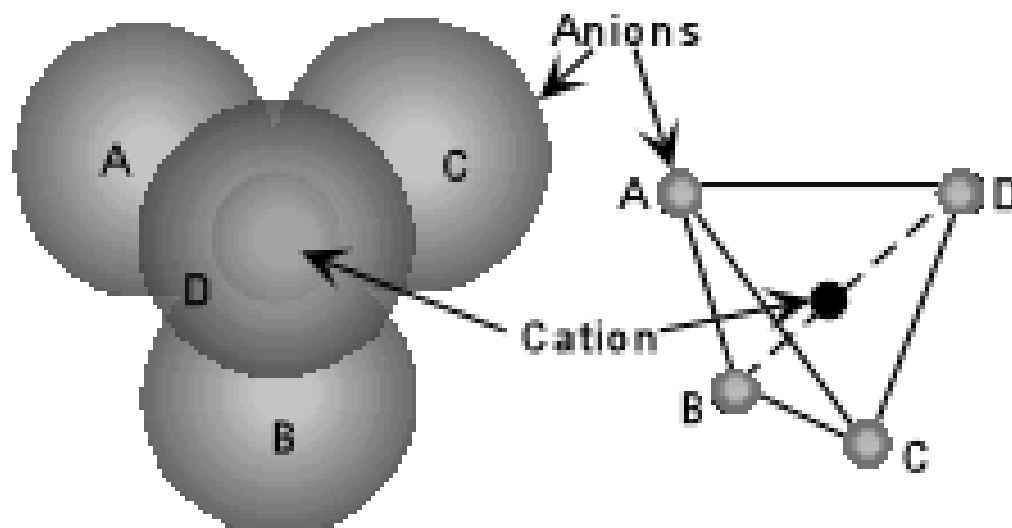
- Transition metals - strong bonds
- Alkali metals - ionic bonds



**Free Cyanide is the CN ion and HCN, generate HCN at pH 6**

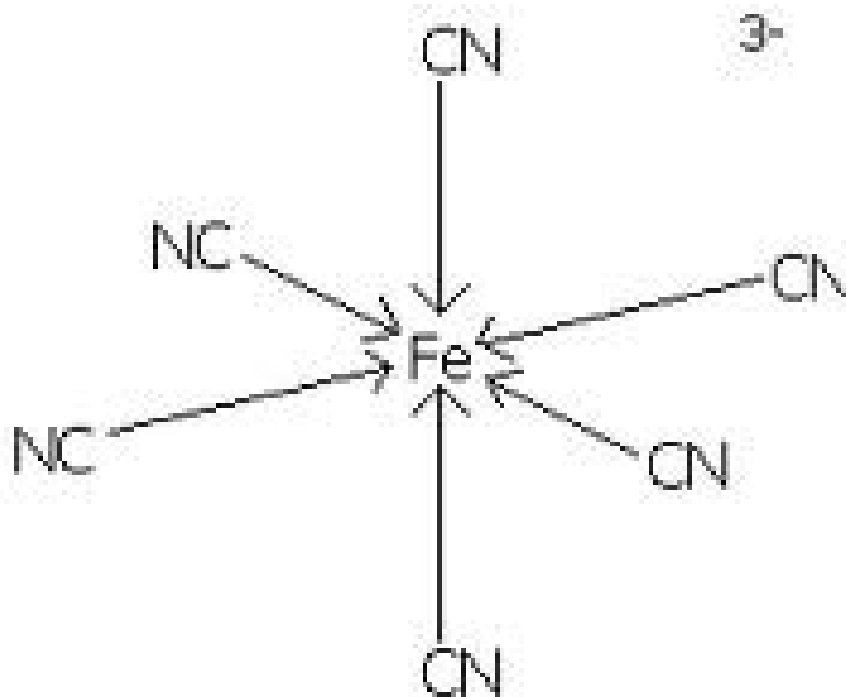


# Metal Complexes require acid to generate HCN



**Zn, Cd, Cu, Ni, Ag**

# Strong Metal Complexes are stable in acid solution



# Iron Cyanide Complexes are very stable in the environment





# Toxicity of Cyanide Complexes is related to its ability to produce HCN

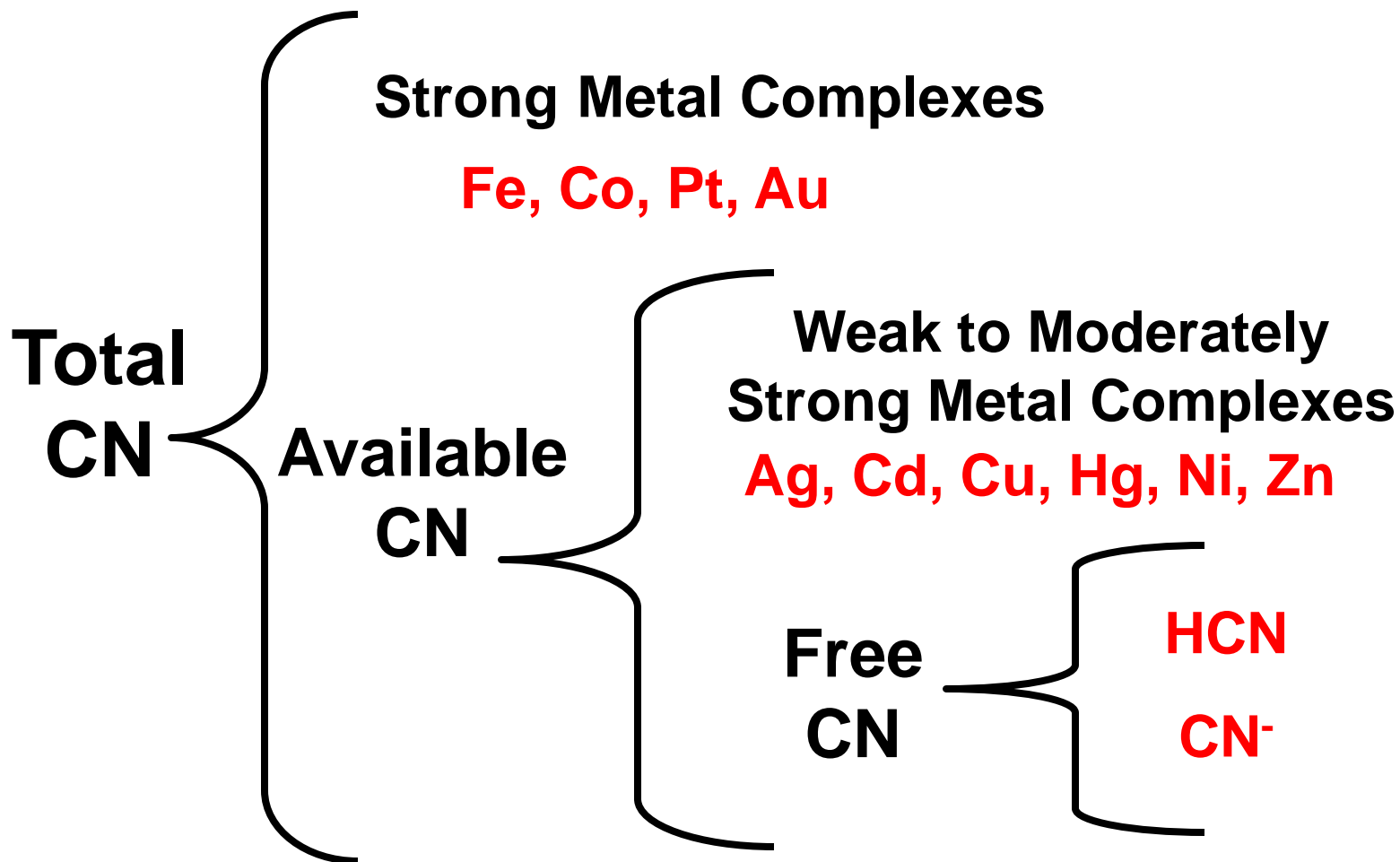


**Weak – Free Cyanide**

**Moderate – Available Cyanide**

**Strong – Total Cyanide**

# Cyanide methods measure the various cyanide “species”



# Cyanide methods measure the various cyanide “species”

## Total Cyanide

Fe

Co

## Available Cyanide

Ag

Hg

Ni

Cu

Zn

Cd

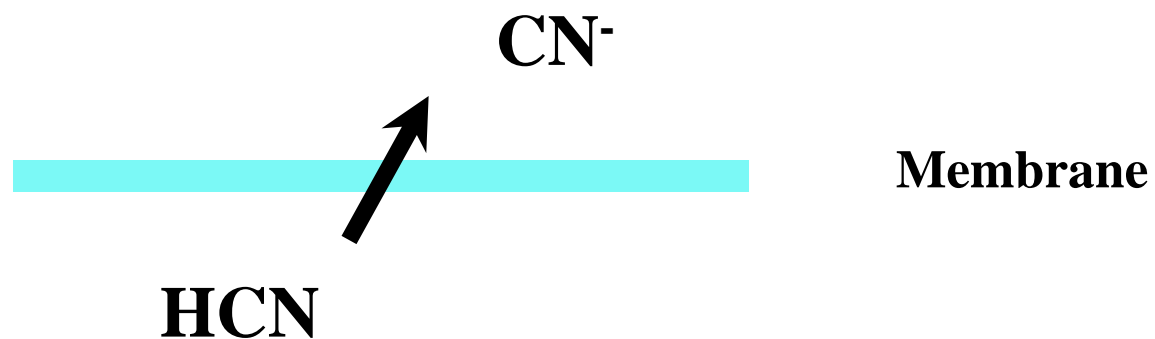
## Free Cyanide

CN<sup>-</sup>

HCN

# How Do You Measure Cyanide?

# Gas diffusion - Amperometry



**Sulfide > 50 ppm**

# Titration by silver ion



S-2

Cl-

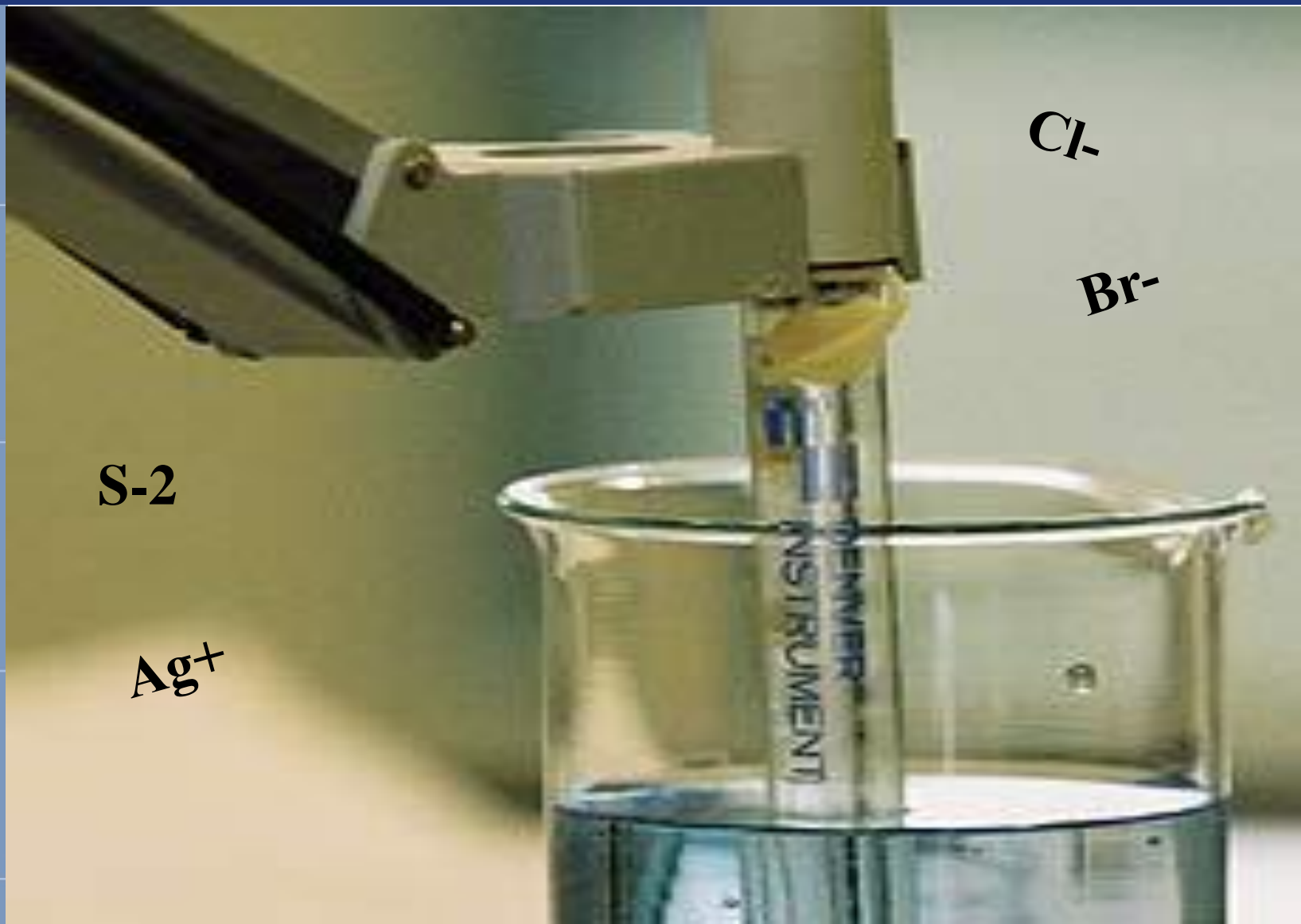
CN

NH<sub>3</sub>

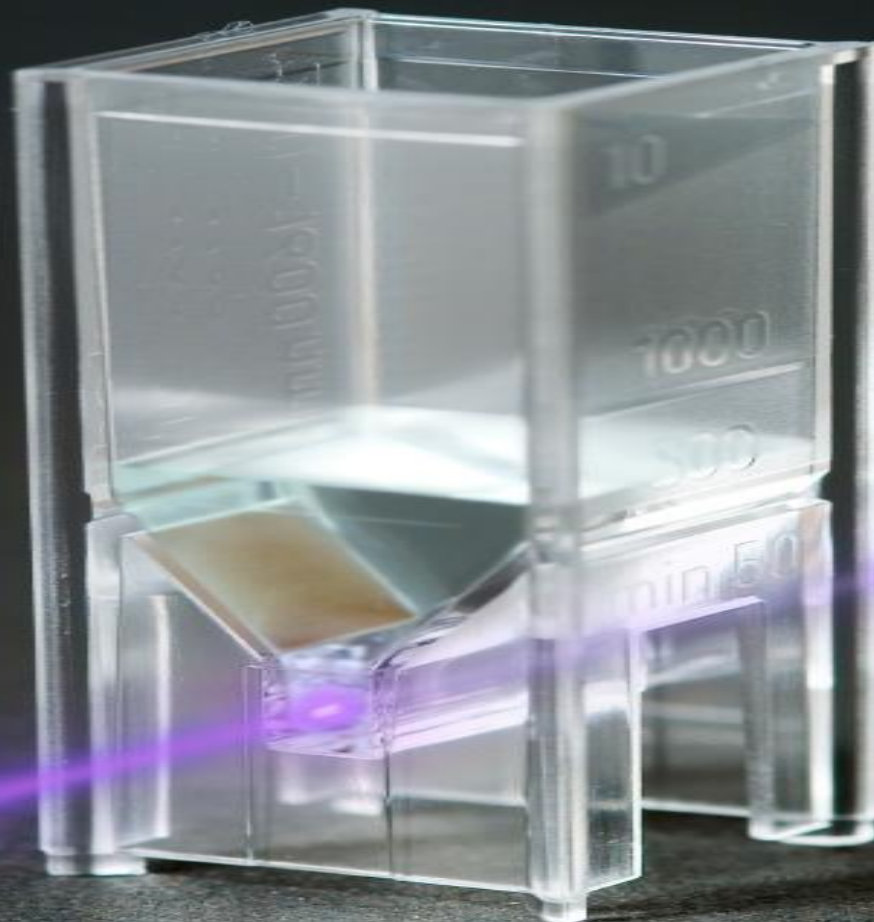
PO<sub>4</sub>

NO<sub>3</sub>

# Ion Selective Electrode (ISE)



# Colorimetric methods



$S^{2-}$

$SCN^{-}$

$SO_3^{-}$

Color

TDS

Turbidity

CN

$NH_3$

$PO_4$

$NO_3$



**Manual “distillation” is used to dissociate as HCN**



**Macro Distillation**



**MIDI  
Distillations**

# Distillation can be automated on a continuous flow analyzer



**Distillation and  
condenser**

**Distillate**



# Cyanide Methods

# Cyanide methods require separation of CN from matrix

- Separated from interferences, cyanide measurement is no different than running standards.



# APPROVED METHODS

<b>Free CN</b>	GD-Amperometry	ASTM D7237
<b>WAD CN</b>	Distillation/Colorimetry	ASTM D2036 or SM4500
<b>CATC</b>	Distillation/Colorimetry	ASTM D2036 or SM4500
<b>Available CN</b>	GD-Amperometry	OIA 1677 or ASTM D6888



# Unlike colorimetry, GD amperometry is easy to visualize

- $\text{CN}^- + \text{H}^+ \rightarrow \text{HCN}$
- $\text{HCN} + \text{OH}^- \rightarrow \text{CN}^- + \text{H}_2\text{O}$
- $\text{Ag} + 2\text{CN}^- \rightarrow \text{Ag}(\text{CN})_2^- + \text{e}^-$



**measure**

CN

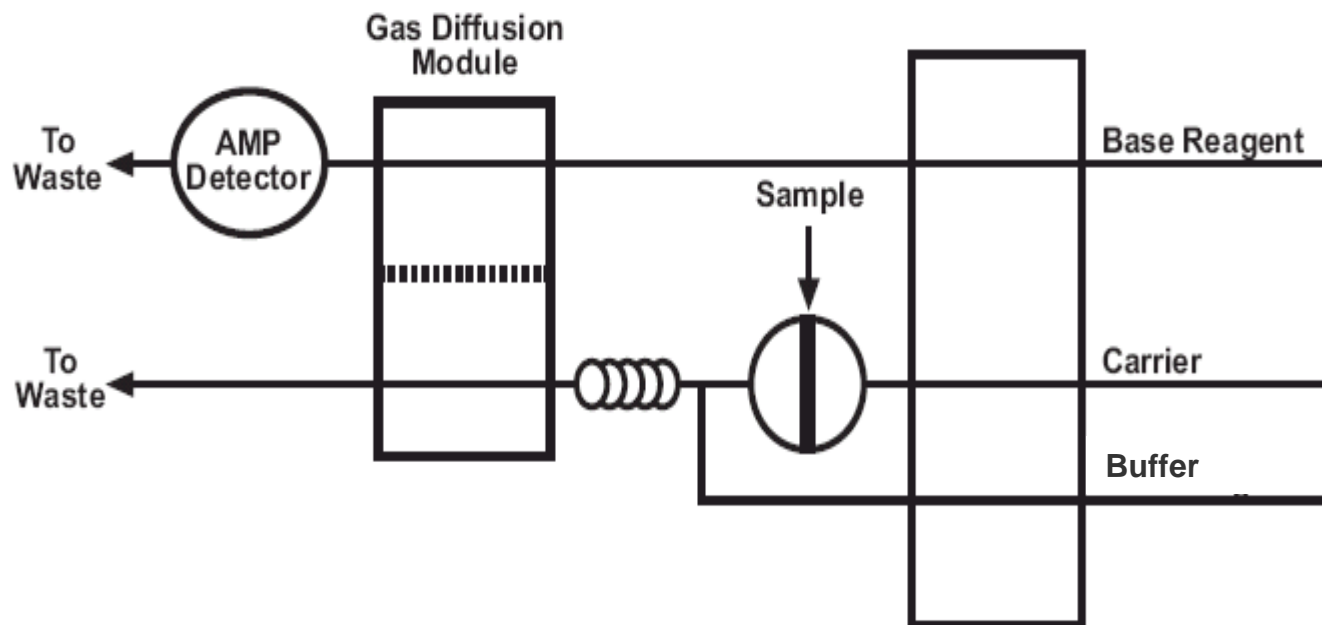
NH<sub>3</sub>

PO<sub>4</sub>

NO<sub>3</sub>

# Aquatic Free Cyanide Analysis

# ASTM D7237-06 can be run on a OIA1677 CNSolution analyzer



**Same Cartridge as OIA 1677**



# WAD Cyanide methods measure “available cyanide”

Method Number	Description	Measurement
SM 4500-CN I	Buffered pH 4.5 manual Distillation	Colorimetry
ASTM D 2036	Buffered pH 4.5 manual distillation	Colorimetry, Gas Diffusion - Amperometry

# Available Cyanide Analysis

# WAD Cyanide methods measure “available cyanide”

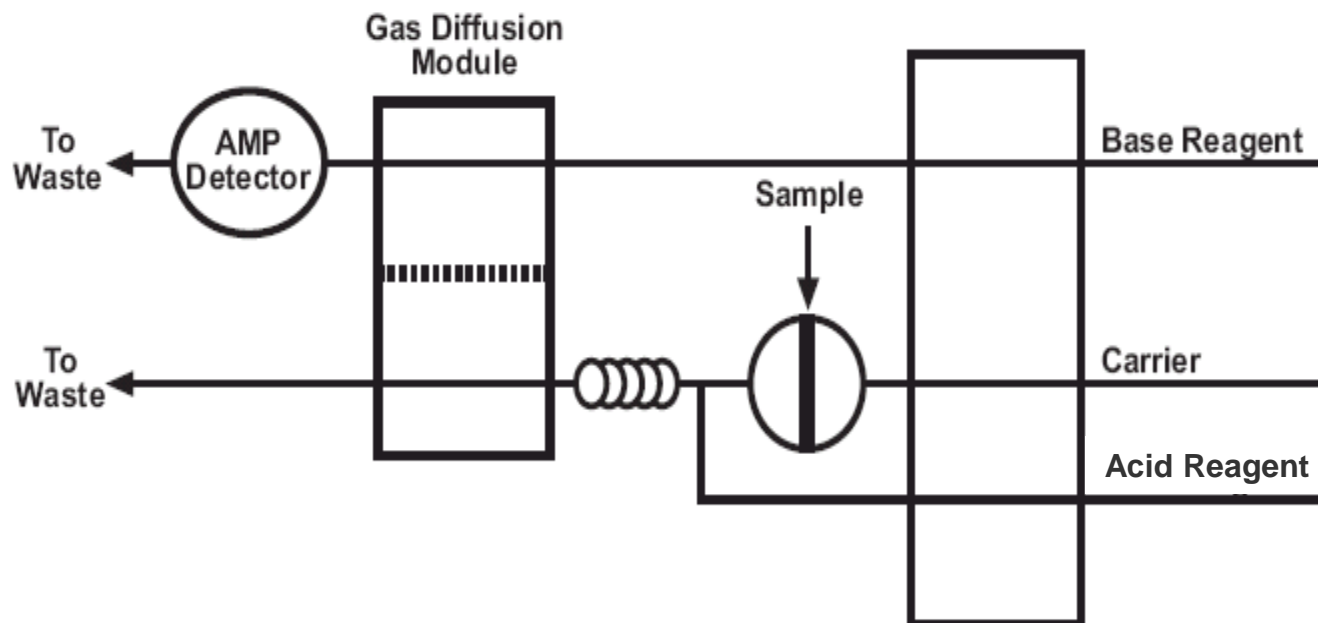
Method Number	Description	Measurement
SM 4500-CN I	Buffered pH 4.5 manual Distillation	Colorimetry
ASTM D 2036	Buffered pH 4.5 manual distillation	Colorimetry, Gas Diffusion - Amperometry

# Ligand Exchange methods measure available cyanide

Method Number	Description	Measurement
OIA 1677	Ligand Exchange / Flow Injection Analysis	Gas Diffusion - Amperometry
ASTM D 6888	Ligand Exchange / Flow Injection Analysis	Gas Diffusion - Amperometry

**GD-amperometry methods do not require distillation**

# OIA 1677 or ASTM D6888 flow diagram



# Ligand Exchange GD-amperometry methods have fewer interferences

CATC	WAD	OIA 1677
N-organics	Excessive Iron Cyanide	None
SCN, NH <sub>3</sub> , NO <sub>2</sub>	Concentration Dependent	—
S <sub>2</sub> O <sub>3</sub> , H <sub>2</sub> O <sub>2</sub>	—	—
Concentration Dependent	—	—



# Ligand Exchange GD-amperometry methods provide the best benefits

- No distillation (eliminates 1 – 4 hours preliminary sample treatment)
- Low MDL (0.5 ppb)
- No Interferences
- High Throughput (up to 90 samples per hour)
- Ease of Operation, very simple chemistry.

CN

NH<sub>3</sub>

PO<sub>4</sub>

NO<sub>3</sub>

# Total Cyanide Methods

**Automated gas diffusion distillation and non-distillation methods**



# Total Cyanide Methods – Manual Distillation

Method Number	Description	Measurement
SM 4500-CN C	Manual Macro Distillation – $\text{Mg Cl}_2$	Manual colorimetry / ISE
ASTM D 2036	Manual Macro Distillation – $\text{MgCl}_2$	Manual colorimetry, ISE, GD-amperometry, IC
EPA 335.4	Midi Distillation – $\text{MgCl}_2$	Automated Colorimetry
ASTM D 7284	Midi / Micro Distillation – $\text{MgCl}_2$	Gas Diffusion - Amperometry

# Automated total cyanide methods use UV to liberate HCN from Fe

Descriptive Name	Method Number	Description	Measurement
Total Cyanide	ASTM D4374 (Kelada 01)	High power <b>UV- Auto distillation</b> Alkaline pH	Automated colorimetry
	EPA 335.3	Low power <b>UV- Auto distillation</b> pH <2	Automated Colorimetry
	OIA 1678/ASTM D7511	Low power <b>UV-</b> pH <2	Gas Diffusion - Amperometry

# Comparison of Kelada and OIA 1678

	Kelada 01	ASTM D7511
Pump Tubes	15	5
Reagents	Pyridine	No Pyridine
Distillation	Yes	No
SCN Interaction	0.25 – 0.5 %	0.01 – 0.03 %

# Comparison of Total CN methods

	<b>335.4</b>	<b>ASTM D7284</b>	<b>ASTM D7511</b>
<b>Sample Preparation</b>	2 – 3 hour distillation	1 – 3 hour distillation	No distillation
<b>Analysis</b>	1 – 2 minutes	1 – 2 minutes	1 – 2 minutes
<b>Total Time</b>	3 – 4 hours	2 – 4 hours	1 – 2 minutes

# Advantages - CNSolution 3100

<b>Total Cyanide</b>	Distillation/ Colorimetry	EPA 335.4
	Distillation / GD- Amperometry	ASTM D7284
	No Distillation / GD- Amperometry	OIA 1678 (ASTM D7511-09)

One Instrument – multiple methods

# How do you solve interference problems caused by distillation?



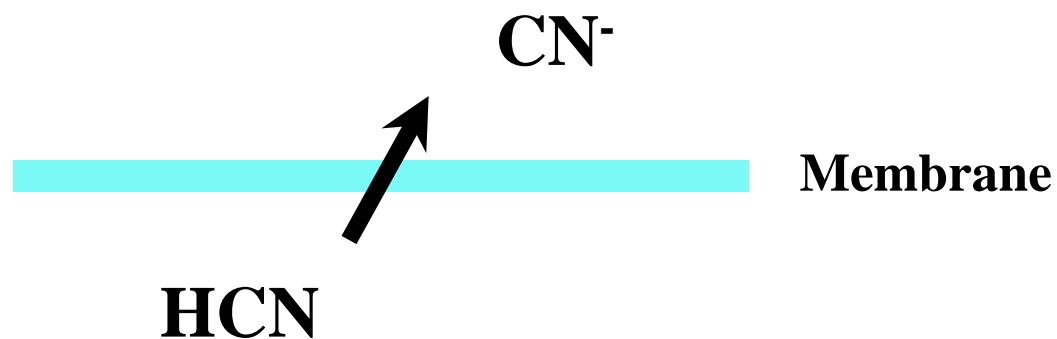
CN

NH<sub>3</sub>

PO<sub>4</sub>

NO<sub>3</sub>

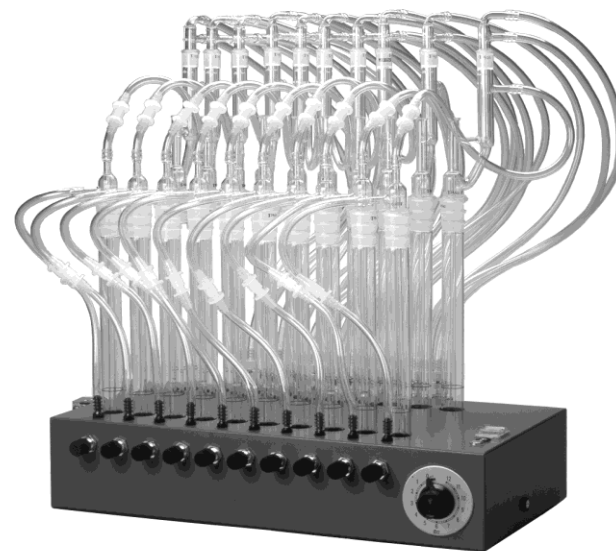
# Replace distillation with gas diffusion



# Distillation most common technique to remove interference



**Macro Distillation**



**MIDI  
Distillations**



# Many cyanide interferences result from distillation

- Destroy CN
- Create CN
- UV distillation colorimetry - worst



# Distillation actually creates CN interferences

- **Boiling acid**
- **Automated UV-Distillation**
  - Boiling acid



# Interferences –

**Thiocyanate**

**Destroy CN**

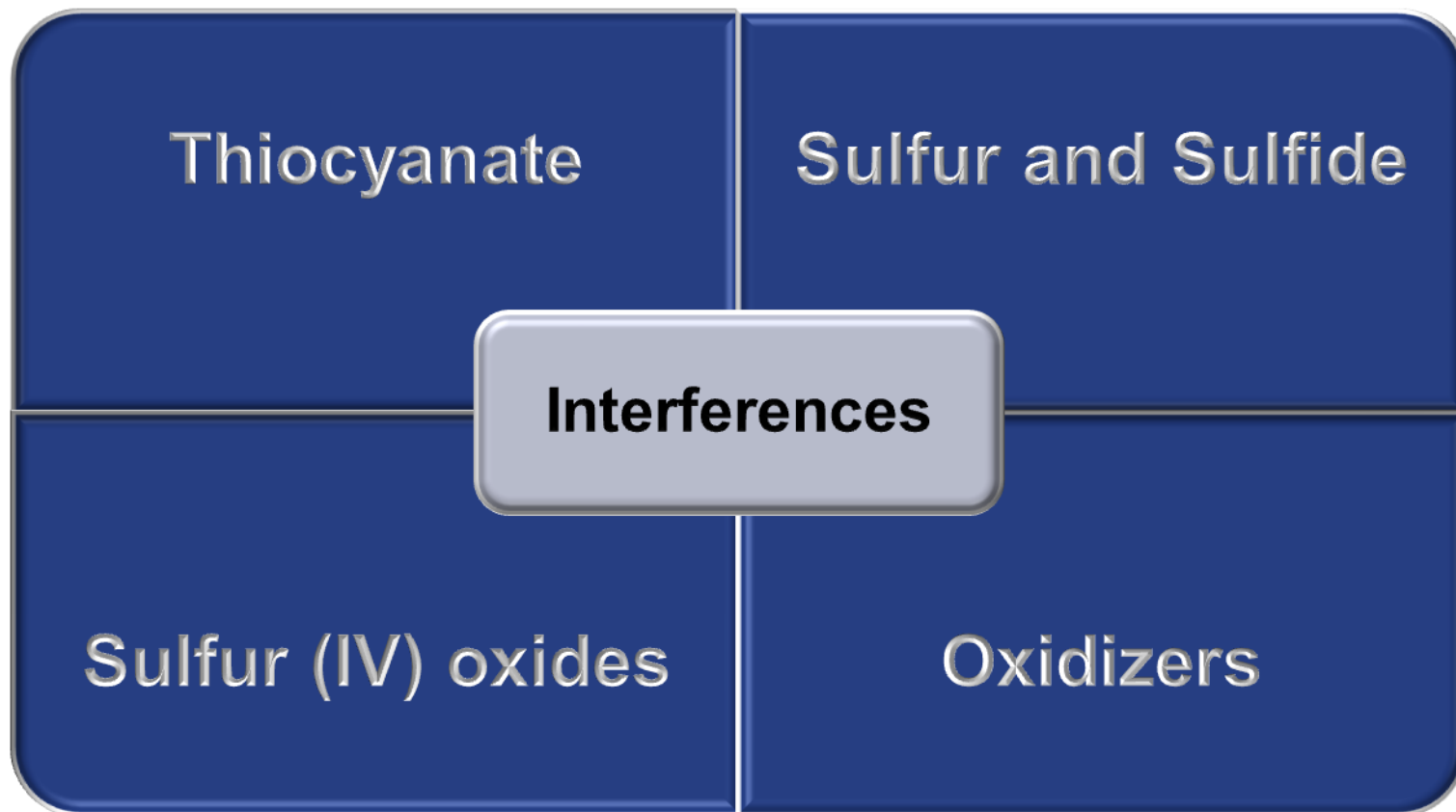
**Create CN**



# Thiosulfate reacts with cyanide during distillation

- **0.200 mg/L  $\text{CN}^-$  + 200 mg/L  $\text{S}_2\text{O}_3^{2-}$** 
  - Cyanide Found = 0.160 mg/L
  - Recovery = 80%\*
- \* Double Chloramine T added, or recovery would be lower.

**These compounds are in almost every sample  
and interfere significantly**

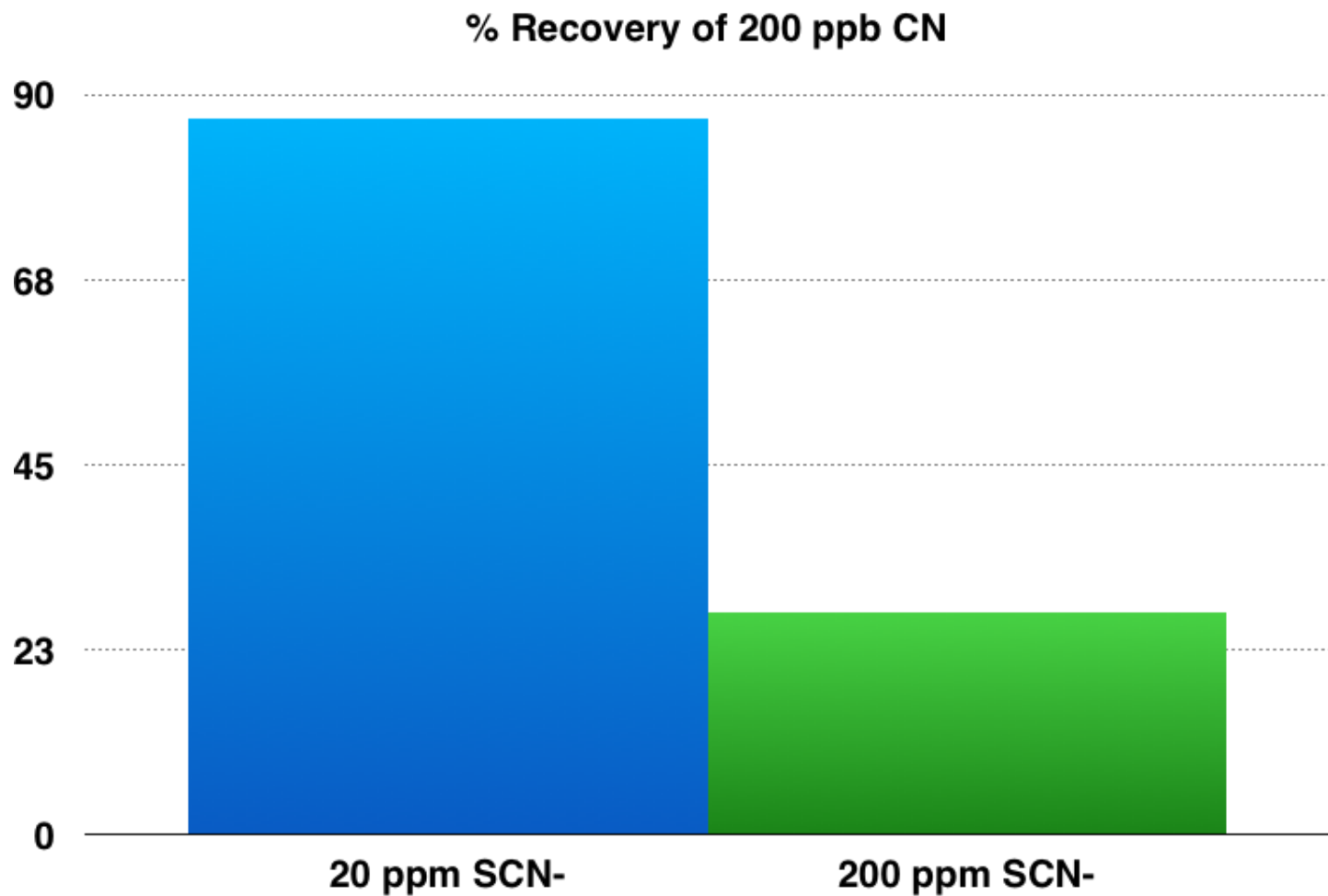


The left side of the slide features a vertical blue bar with a white chromatogram line. Along this line are four boxes containing chemical symbols:  $\text{CN}^-$ ,  $\text{NH}_3$ ,  $\text{PO}_4$ , and  $\text{NO}_3^-$ .

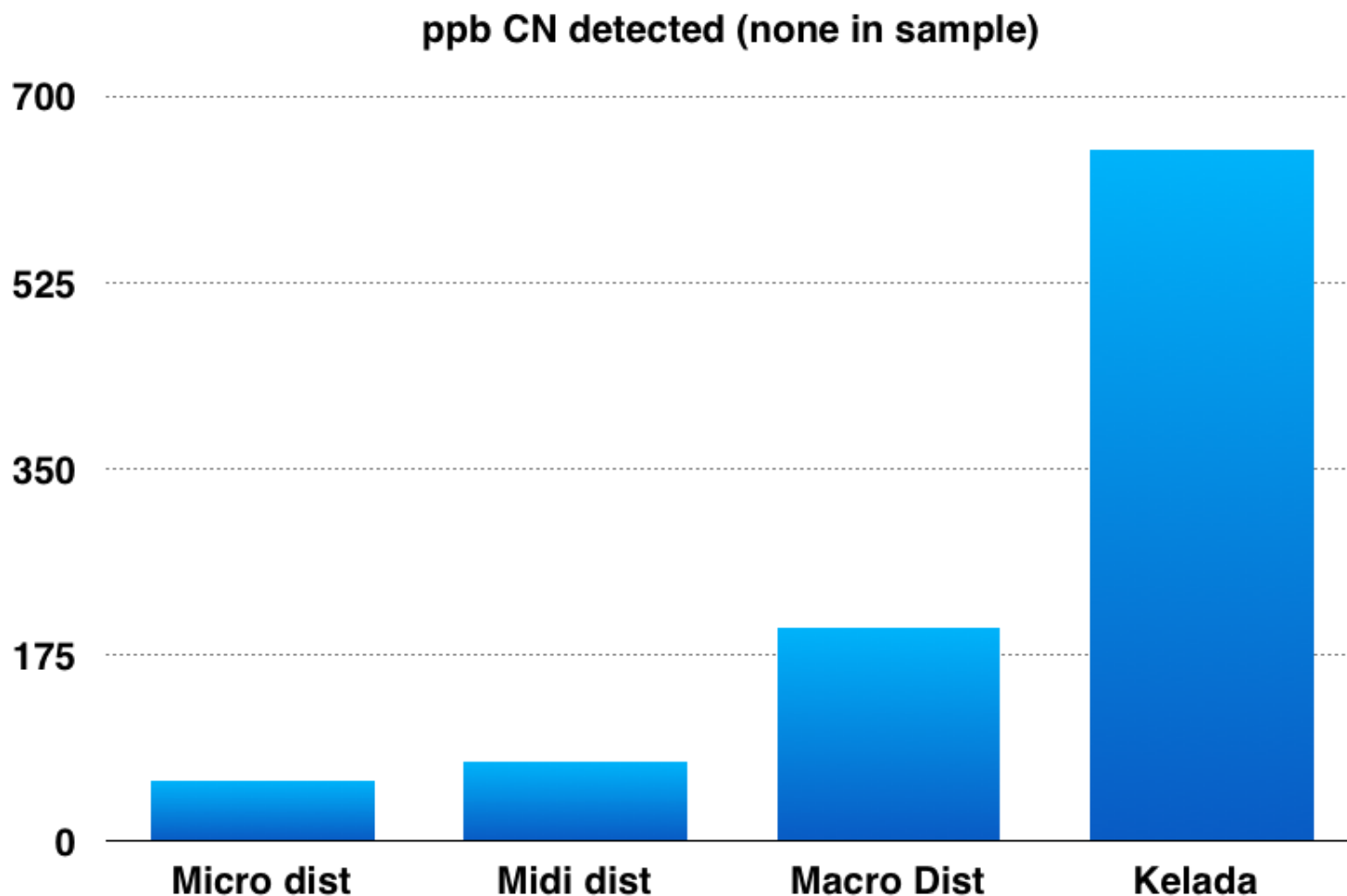
# Thiocyanate + Nitrate results in positive bias

- The addition of Sulfamic acid does not sufficiently reduce this interference.
  - A real POTW sample with 0.1 mg/L  $\text{SCN}^-$  and 63.5 mg/L  $\text{NO}_3^-$  detected total  $\text{CN}^-$  at **0.10 mg/L** even after the addition of Sulfamic Acid

# Interferences – Thiocyanate



# Interferences – Thiocyanate and Nitrate



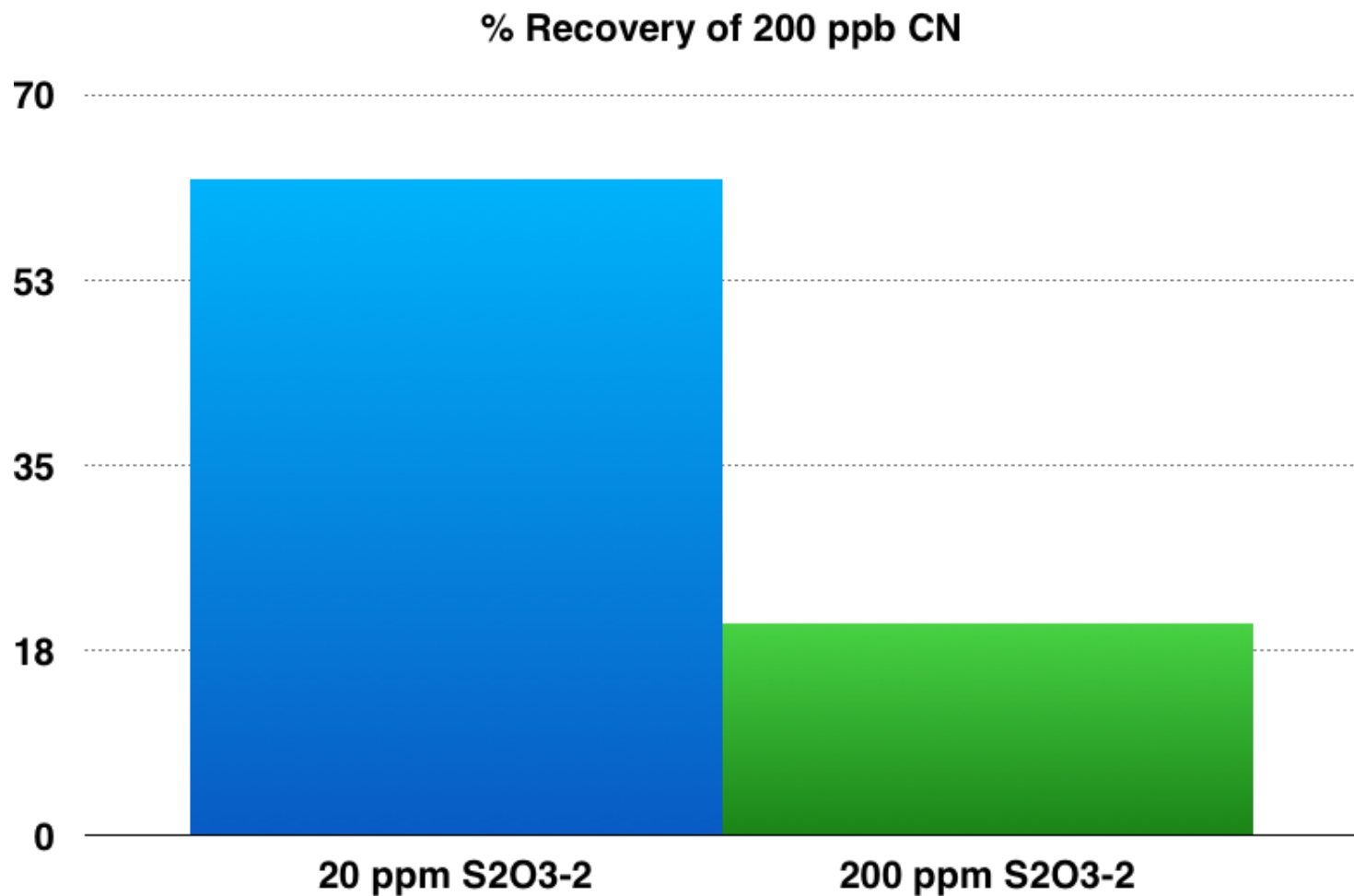




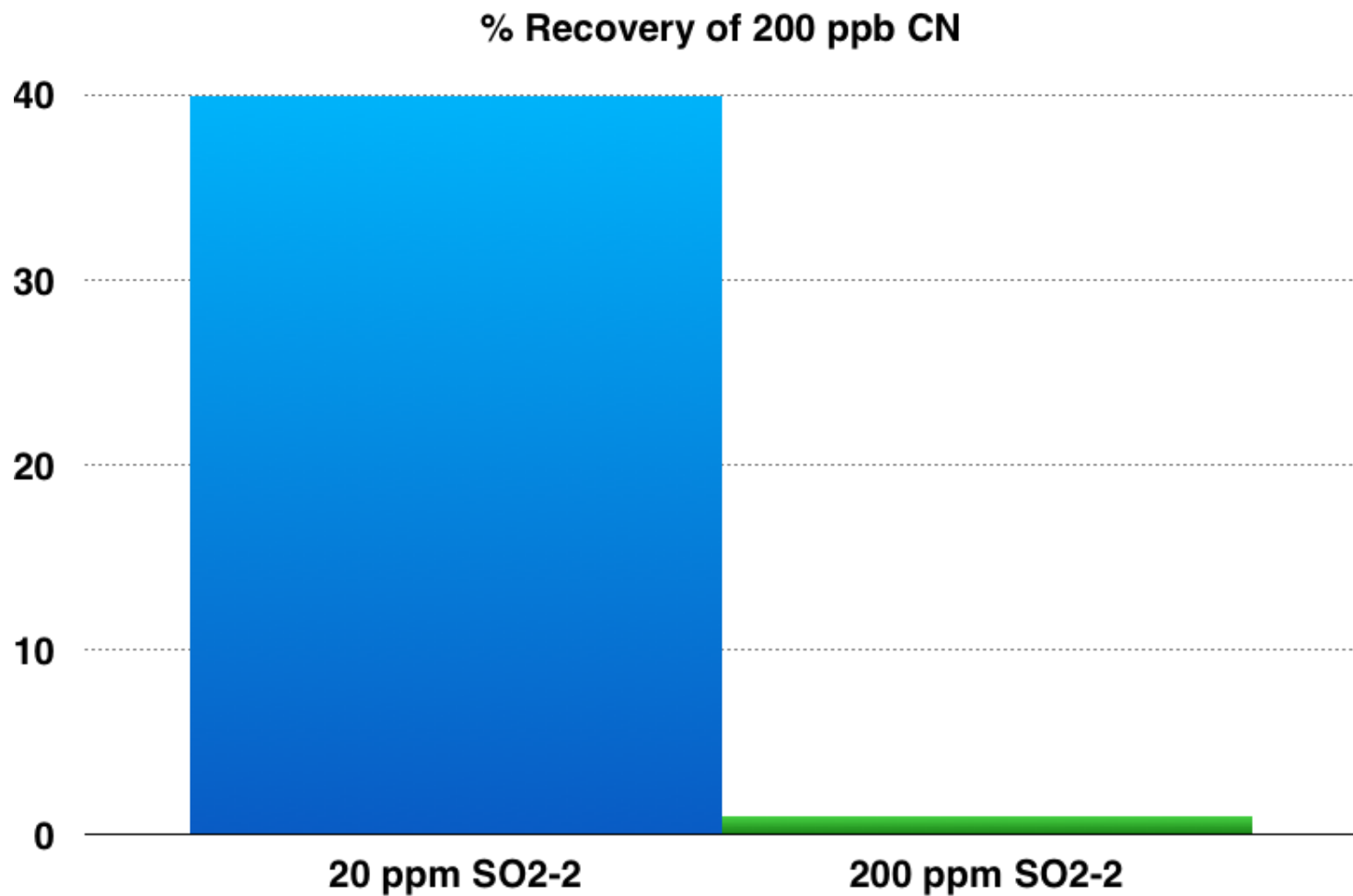
# Sulfite reacts rapidly with CN in basic solutions

- **0.200 mg/L CN<sup>-</sup> + 200 mg/L SO<sub>3</sub><sup>-2</sup>**
  - Cyanide Found = 0.000 mg/L
  - Recovery = 0%
- This reaction occurs in absorber solution, or in preserved sample

# Interferences – Thiosulfate



# Interferences – Sulfite



# Interferences – Sulfur Compounds

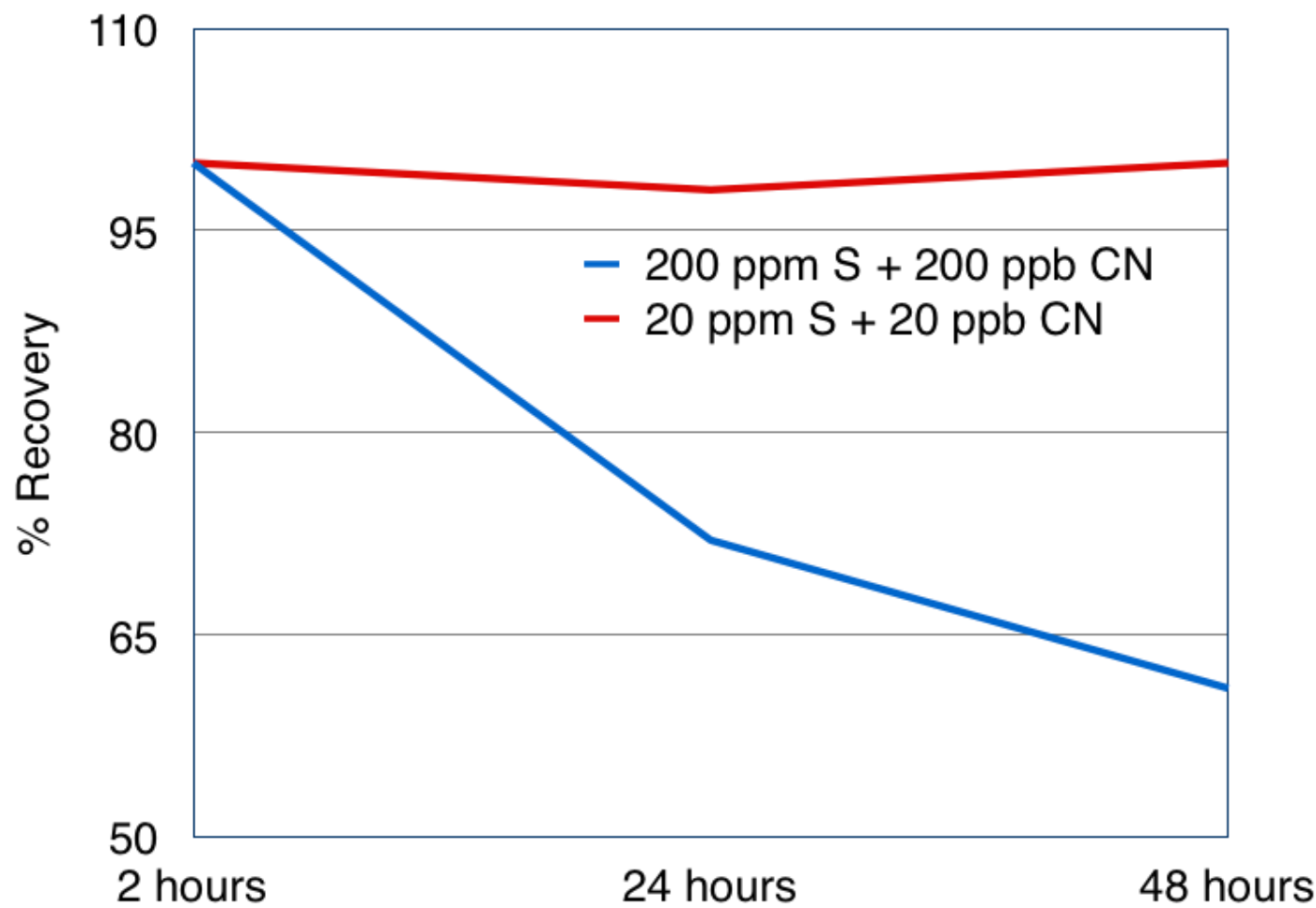
- No Spot Tests
- Dechlorination



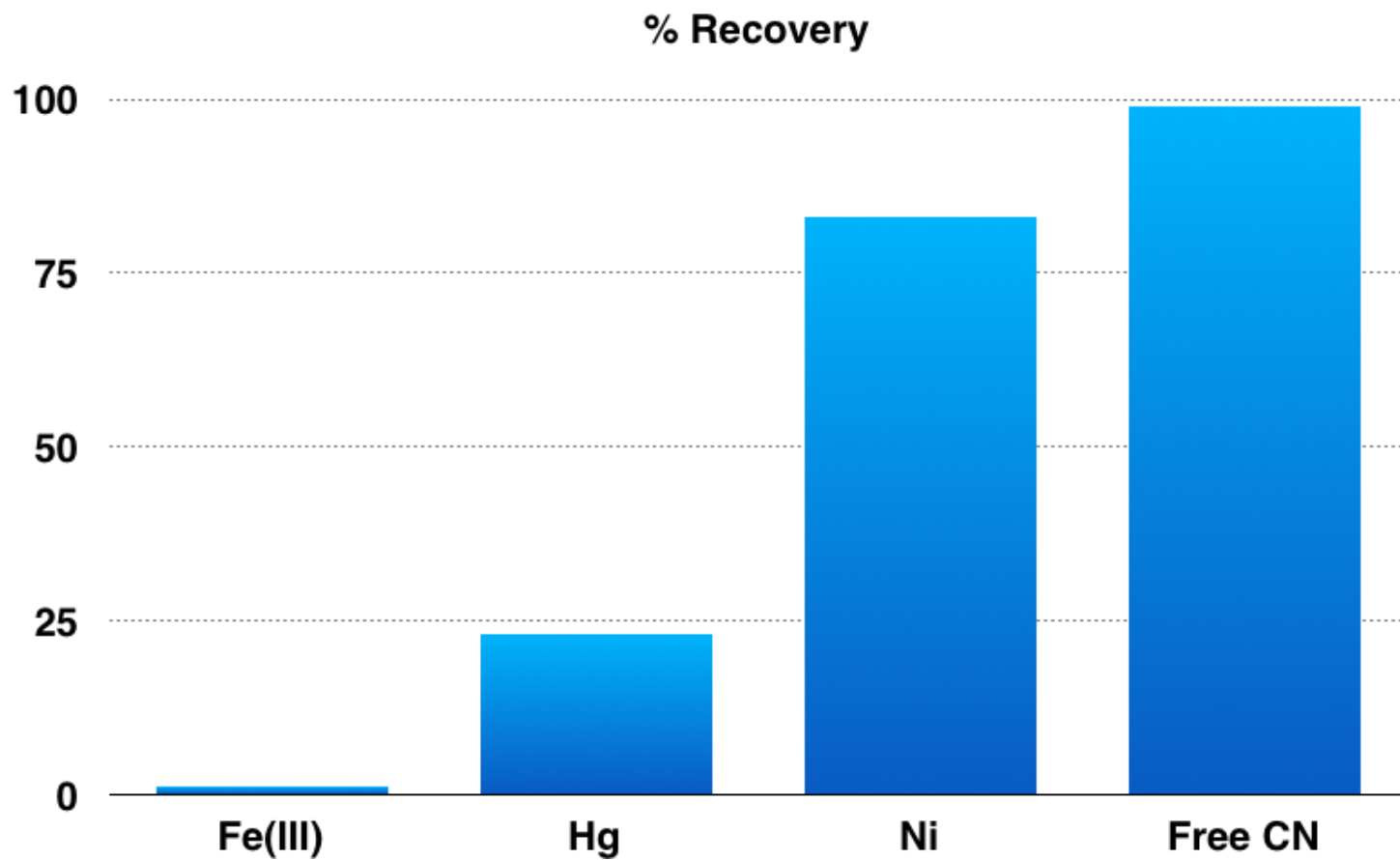
# Sulfur compounds react rapidly with CN

- **Elemental Sulfur**
  - $8\text{CN}^- + \text{S}_8 \rightarrow \text{SCN}^-$
- **Metal Sulfides**
  - $\text{Cu}_2\text{S}$ ,  $\text{FeS}$ ,  $\text{PbS}$ ,  $\text{CuFeS}_2$ ,  $\text{CdS}$ ,  $\text{ZnS}$ , etc.
  - S reacts with  $\text{CN}^-$  to form  $\text{SCN}^-$

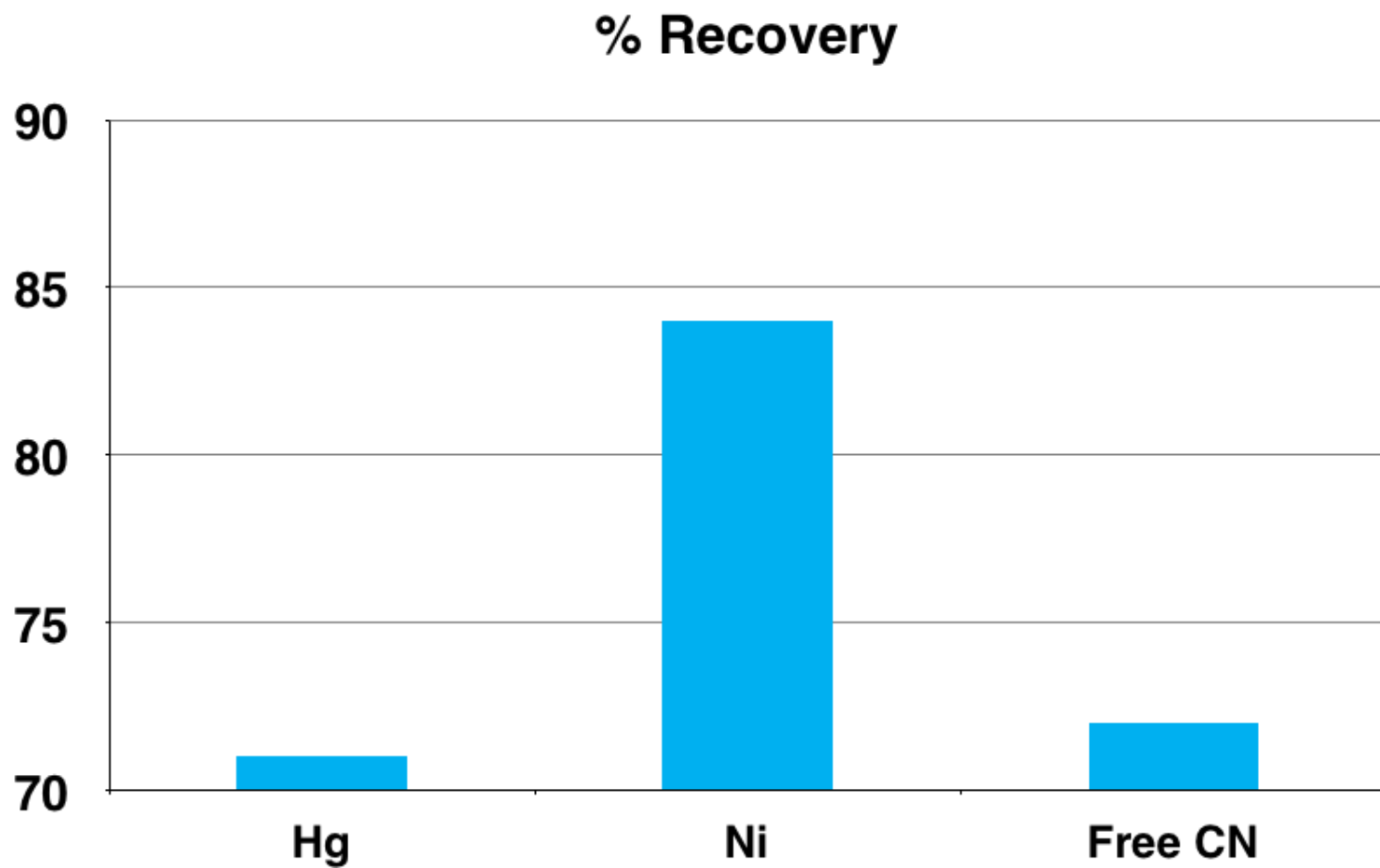
# Holding Time Study – Sulfide Bearing Samples



# Cannot use Cadmium to Treat Sulfide

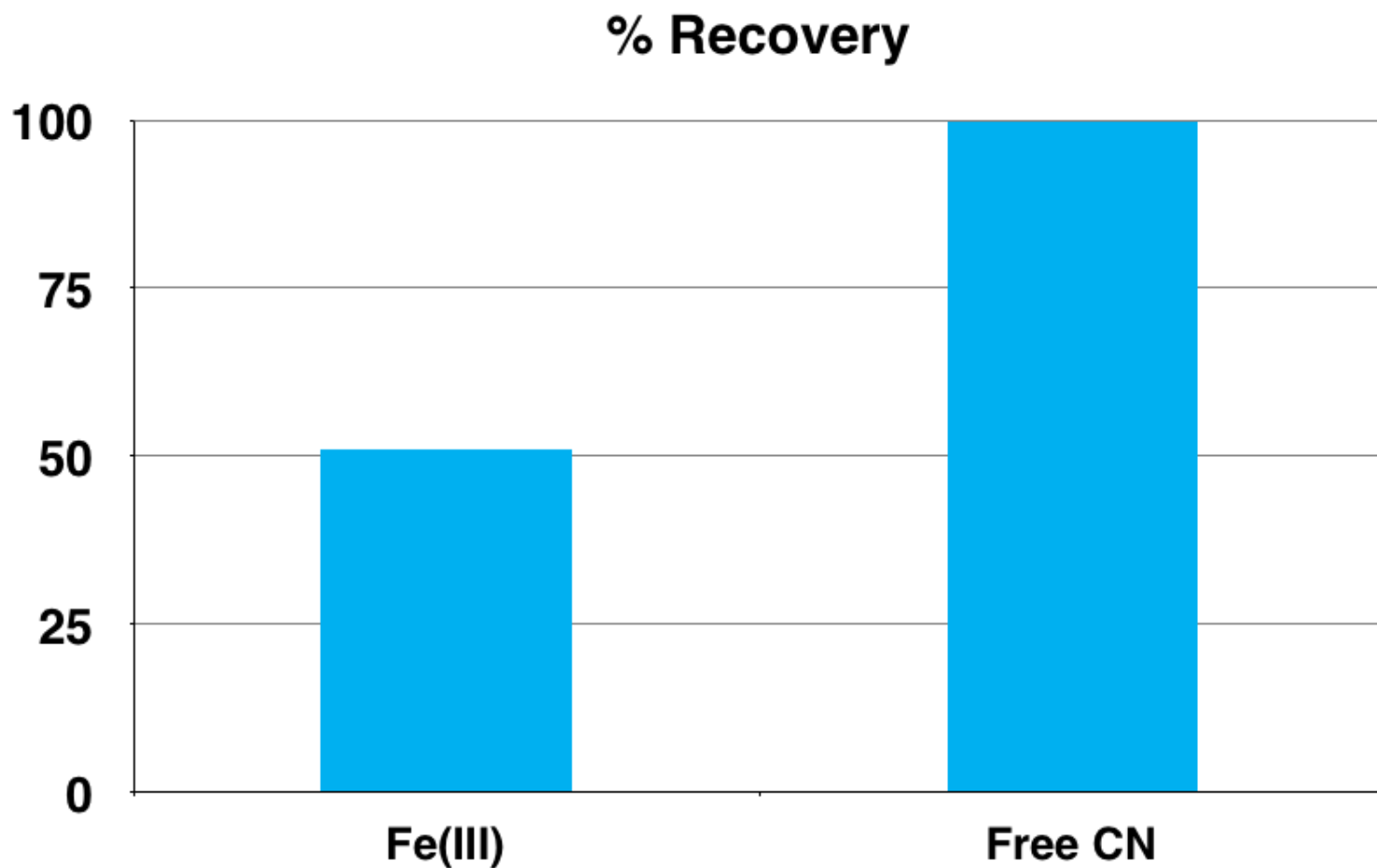


# Headspace to Treat Sulfide





# Bismuth to Treat Sulfide then distillation



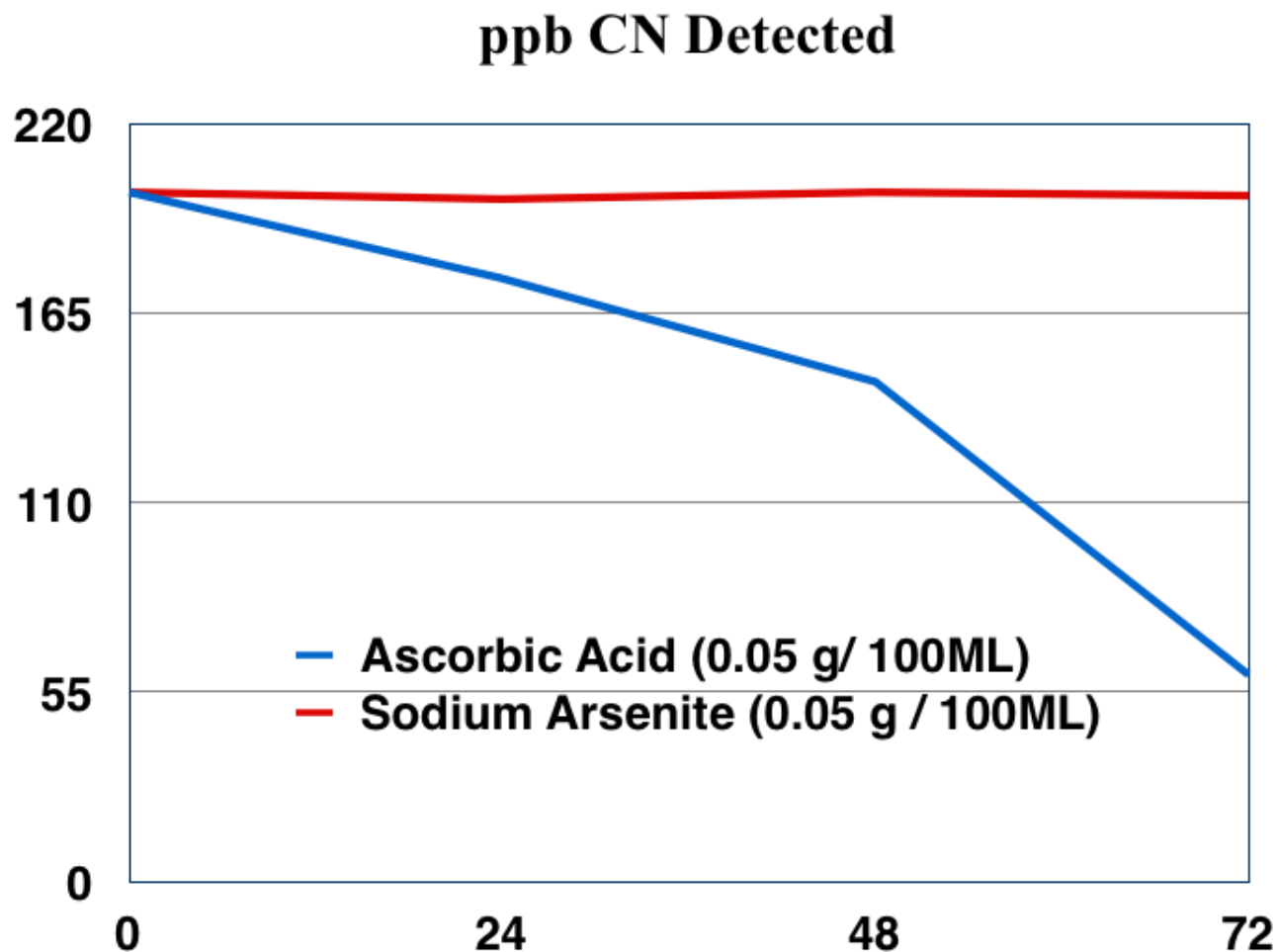


# Oxidizers destroy cyanide before or during distillation

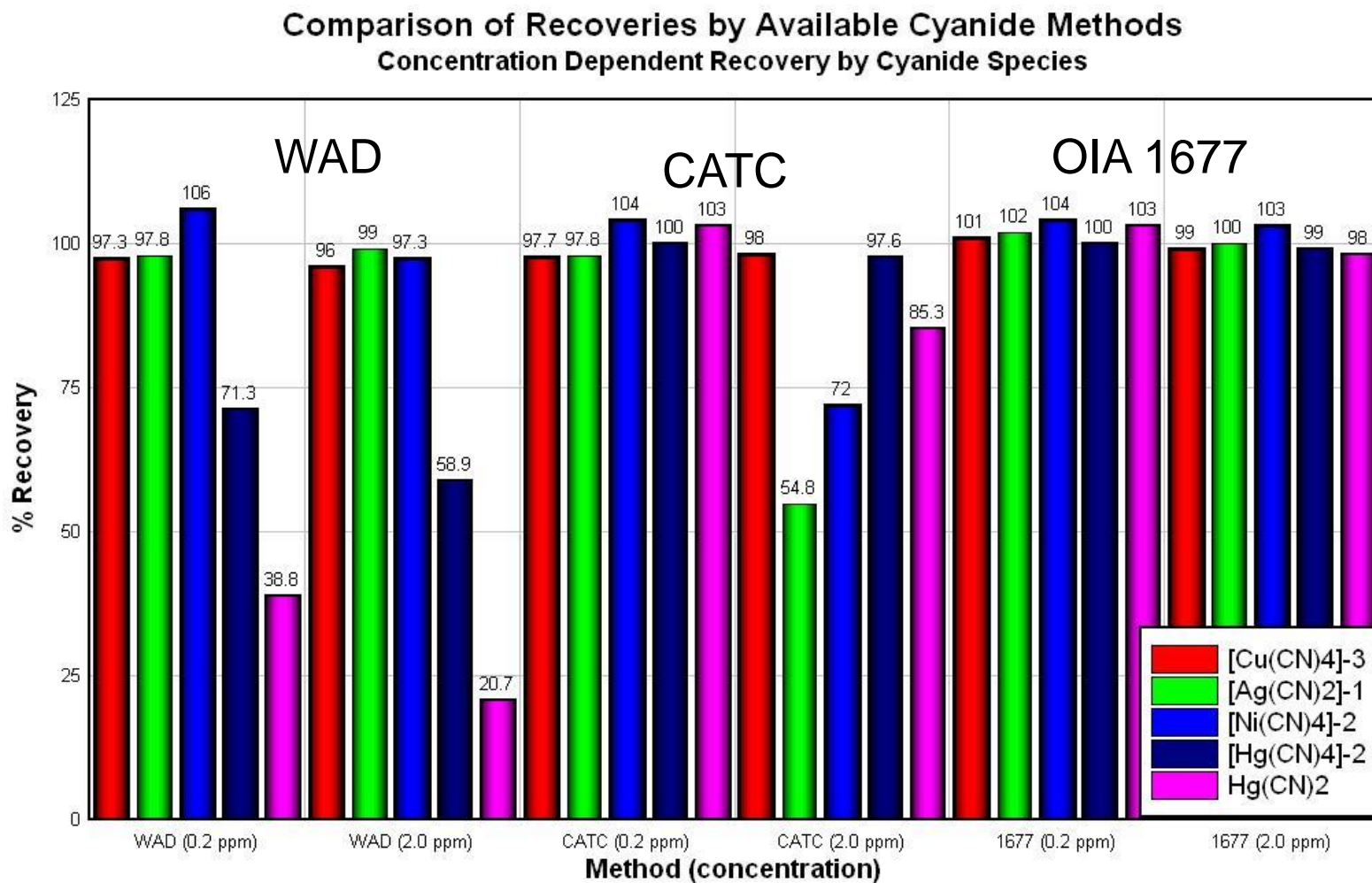
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- Hypochlorite
- Peroxide
- Caro's Acid
- Chloramines

# Oxidizer Removal



# Ligand Exchange GD-amperometry methods get better recovery

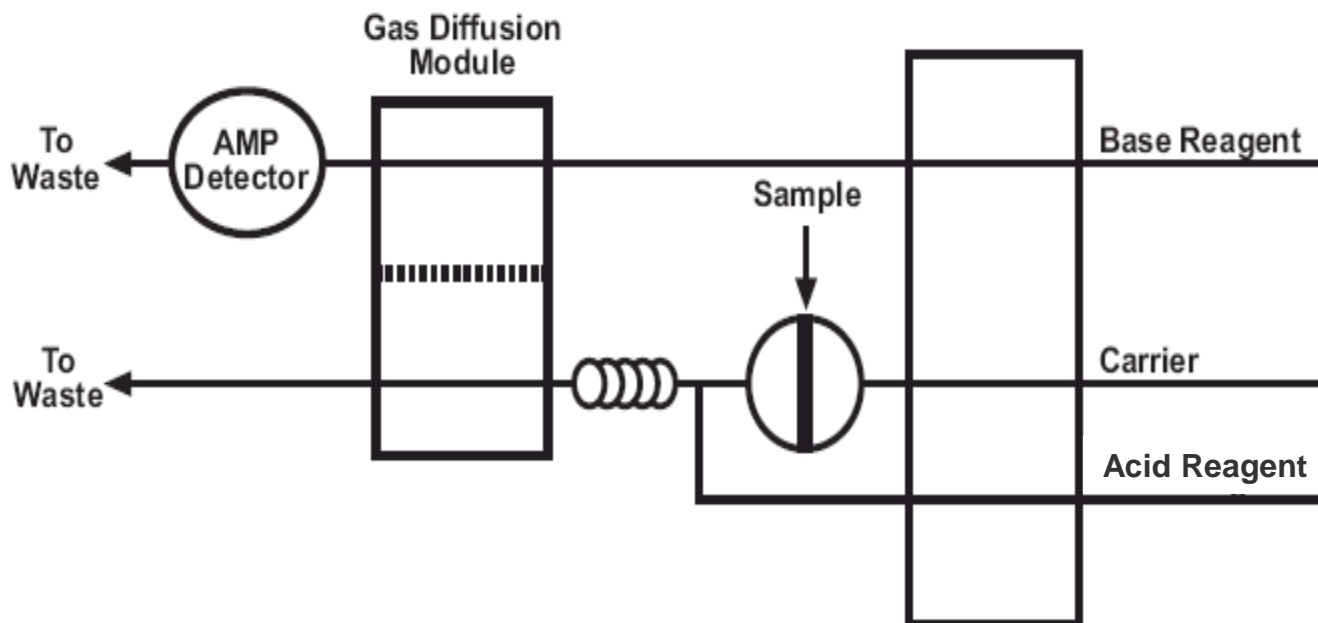


# Cyanide methods that utilize distillation have significant disadvantages

- **Time Consuming**
  - One hour long distillation (does not take into account setup and teardown)
  - CATC requires two, one hour distillations
- **Bulky and Relatively Expensive Glassware**
- **Operator-dependent results (technique)**
- **Multiple Interferences**

# GD-amperometry provides the safest, easiest, and most accurate technique

No pyridine



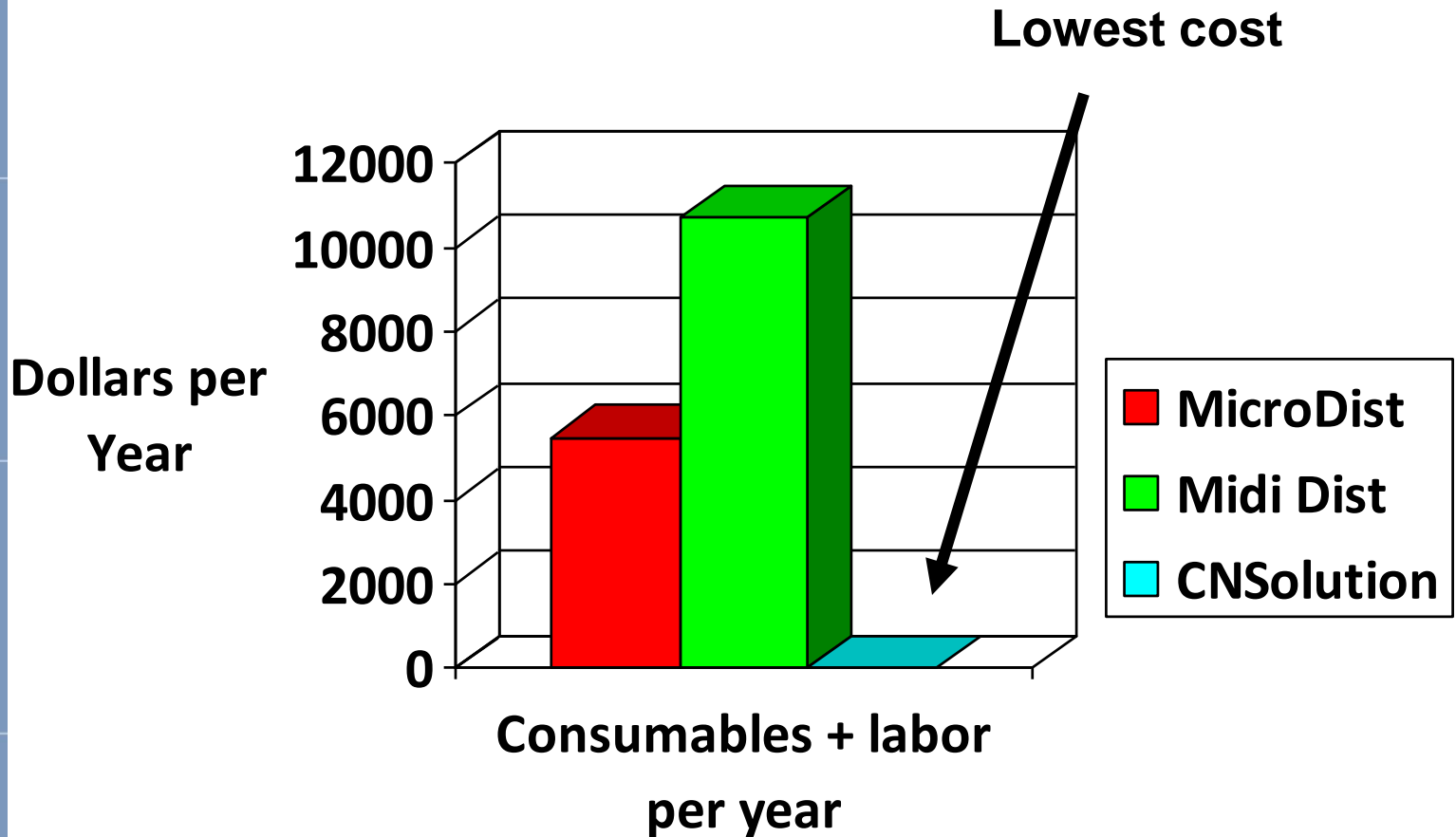
ASTM D7284-08 is a “green” chemistry

The left side of the slide features a vertical blue bar with a white chromatogram line. Four chemical formulas are enclosed in white boxes: CN, NH<sub>3</sub>, PO<sub>4</sub>, and NO<sub>3</sub>.

# CNSolution 3100

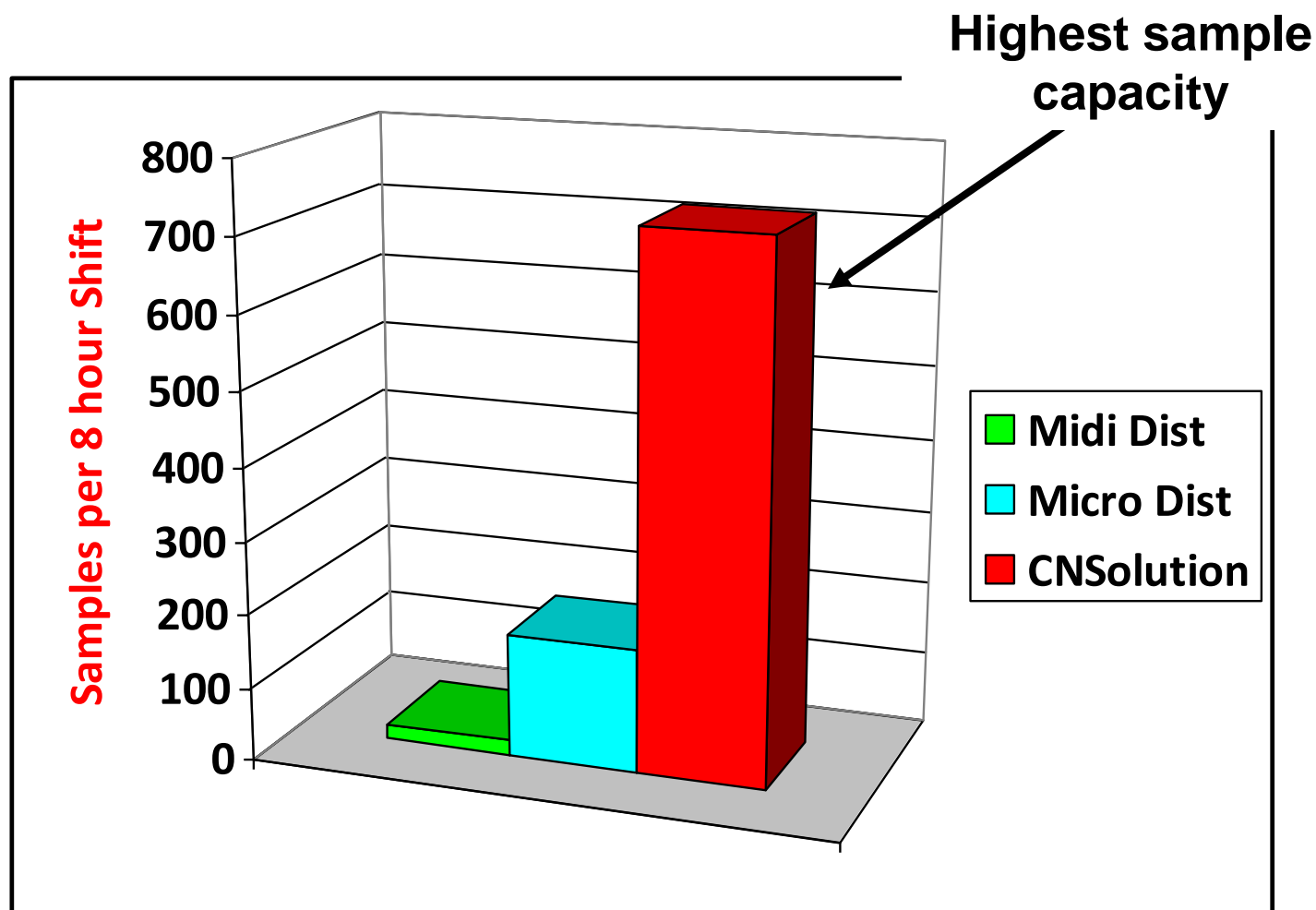
- Rapidly Analyze Cyanide with Confidence in Results
  - Free Cyanide
  - Available Cyanide
  - Total Cyanide
- Eliminate Time Consuming, Error Causing Distillations
- Expand Capability to Colorimetric FIA and SFA

# Gas diffusion eliminates distillation and associated cost





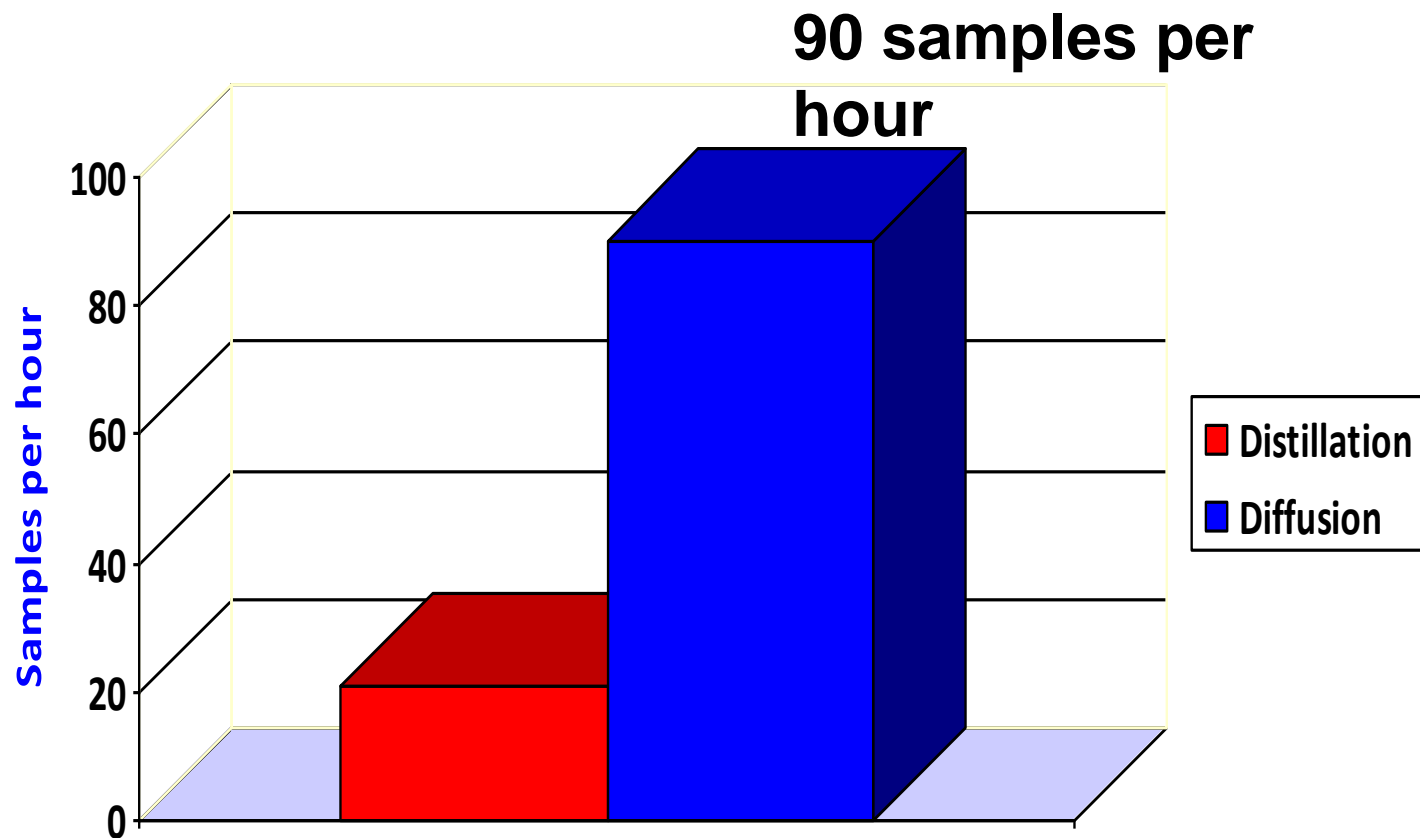
# Eliminating distillation increases laboratory capacity



# Ligand Exchange GD-amperometry methods give you results in minutes

	CATC	WAD	OIA 1677
Sample Preparation	2 distillations 2 – 3 hours	1 distillation 2 – 3 hours	No distillation
Analysis	1 – 2 minutes	1 – 2 minutes	1 – 2 minutes
Total Time	3 – 4 hours	3 – 4 hours	1 – 2 minutes

# Ligand Exchange GD-amperometry methods means more samples





# The CNSolution is your solution to cyanide analysis

- **Gas diffusion amperometry methods:**
  - Save time
  - Save money
  - Are more accurate
  - Have fewer interferences
  - Are “green”
- **The CNSolution = the FS3100**
  - Change to a photometric detector and run any colorimetric chemistry

CN

NH<sub>3</sub>

PO<sub>4</sub>

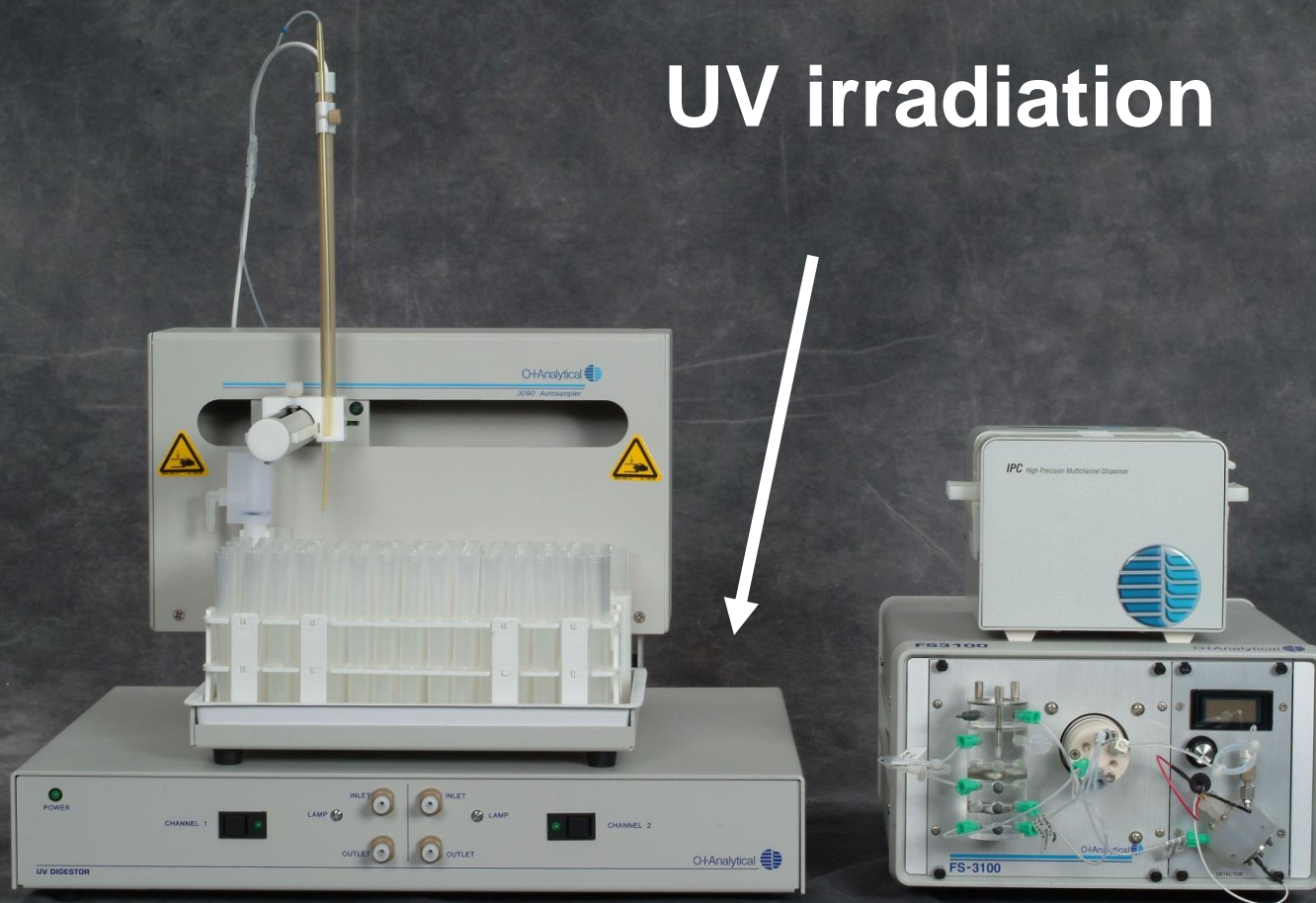
NO<sub>3</sub>

# The CNSolution 3100 available cyanide system



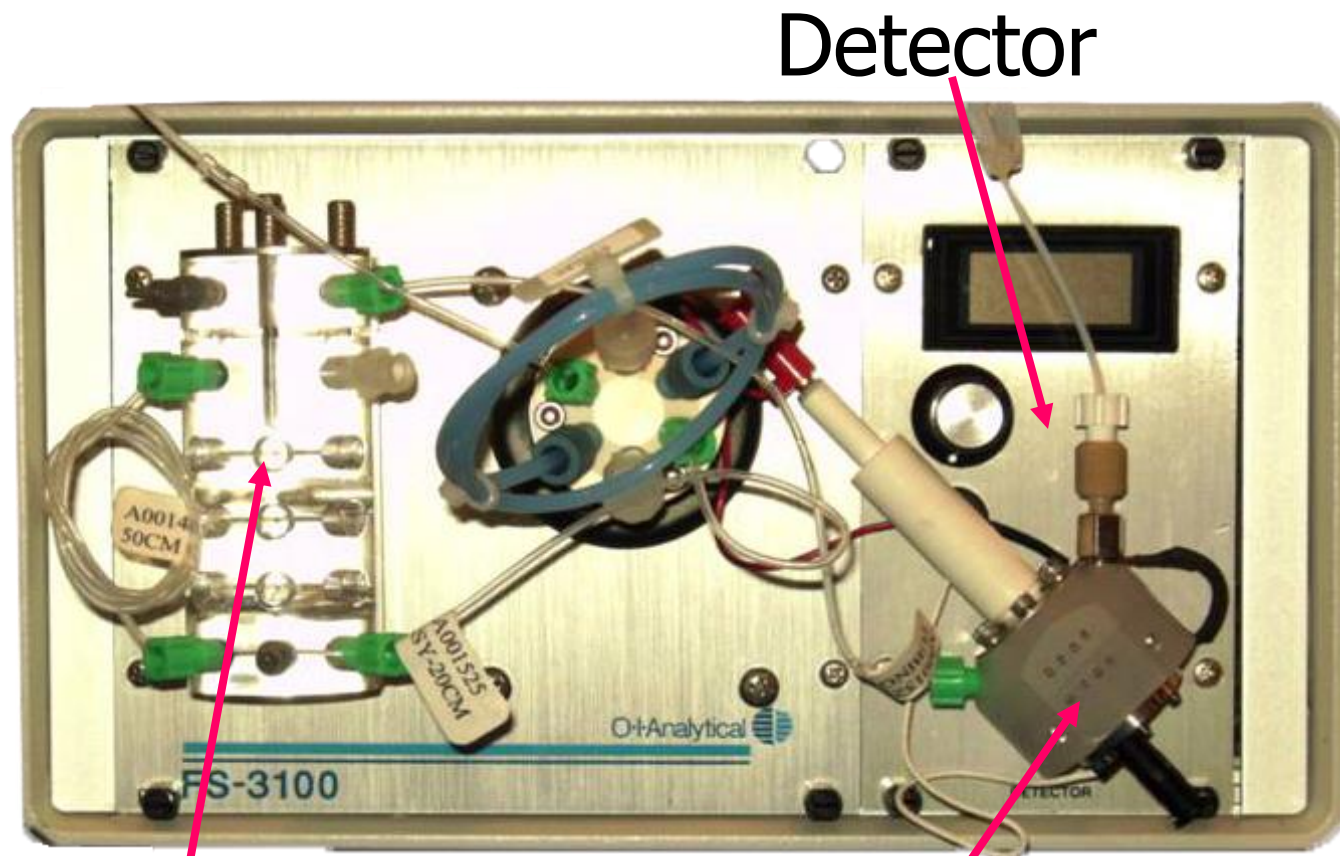
# The CNSolution 3100 total cyanide system

UV irradiation





# OIA 1677 - CNSolution



Gas Diffusion  
and Mixing

Flow-cell

# Flow Solution 3100

- Flow Injection Analyzer
  - Colorimetric detector added it becomes the FS 3100 FIA analyzer
  - All FS 3100 methods
    - $\text{NO}_3\text{-N}$
    - $\text{NH}_3\text{-N}$
    - $\text{PO}_4$
    - TKN



# A high quality peristaltic pump



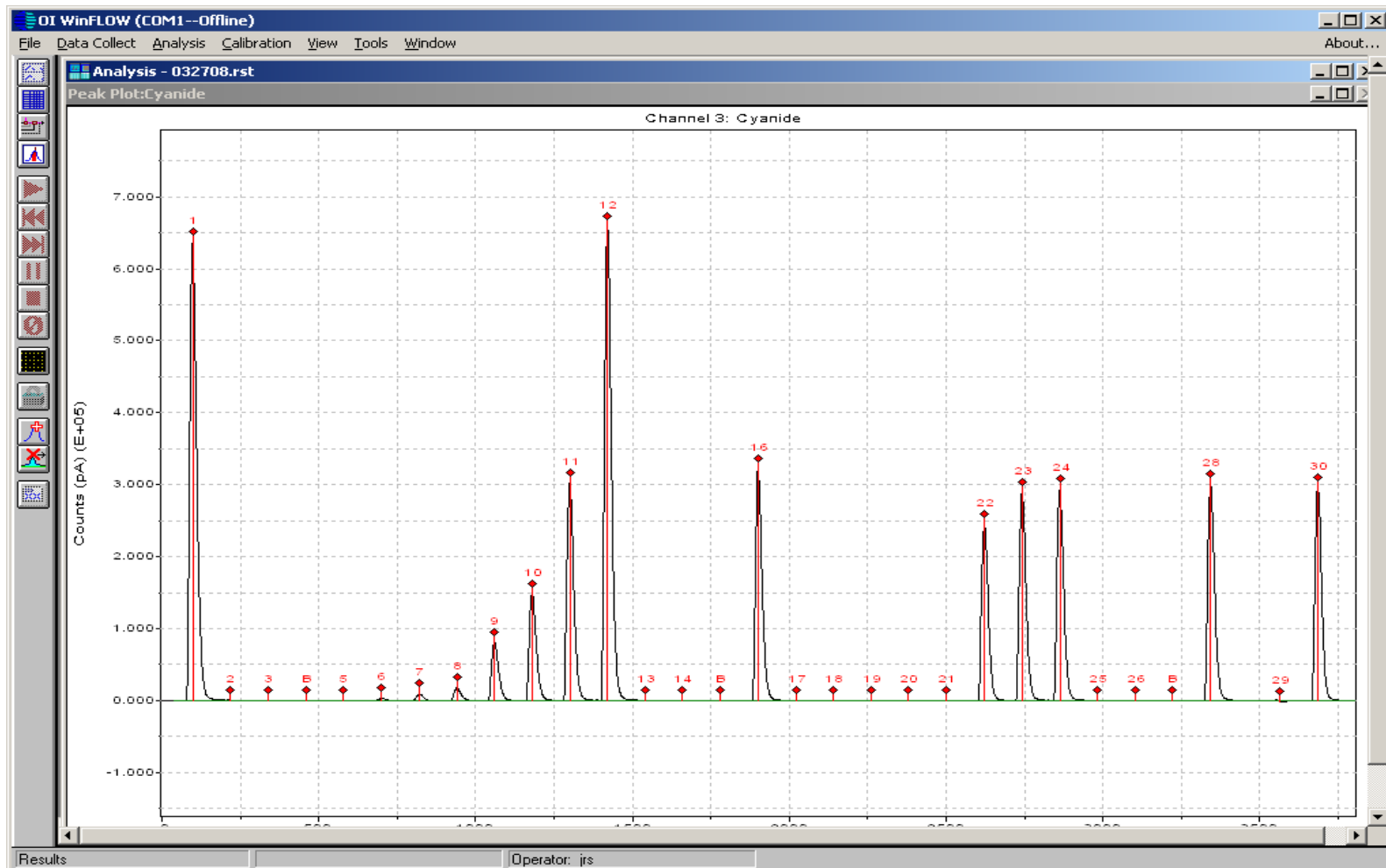
**Separate module, individual channels, long pump tube life.**

# A random access xyz sampler



- Integrated circulating wash station
- 90 or 360 positions
- Separate standard and QC vials

# Winflow software provide accurate quantitation of CN results





# The CNSolution 3100 is accurate, and cost effective

- **Rapidly analyze cyanide**
- **Eliminate time consuming error causing distillations**
- **Expand capability to colorimetric FIA and SFA**

CN

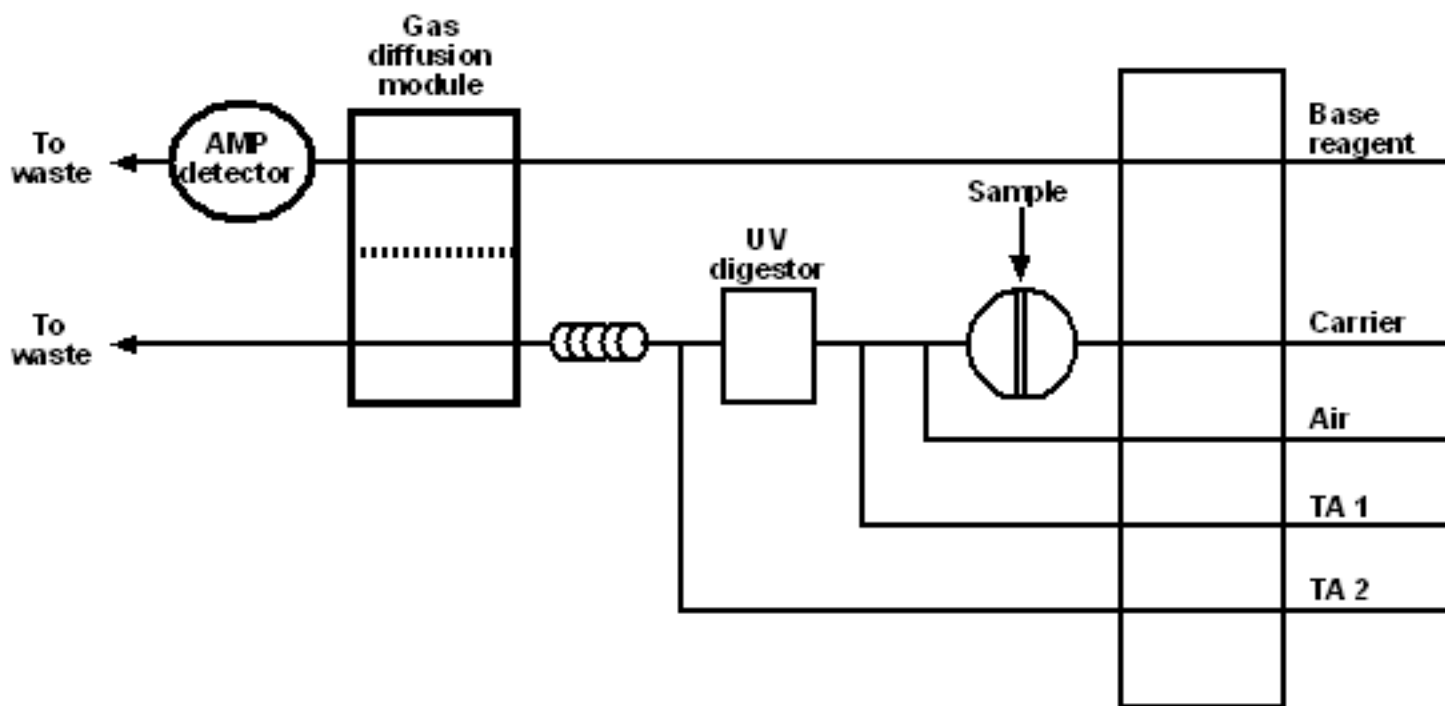
NH<sub>3</sub>

PO<sub>4</sub>

NO<sub>3</sub>

# ASTM D7511 is easy to understand and operate and does not distill.

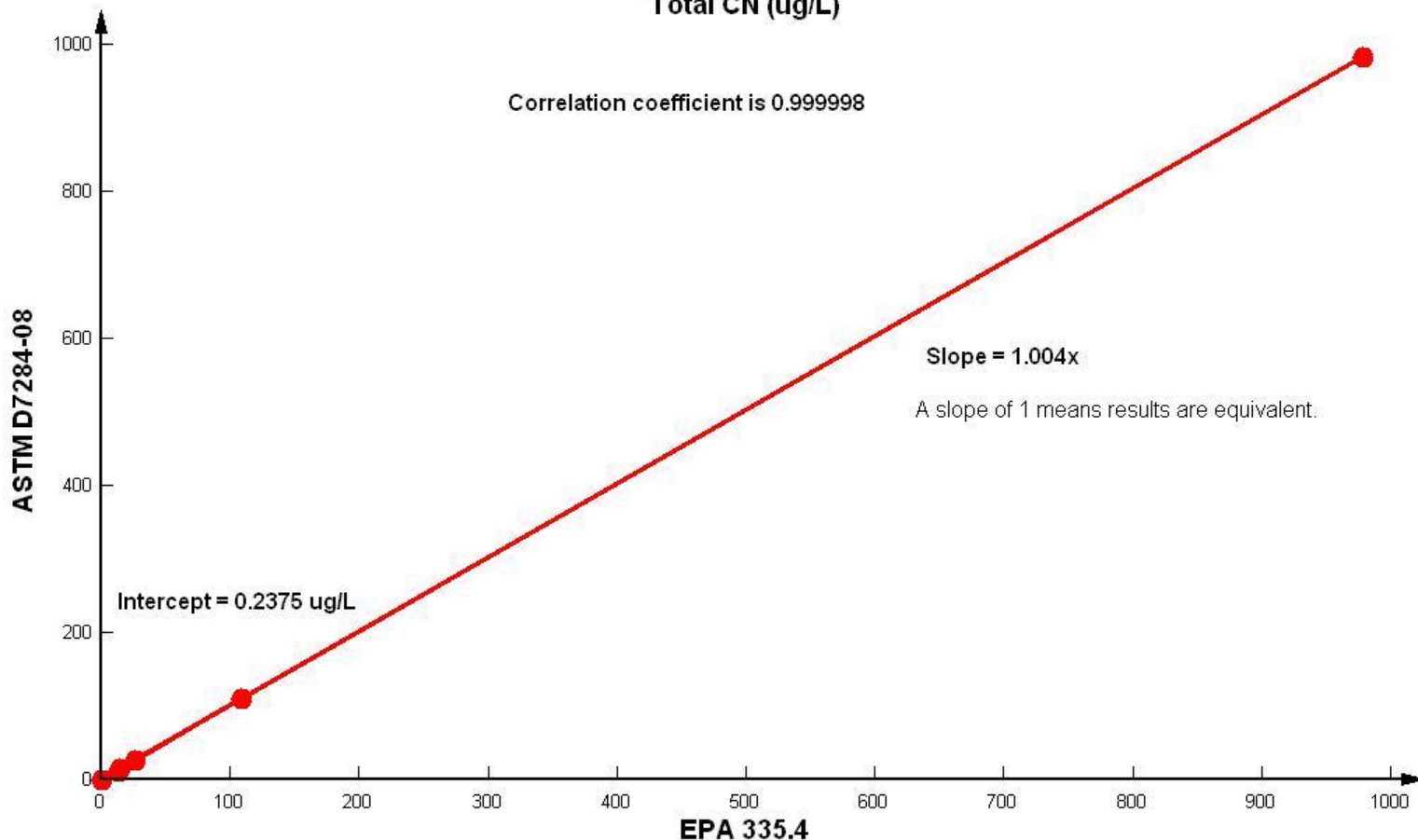
No pyridine



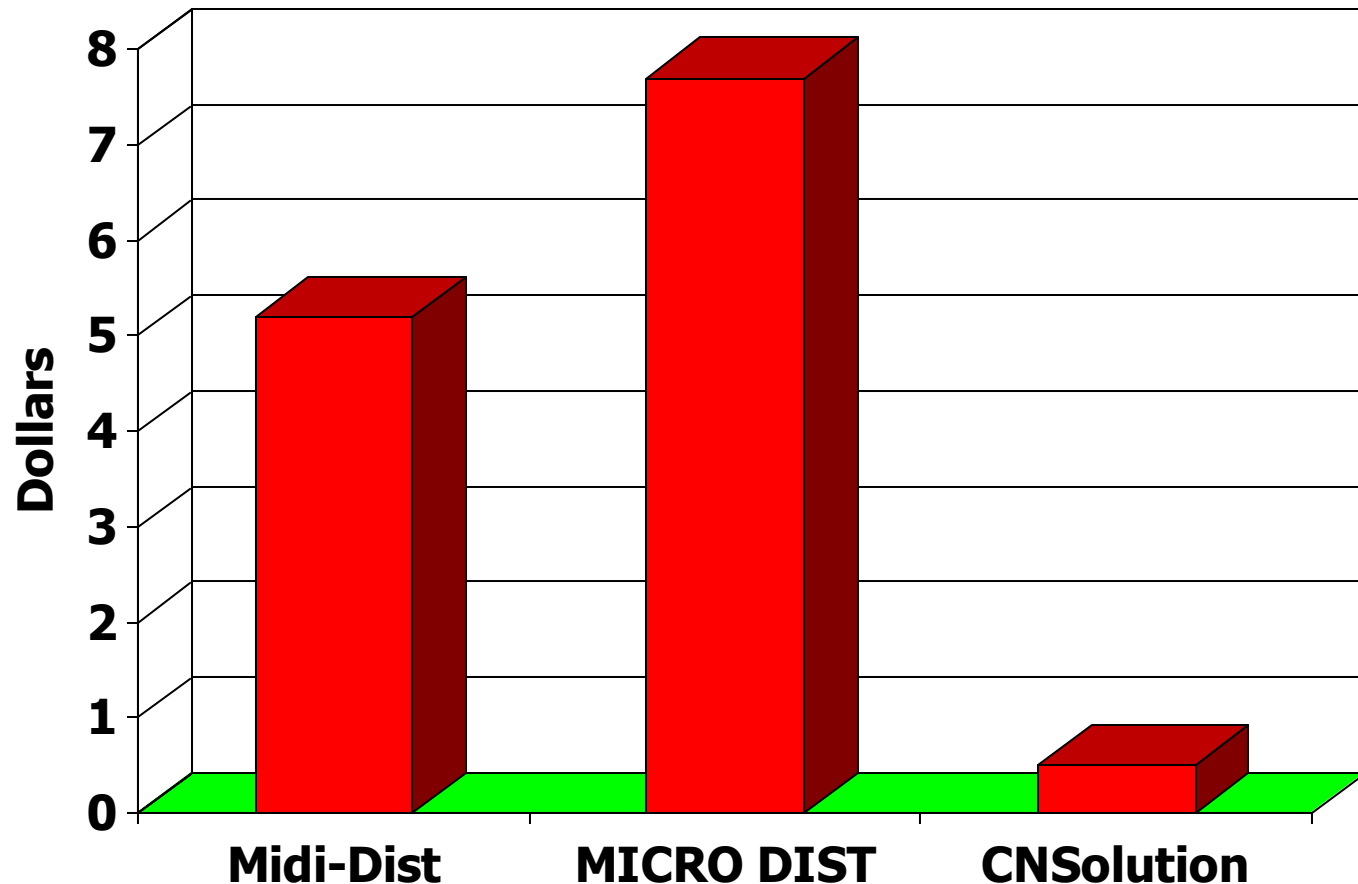
“Green” Chemistry

# GD-Amperometry Equivalent to Colorimetry

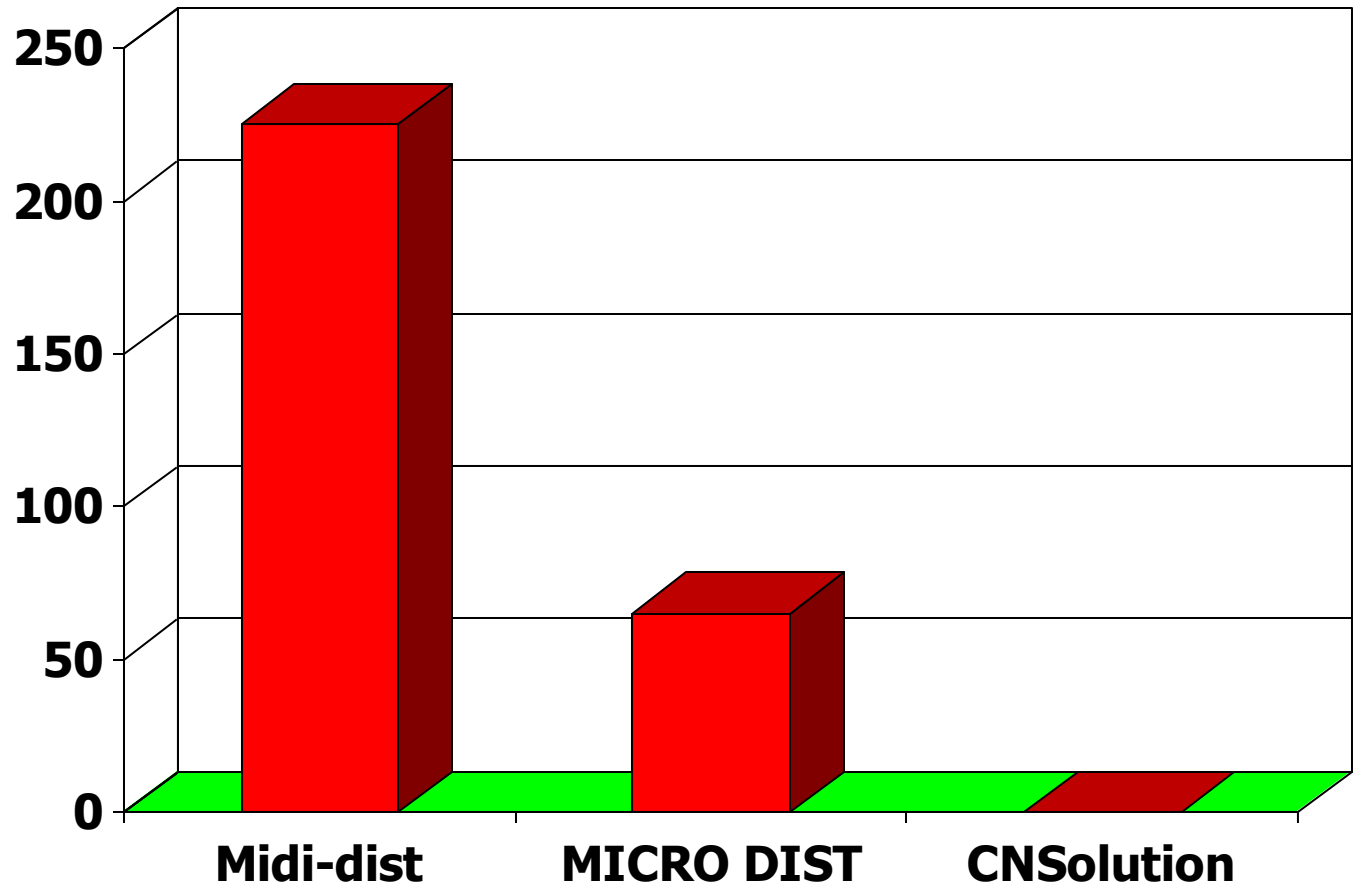
Comparison of EPA 335.4 and ASTM D7284-08  
Total CN (ug/L)



# Estimated Reagent and Consumable Cost per Analysis for Distillation (US Dollars)

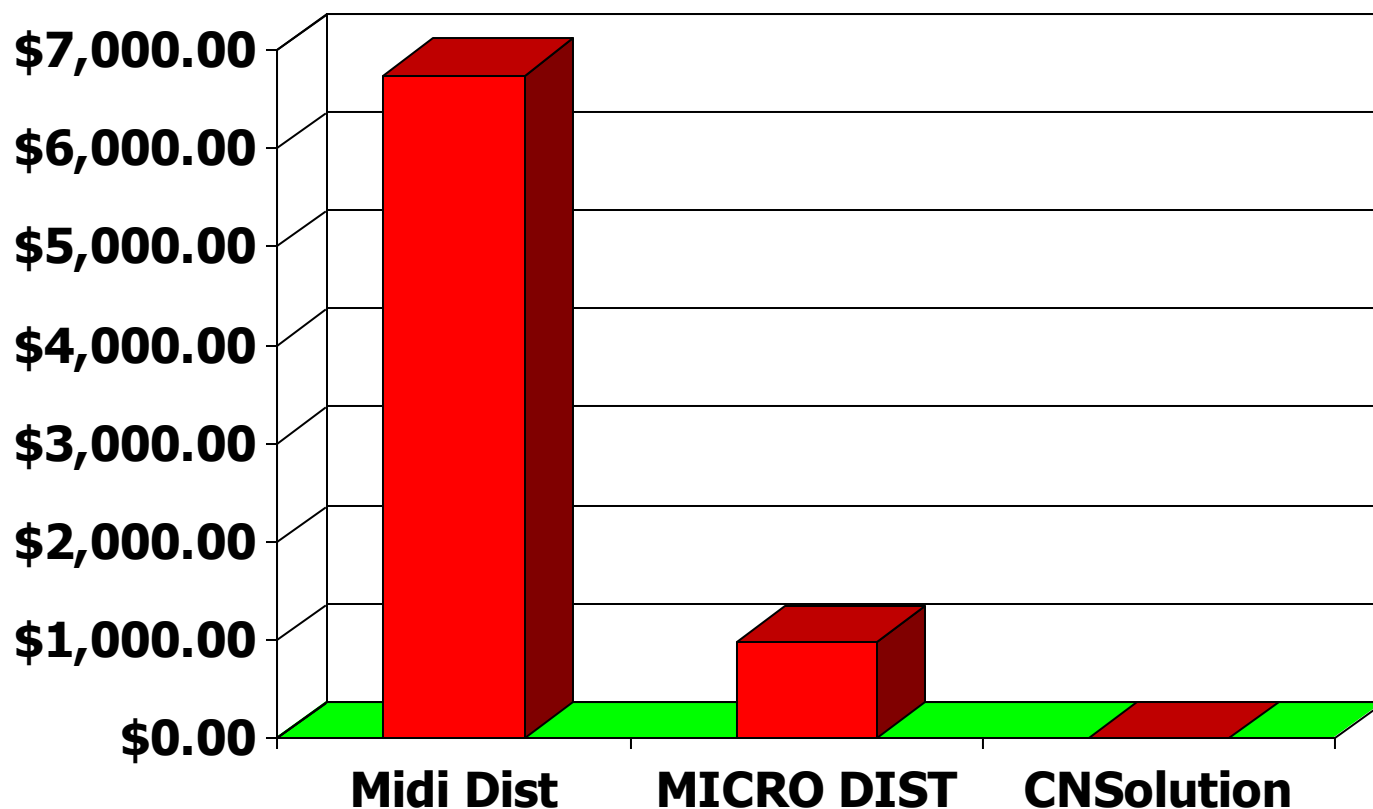


# Estimated Labor (in minutes) Required to Distill 20 Samples for Analysis



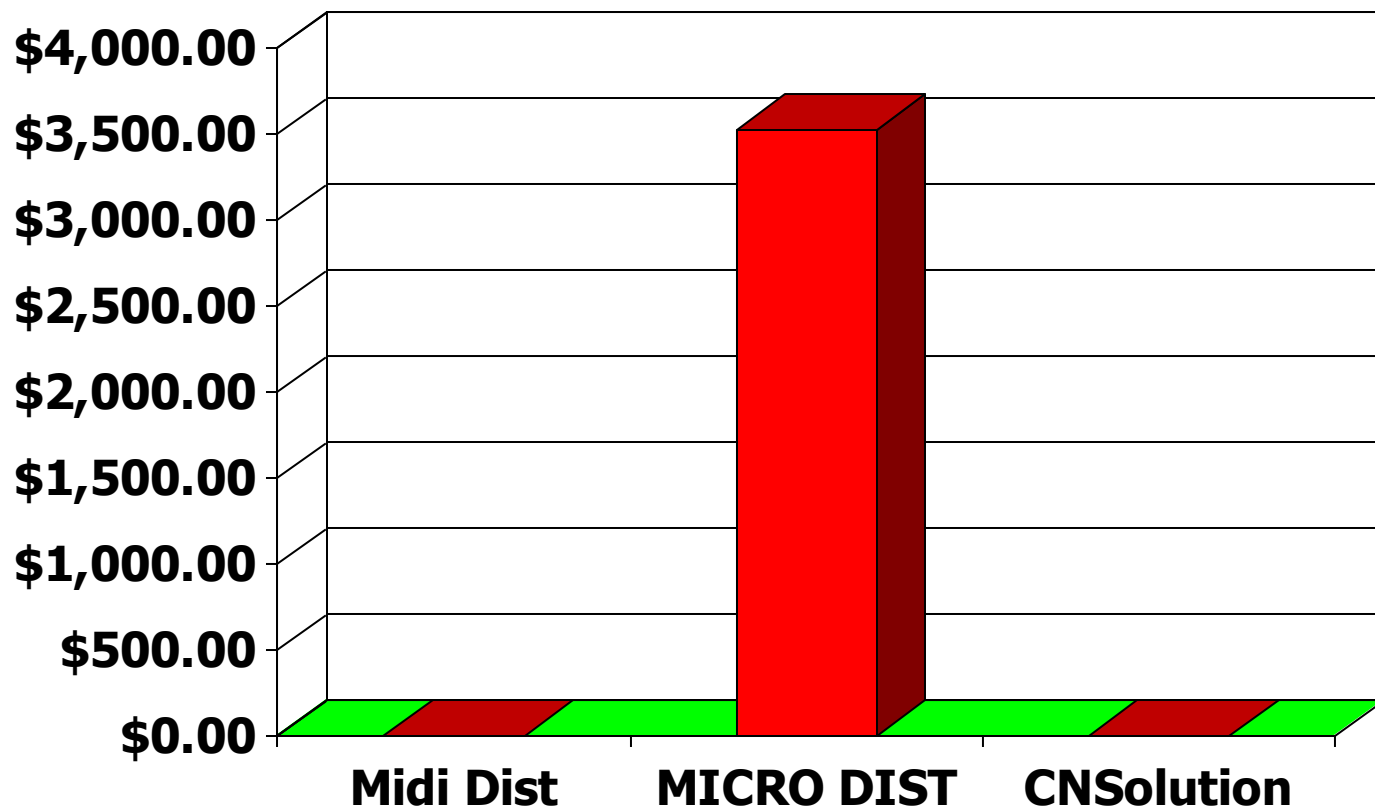


# Estimated Annual Labor Cost to Distill and Analyze 40 Samples per Month (Including Overhead)

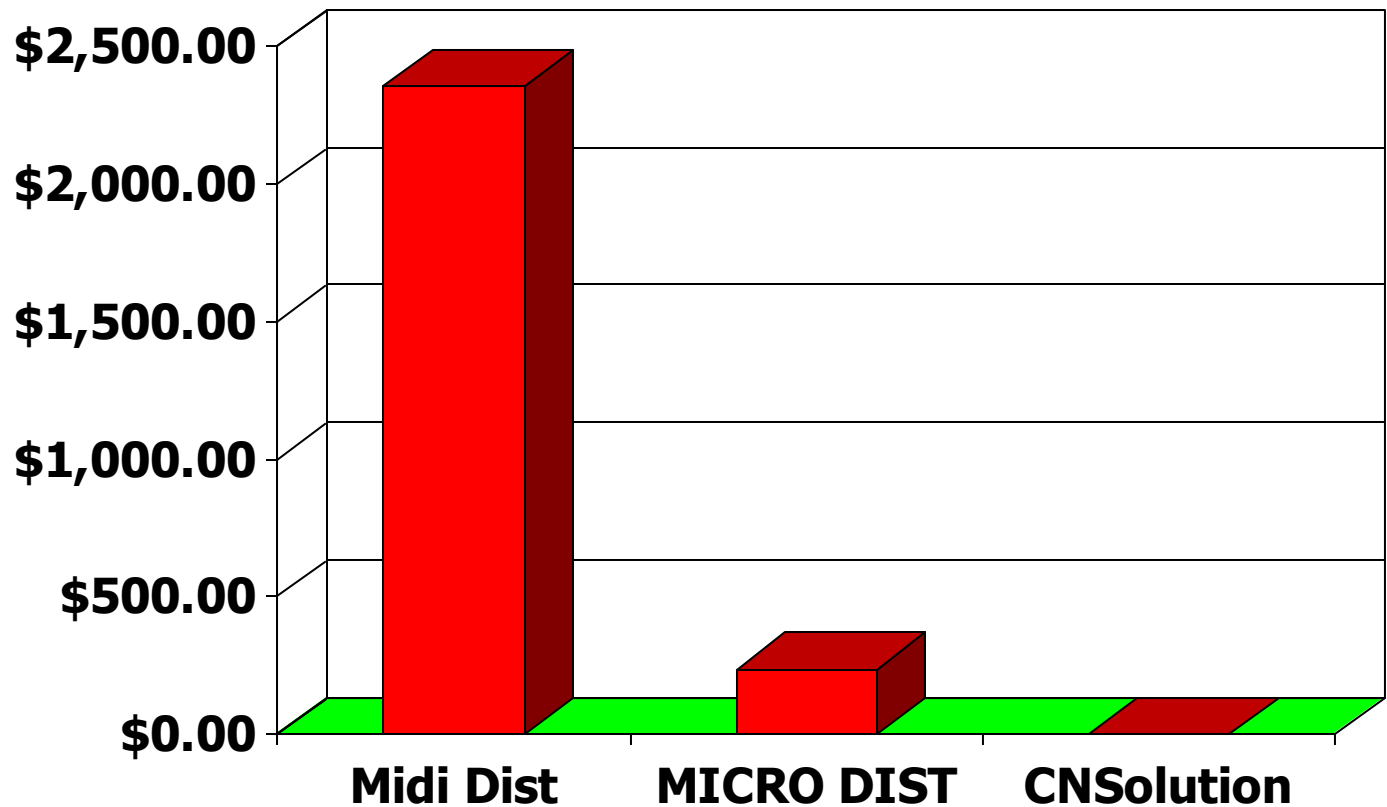


Assume tech pay at  
\$15/hour

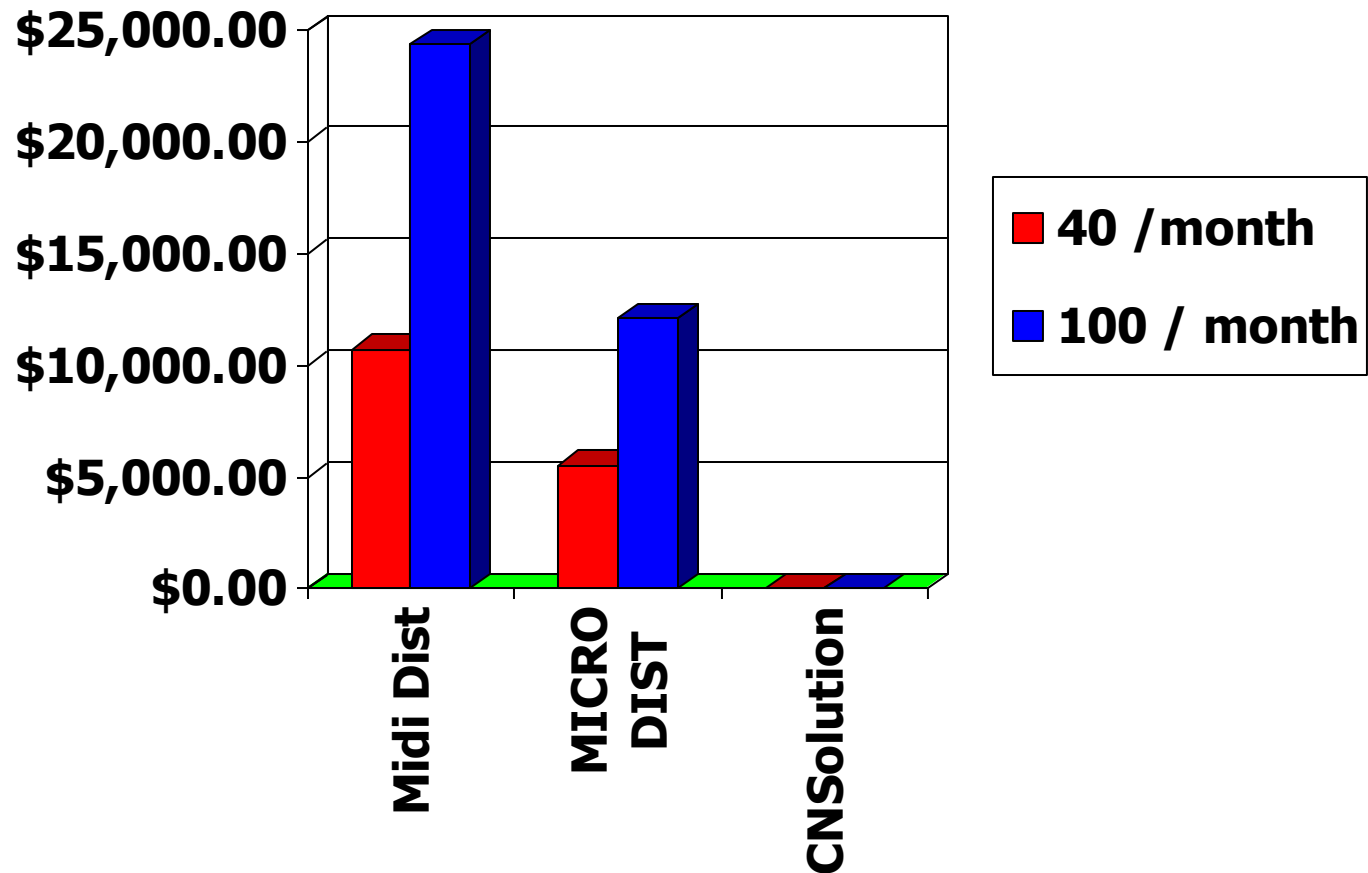
# Estimated Annual Cost for Consumables to Distill and Analyze 40 Samples per Month



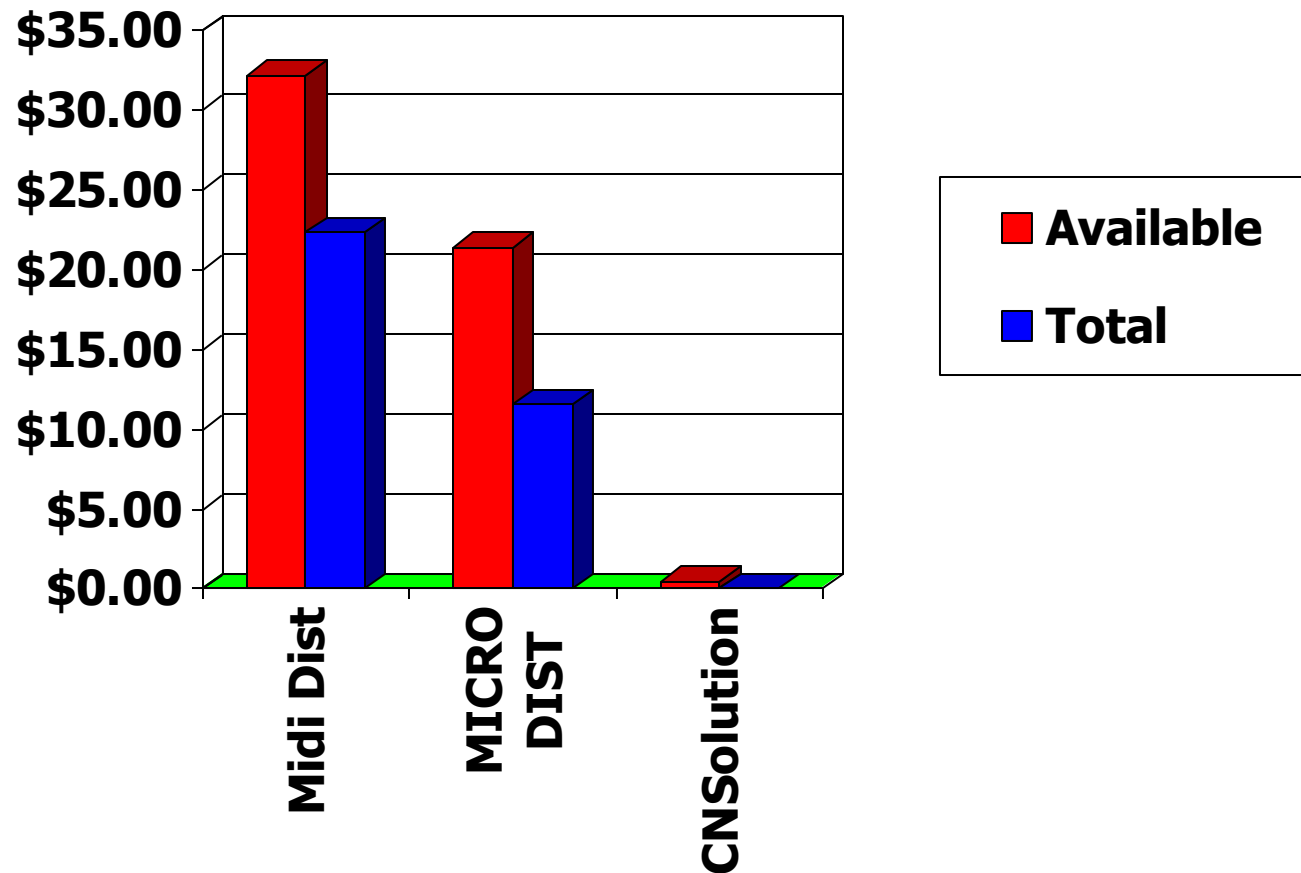
# Estimated Annual Cost for Reagents to Distill and Analyze 40 Samples per Month



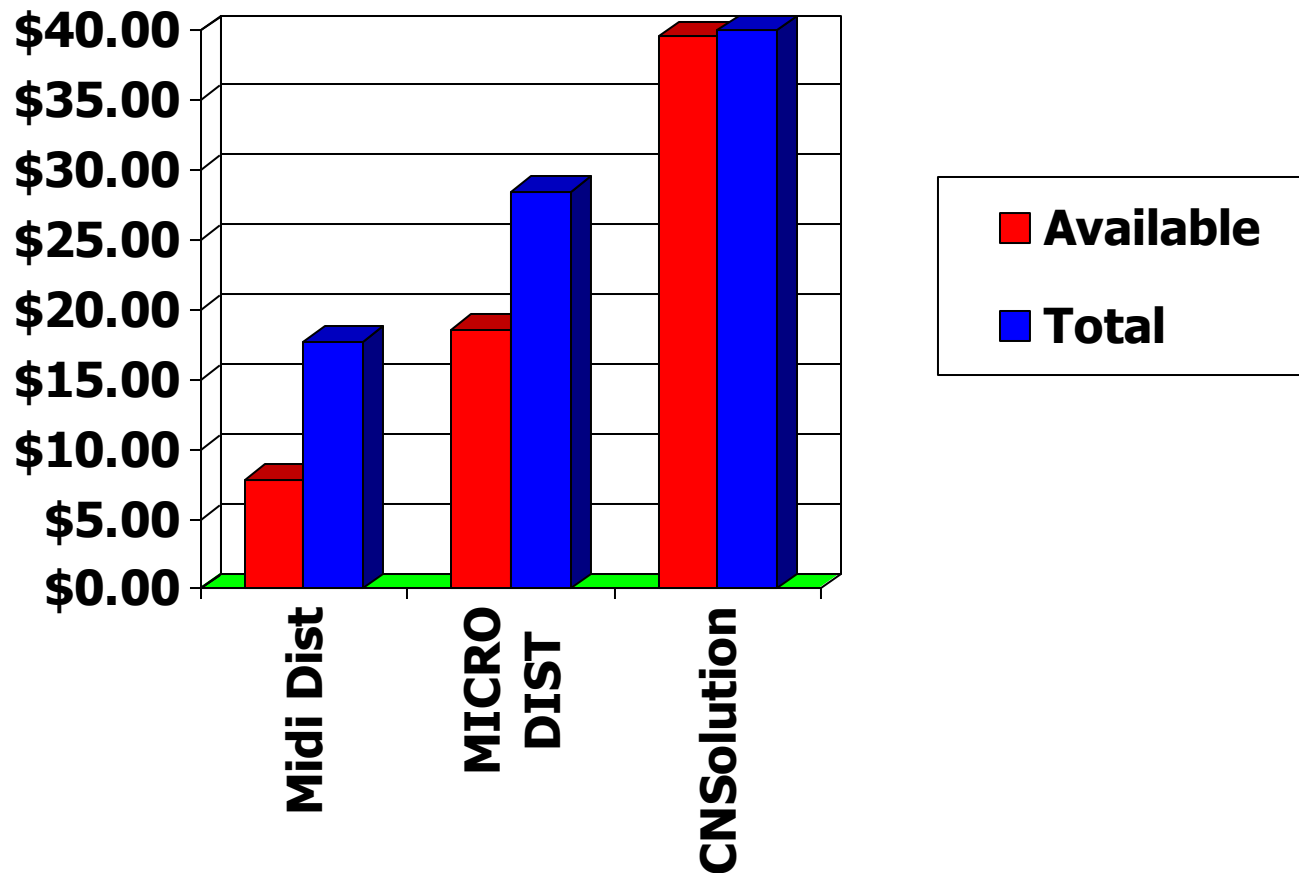
# Annual Cost for Distillation vs. Non-Distillation Techniques



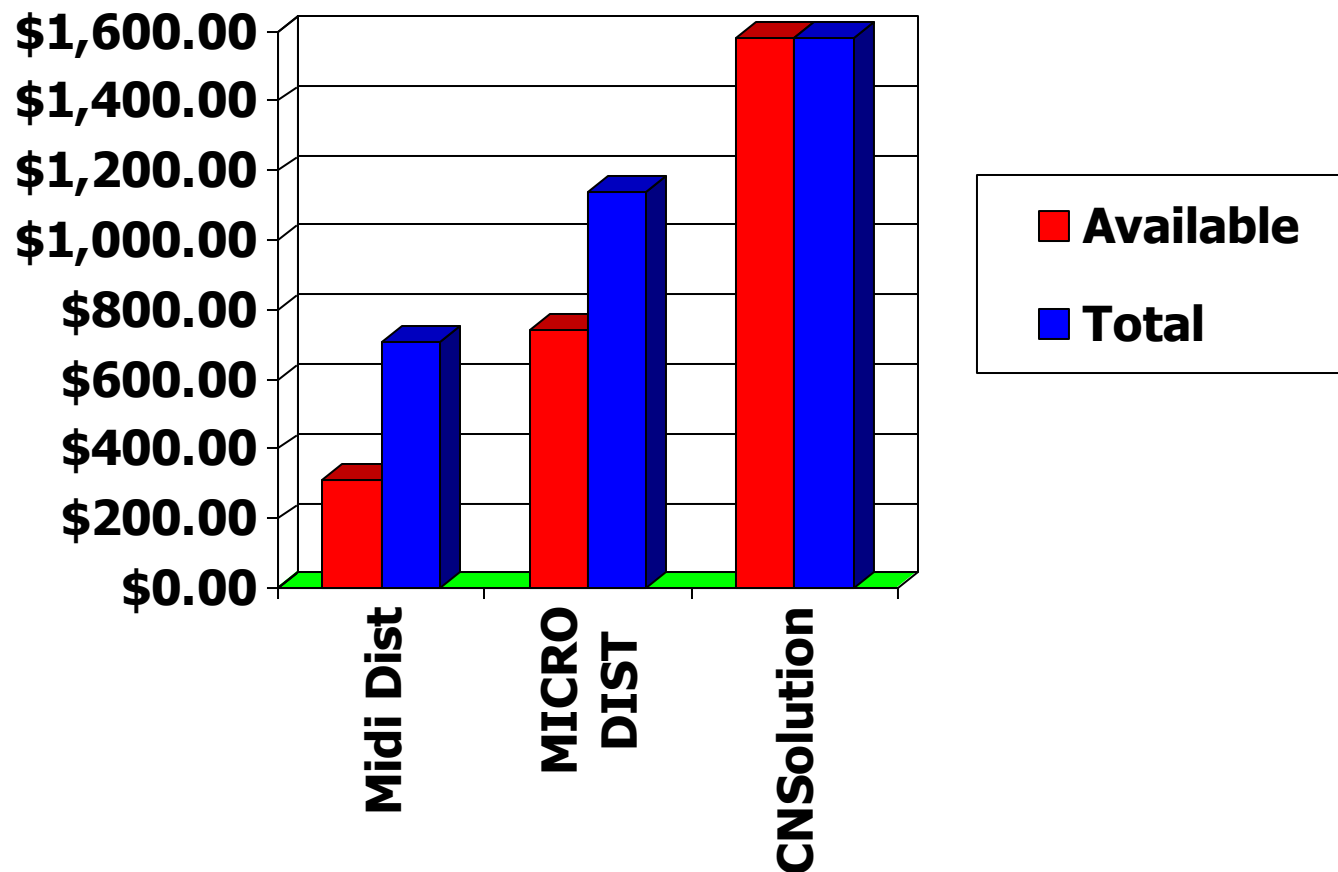
# Comparison of Cost per Sample by Cyanide Species and Method



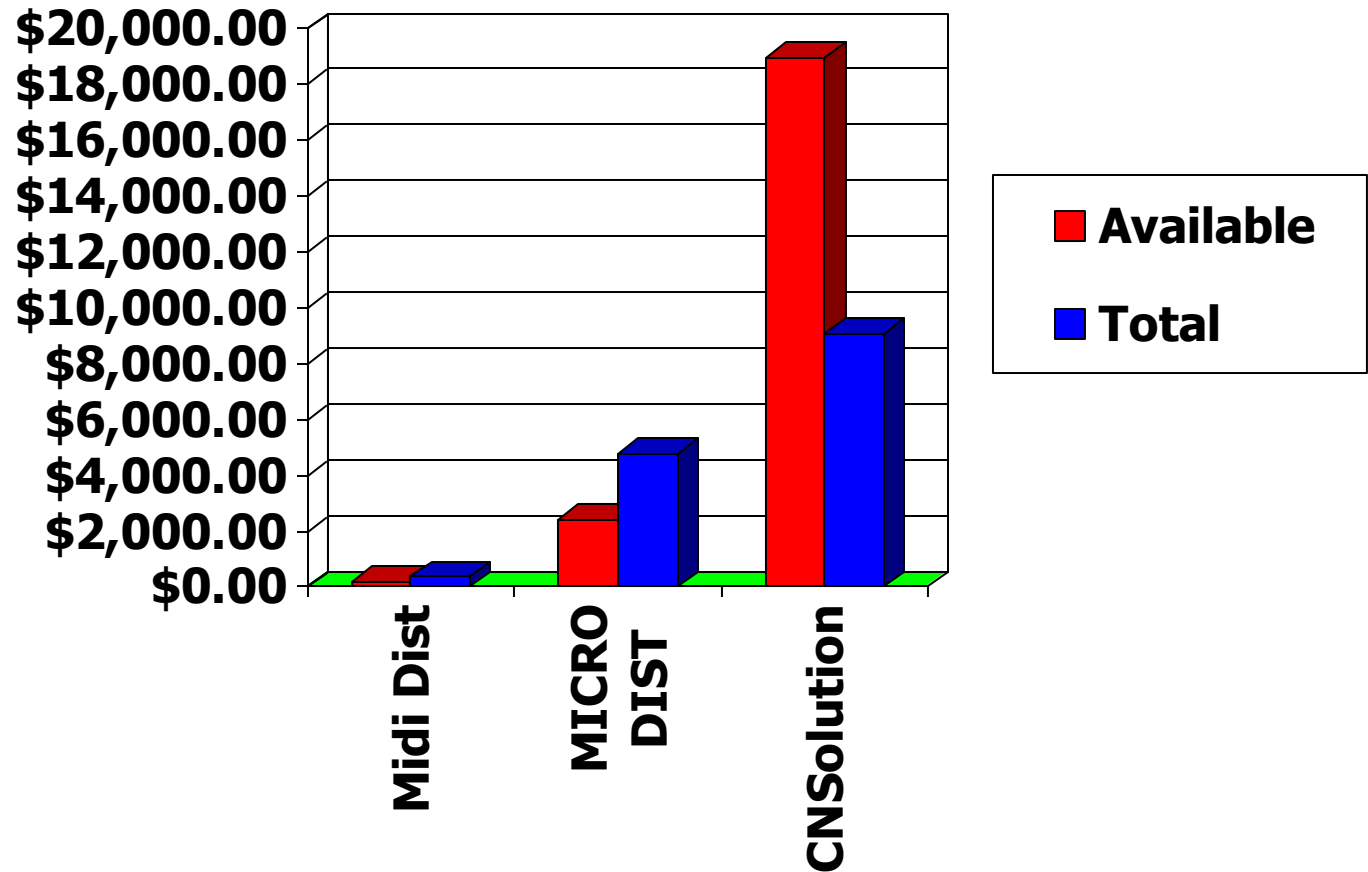
# Profit per Sample by Cyanide Species at \$40 per Test



# Profit per Month by Cyanide Species at \$40 per Test and 40 Samples per Month

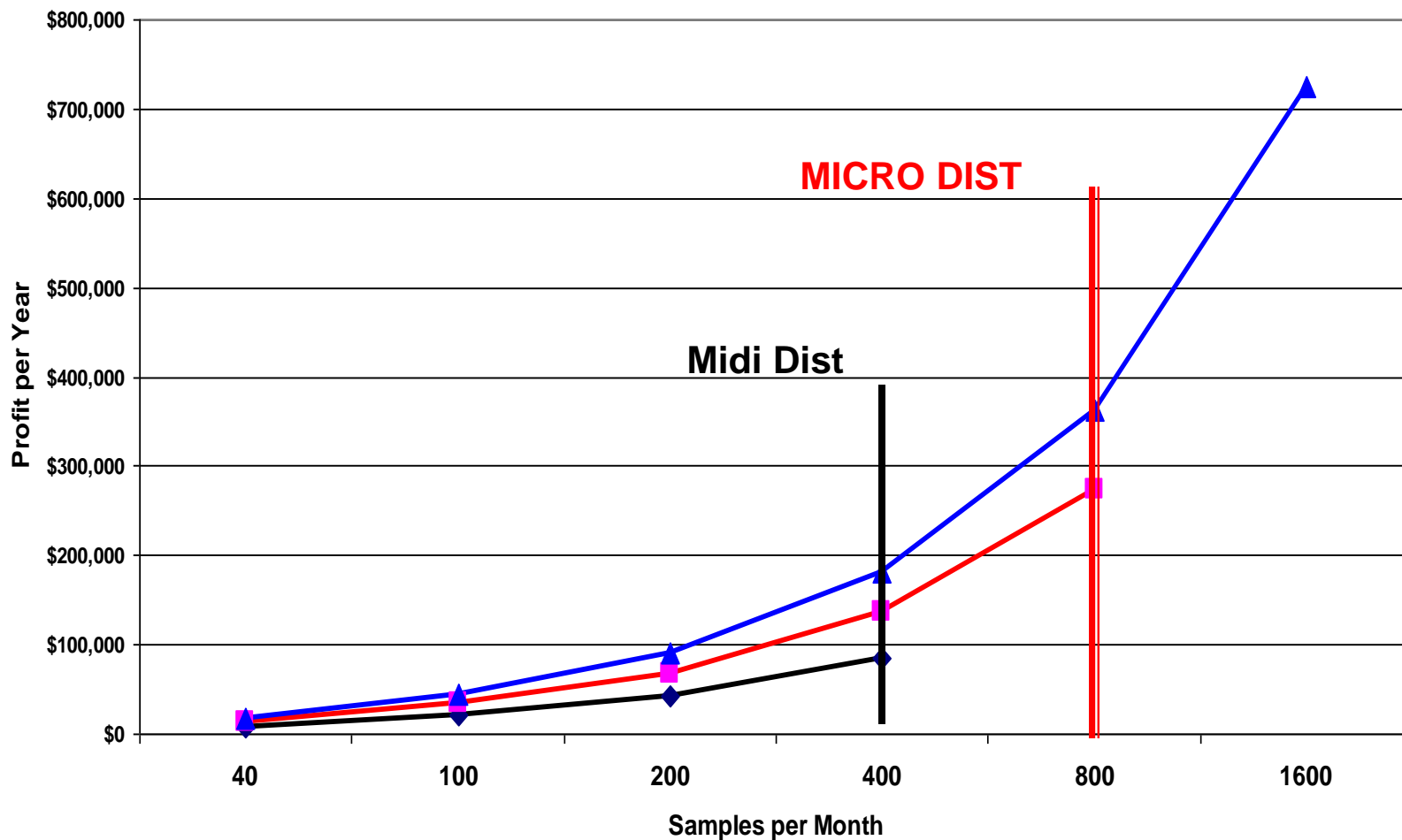


# Profit per Shift by Cyanide Species at \$40 per Test Based on Maximum Capacity





# Estimated Profit by Number of Samples Analyzed at Maximum Capacity

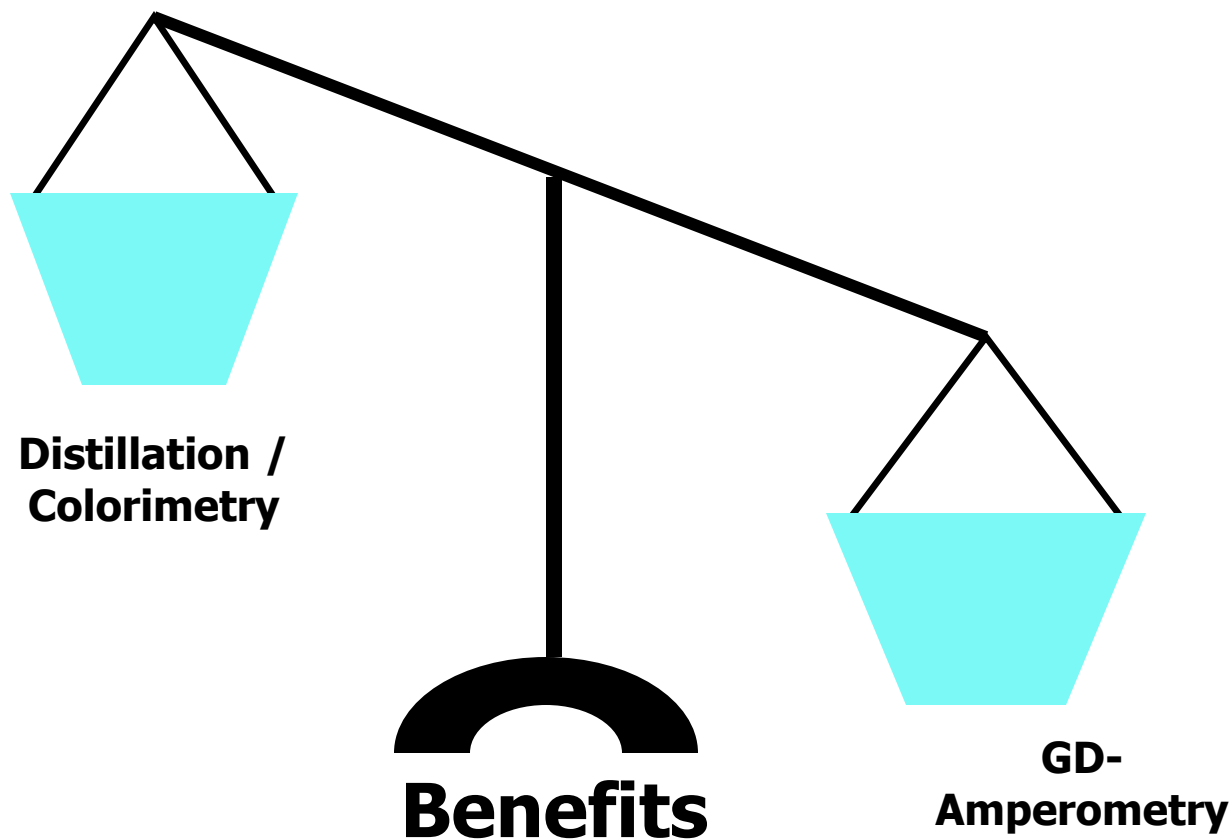


The left side of the slide features a vertical blue bar with a white chromatogram line. Along this line are four boxes containing chemical symbols: CN, NH<sub>3</sub>, PO<sub>4</sub>, and NO<sub>3</sub>.

# Thoughts Regarding “Break Even” Analysis

- **Manual distillation and manual spectrometer versus CNSolution**
  - Manual equipment costs 3 x less
  - CNSolution profit 3 x higher per test
- **Break Even = 1000 samples for both!**
- **That’s 20 samples per week regardless**

In summary, distillation/colorimetry should be replaced





# Thank You

## Questions?

[www.oico.com](http://www.oico.com)



## An Overview and Comparison of Methods for Cyanide Analysis

*Presented at the 2009 Pittsburgh Conference on Analytical Chemistry and Applied Spectroscopy, Chicago, Illinois  
March 8, 2009–March 13, 2009*

### Introduction

Cyanide analysis methods attempt to measure groups of compounds with similar chemical characteristics and report them as a single value. Various techniques are employed to separate specific types of cyanide complexes from each other and potential matrix interferences to achieve accurate quantitation (Figure 1).

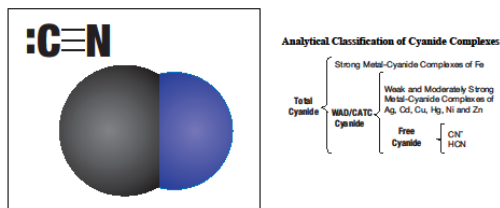


Figure 1. Cyanide refers to a monovalent anion consisting of carbon and nitrogen atoms with triple covalent bonds. Cyanide is very reactive and readily forms metal–cyanide complexes and organic compounds



CN

NH<sub>3</sub>

PO<sub>4</sub>

NO<sub>3</sub>

The regulated community is presented with an array of conflicting definitions of cyanide species, which leads to misunderstandings about what the various techniques and methods actually measure. In fact, the U.S. EPA *Solutions to Analytical Chemistry Problems with Clean Water Act Methods*<sup>(1)</sup>, or "Pumpkin Guide", states, "Next to oil and grease, cyanide is the pollutant for which the most matrix interferences have been reported to EPA."

This study presents an overview and assessment of the most commonly used cyanide analysis methods including cyanide species measured, analytical techniques employed, potential matrix interferences, and final determinative steps.

### Commonly Used Cyanide Analysis Methods

#### *Free Cyanide*

Free cyanide refers to the sum of hydrogen cyanide (HCN) and cyanide ion (CN<sup>-</sup>) in a sample<sup>(2,3)</sup>. Free cyanide is bioavailable and approximately a thousand times more toxic to aquatic organisms than it is to humans<sup>(4)</sup>. Analytically, free cyanide is referred to as the amount of HCN liberated from solution at pH 6.0.

#### *Weak to Moderately Strong Metal - Cyanide Complexes*

Weak to moderately strong metal-cyanide complexes are compounds that dissociate and release hydrogen cyanide gas under mildly acidic conditions (pH 3 to 6). Cyanide species within this category include: simple cyanides-soluble/dissociable alkali metal and alkali earth metal-cyanide complexes (NaCN, KCN, Ca(CN)<sub>2</sub>); weak metal-cyanide complexes (Zn(CN)<sub>2</sub><sup>5</sup>, Cd(CN)<sub>2</sub>); and moderately strong metal-cyanide complexes (Cu(CN)<sub>2</sub><sup>-</sup>, Ni(CN)<sub>2</sub><sup>2-</sup>, Ag(CN)<sub>2</sub><sup>-</sup><sup>(2,3)</sup>. Weak Acid Dissociable (WAD), Cyanide Amenable to Chlorination (CATC), and Ligand Exchange methods were developed to quantify the sum of these cyanide species as well as any free cyanide present in a sample.

Methods intended to measure weak to moderately strong metal-cyanide complexes also measure simple cyanides. Simple cyanides include free cyanide, alkali metal cyanides, alkali earth metal cyanides, and ammonium cyanide.

#### *Strong Metal-Cyanide Complexes*

Strong metal-cyanide complexes are compounds that require strongly acidic conditions (pH < 2) to dissociate and release hydrogen cyanide gas<sup>(2)</sup>. Examples of strong metal-cyanide complexes include Fe(CN)<sub>6</sub><sup>3-</sup>, Fe(CN)<sub>6</sub><sup>4-</sup>, Co(CN)<sub>6</sub><sup>4-</sup>, and Au(CN)<sub>2</sub><sup>-</sup>. The strong acidic conditions used to dissociate these resistant metal-cyanide complexes readily dissociates all other cyanide species present in a sample. "Total Cyanide" is the term used in U.S. EPA methods to refer to the sum of all cyanide species that are converted to hydrogen cyanide following reflux distillation of a sample acid in a strong solution.

For a summary of cyanide analysis methods, refer to Table 1.

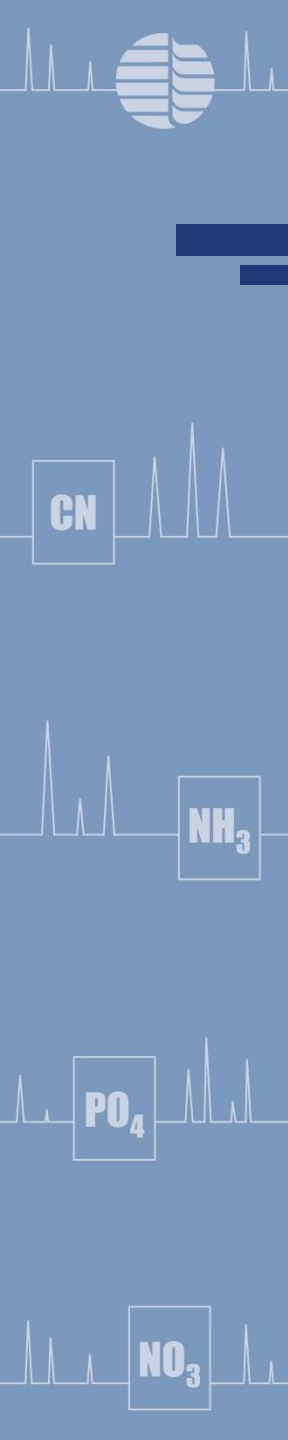


CN

NH<sub>3</sub>PO<sub>4</sub>NO<sub>3</sub>

Table 1: Summary of CN Analysis Methods

Description Name	Method	Potential Interferences	Measurement
Free Cyanide	AOTN D-432	Storage Measurement	Manual observation of cyanide with chloramine T and rapid reaction with (cyano)hydrazine acid. Maximum observation sensitivity by manual observation.
	AOTN D-507	Storage Measurement	Automatic observation of cyanide with chloramine T and rapid reaction with (cyano)hydrazine acid. Maximum observation sensitivity by manual observation.
Acid-Soluble Cyanide (AS-CN)	AOTN D-432	Storage Measurement	Manual observation of cyanide with chloramine T and rapid reaction with (cyano)hydrazine acid. Maximum observation sensitivity by manual observation.
	AOTN D-507	Storage Measurement	Automatic observation of cyanide with chloramine T and rapid reaction with (cyano)hydrazine acid. Maximum observation sensitivity by manual observation.
	AOTN D-507	Storage Measurement	Automatic observation of cyanide with chloramine T and rapid reaction with (cyano)hydrazine acid. Maximum observation sensitivity by manual observation.
	AOTN D-507	Storage Measurement	Automatic observation of cyanide with chloramine T and rapid reaction with (cyano)hydrazine acid. Maximum observation sensitivity by manual observation.
	AOTN D-507	Storage Measurement	Automatic observation of cyanide with chloramine T and rapid reaction with (cyano)hydrazine acid. Maximum observation sensitivity by manual observation.
	AOTN D-507	Storage Measurement	Automatic observation of cyanide with chloramine T and rapid reaction with (cyano)hydrazine acid. Maximum observation sensitivity by manual observation.
Total Cyanide	AS-CN D-432	Storage Measurement	Cyanide in the solution solution is observed with chloramine T.
	AS-CN D-507	Storage Measurement	Cyanide in the solution solution is observed with chloramine T.
	AS-CN D-507	Storage Measurement	Cyanide in the solution solution is observed with chloramine T.
	AS-CN D-507	Storage Measurement	Cyanide in the solution solution is observed with chloramine T.
	AS-CN D-507	Storage Measurement	Cyanide in the solution solution is observed with chloramine T.
	AS-CN D-507	Storage Measurement	Cyanide in the solution solution is observed with chloramine T.
	AS-CN D-507	Storage Measurement	Cyanide in the solution solution is observed with chloramine T.
	AS-CN D-507	Storage Measurement	Cyanide in the solution solution is observed with chloramine T.
	AS-CN D-507	Storage Measurement	Cyanide in the solution solution is observed with chloramine T.
	AS-CN D-507	Storage Measurement	Cyanide in the solution solution is observed with chloramine T.
	AS-CN D-507	Storage Measurement	Cyanide in the solution solution is observed with chloramine T.



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## Potential Method Interferences

### *Sampling, Preservation, and Storage Interferences*

Certain interferences occur during the sampling, preservation, and storage process and apply, to some extent, to all cyanide methods. Chemical preservatives added to samples to preserve the original concentration of cyanide in the sample need to be chosen according to the analytical method that will be used. In general, distillation methods are less forgiving and require extra care in sample pretreatment. Table 2 contains a summary of interferences that result from sample storage and preservation.

### *Sample Processing Interferences*

The sample processing necessary to break metal-cyanide bonds and liberate the cyanide generated from the aqueous sample solution is not interference free. Often, in some sample matrices, the sample processing introduces so much doubt into the measurement that the results cannot be trusted at all. Table 2 contains a summary of interferences that can be introduced during sample processing.

### *Measurement Interferences*

The final measurement technique used to determine the cyanide concentration is also not interference free. Since the final measurement is performed under the assumption that all interferences were previously removed, these interferences often go undetected. This is especially true for automated colorimetric methods. Table 2 contains a summary of interferences that may occur during the final cyanide measurement step.



CN

NH<sub>3</sub>PO<sub>4</sub>NO<sub>3</sub>

Table 2: Summary of CN Method Interferences

Analysis Step	Compound	Process/Measurement Technique	Description of Interference
Sampling, Preservation, and Storage	Residual chlorine, peroxide, or other oxidizers	N/A	React with cyanide in solution rapidly decreasing the cyanide concentration. Oxidizers can co-exist with cyanide.
	Chloramines	N/A	React with sample at pH>10 increasing the cyanide concentration.
	Sulfide	N/A	Reacts with cyanide to form thiocyanate decreasing the cyanide concentration. Reaction is especially rapid if metal sulfides, such as lead sulfide, are present. The reaction is fairly slow without metal sulfides.
	Native Sulfur (colloidal sulfur)	N/A	Reacts with cyanide to form thiocyanate decreasing the cyanide concentration. Reaction is very fast with colloidal sulfur.
	Sulfite	N/A	Reacts with strong cyanide complexes at pH>10 decreasing the cyanide concentration. The reaction is almost immediate at pH>12.
	Light (<350 nm)	N/A	Reacts with strong metal-cyanide complexes releasing free cyanide.
	Ascorbic acid	N/A	Reacts with cyanide decreasing the cyanide concentration. Sample holding time when ascorbic acid is added is less than 48 hours. In some samples, ascorbic acid can react with ammonia or other nitrogen sources and increase the cyanide concentration.
	Formaldehyde	N/A	Reacts with cyanide decreasing the concentration. In some samples, formaldehyde reacts with ammonia or other nitrogen sources and increases the cyanide concentrations.
Sample Processing	Oxidizers	Distillation	Reacts with cyanide decreasing its concentration.
	Sulfide	Distillation	Distills into absorber solution and reacts with CN forming thiocyanate.
		Gas diffusion	Passes through diffusion membrane.
	Sulfite (or Sulfur Dioxide)	Distillation	Reacts with cyanide decreasing its concentration. Distills into absorber solution and reacts with cyanide decreasing its concentration.
	Thiosulfate and other oxidized sulfur species (except sulfate)	Distillation	Decompose to form native sulfur and sulfur dioxide. React with cyanide decreasing its concentration. Sulfur Dioxide distills into absorber solution and reacts with cyanide decreasing its concentration.
	Thiocyanate	Cyanide Amenable to Chlorination (C/ATC)	Reacts with chlorine during alkaline chlorination and generates cyanide. Causes negative C/ATC results.
		Distillation	Decomposes to sulfur dioxide and reacts with cyanide decreasing its concentration. Sulfur dioxide distills into absorber solution.
		UV Irradiation	Can react at < 280 nm to form cyanide
	Thiocyanate + Nitrate or Nitrite	Distillation	Decompose to form cyanide
	Misc. Organics + Nitrate or Nitrite	Distillation	Decompose to form cyanide
Measurement	Sulfide	Distillation	Excessive foaming and possible violent release of carbon dioxide
		Gas diffusion	Passes through diffusion membrane
		Titration	Detected as cyanide
		Ion Selective Electrode	Detected as cyanide
	Thiocyanate	Colorimetry	Detected as cyanide (at > 10-mg S/L)
		GD-Amperometry	Detected as cyanide (at > 50-mg S/L)
		Colorimetry	Detected as cyanide
	Fatty Acids	Titration	Mask end point
	Sulfur Dioxide	Colorimetry	Increase chloramine-T demand resulting in a negative bias. The increased demand may not be noticed with automated methods.
	Carbonates	GD-Amperometry	Negative bias at > 1,500-mg CO <sub>3</sub> /L



CN

NH<sub>3</sub>

PO<sub>4</sub>

NO<sub>3</sub>



CN

NH<sub>3</sub>

PO<sub>4</sub>

NO<sub>3</sub>



CN

NH<sub>3</sub>

PO<sub>4</sub>

NO<sub>3</sub>